

TRIPTYCENE ANALOGS

CROSS-REFERENCE TO RELATED APPLICATIONS

This application claims priority to United States provisional application serial number 60/238856, filed on October 6, 2000, which is hereby incorporated by reference to the extent not inconsistent with the disclosure herewith.

BACKGROUND OF THE INVENTION

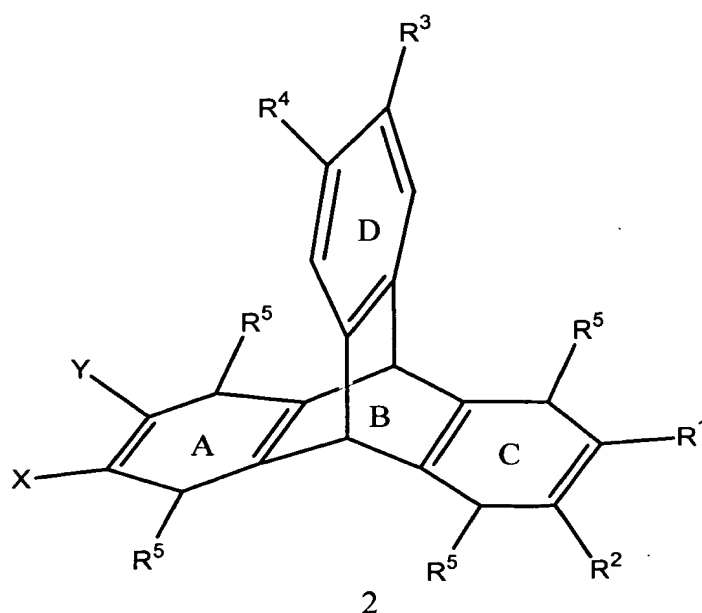
As precursors of reactive quinone methides, many natural and synthetic quinones function as bioreductive alkylating agents and have antitumor activity. (See for example, Thomson RH, Naturally occurring quinones III: recent advances. New York: Chapman and Hall 1987; Moore HW, Science 1977, 197: 527-32; Lin AJ, et al, J Med Chem 1972, 15: 1247-52; Lin AJ, et al, J Med Chem 1973, 16: 1268-71; Lin AJ, et al, J Med Chem 1974, 17: 558-61; Lin AJ, et al, J Med Chem 1975, 18: 917-21). The cytotoxicity of quinones may be due to two competing mechanisms: soft electrophilic arylation and redox cycling oxidation. (See for example, Brunmark A, Cadenas E, Free Radical Biol Med 1989, 7: 435-77; O'Brien PJ, Chem-Biol Interact 1991, 80: 1-41; Monks TJ, et al, Toxicol Appl Pharmacol 1992, 112: 2-16). While complete two-electron reduction of the quinone ring by DT diaphorase produces a stable hydroquinone, partial one-electron reduction of the quinone ring by NADPH-oxidizing enzymes yields an unstable semiquinone free radical (FR) that can spontaneously autoxidize at the expense of molecular O₂ to generate a cascade of reactive O₂ species (ROS) and FRs, which can induce DNA damage, lipid peroxidation and cytotoxicity. However, most quinone antitumor agents used clinically, such as anthracycline antibiotics, mitomycin C and benzoquinone derivatives, have a complex chemical structure with a number of active functional groups and the exact contribution of the quinone group to their antitumor activity remains uncertain. (See for example, Myers CE, Chabner BA, Anthracyclines. In: Chabner BA, Collins JM, eds. Cancer chemotherapy: principles and practice. Philadelphia: Lippincott 1990: 356-81; Rossi L, et al, Arch Biochem Biophys 1986, 251: 25-35; Begleiter A, et al, Cancer Res 1988, 48: 1727-35; Qiu XB, et al, Free Radical Biol Med 1998, 24: 848-54). The anthracycline quinone antibiotics adriamycin (ADR) and daunomycin (DAU) covalently bind to and intercalate into DNA, inhibit

DNA replication and RNA transcription, are DNA topoisomerase (Topo) II poisons, produce oxidative stress and damage biomembranes, induce DNA breakage and chromosomal aberrations, trigger apoptosis and have a wide spectrum of anticancer activity. (See for example, Cadenas E, Free Radical Biol Med 1998, 24: 848-54; Liu LF, Annu Rev Biochem 1989, 58: 351-75; Mimnaugh EG, et al, Cancer Res 1985, 45: 3296-304; Ganapathi R, et al, Biochem Pharmacol 1990, 40: 1657-62; Ling Y-H, et al, Cancer Res 1993, 53: 1845-52; Ramachandran C, et al, Anticancer Res 1997, 17: 3369-76). However, the clinical effectiveness of DOX and DAU is severely limited by their cumulative cardiotoxicity and ability to induce multi-drug resistance, so it is important to develop drugs with improved bioactivity.

SUMMARY OF THE INVENTION

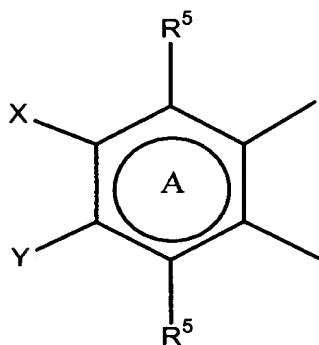
This invention provides analogs of triptycene which are useful as anticancer drugs, as well as for other uses. The potency of these compounds is in a similar magnitude as daunomycin, a currently used anticancer drug. Each compound of the invention produces one or more desired effects (blocking nucleoside transport, inhibiting nucleic acid or protein syntheses, decreasing the proliferation and viability of cancer cells, inducing DNA fragmentation or retaining their effectiveness against multidrug-resistant tumor cells).

More specifically, the invention provides triptycene analogs having the following formula:

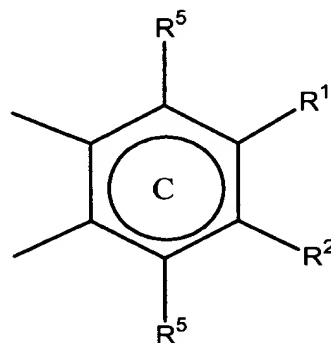


wherein X is selected from the group consisting of: H, R, SR and NR_2 ;

Y is selected from the group consisting of: halogen (preferably Cl, Br, I), R, NR_2 , SR and H;
 R and R^{1-2} are independently selected from the group consisting of: H, halogen, OR, and hydrocarbyl (preferably lower alkyl, allyl, phenyl, aryl, substituted alkyl, substituted allyl, substituted phenyl, $-\text{CH}_2-(\text{CH}_2)_n\text{CO}_2\text{H}$, $-\text{CH}_2-(\text{CH}_2)_n\text{CH}(\text{NH}_2)\text{CO}_2\text{H}$, carboxylic acid, substituted carboxylic acid, amine, substituted amine, NHR , NR_2 , amino acid, $\text{RCO}_2(\text{CH}_2)_n\text{NH}$, where one or both of the hydrogen atoms on CH_2 can be substituted with alkyl, allyl, phenyl, aryl, substituted allyl, substituted phenyl, substituted carboxylic acid, amine, or substituted amine, and where n is an integer from 0 to 8); R^{3-4} , independently of one another, are selected from the group consisting of: H, halogen (preferably bromine), OR, R, SR and NR_2 ; R^5 , independently of other R^5 s, is selected from the group consisting of: $=\text{O}$, $=\text{N-OH}$, and $=\text{CHR}$; and reduced forms thereof; wherein in reduced forms, either ring A or ring C or both is replaced with

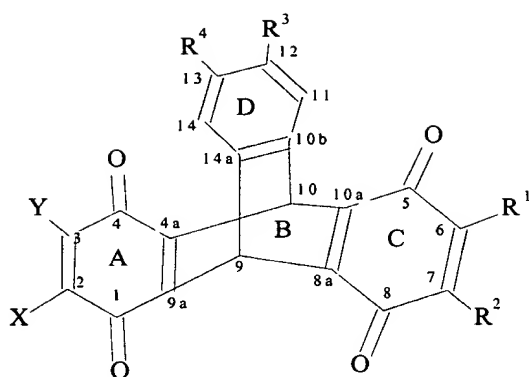


or

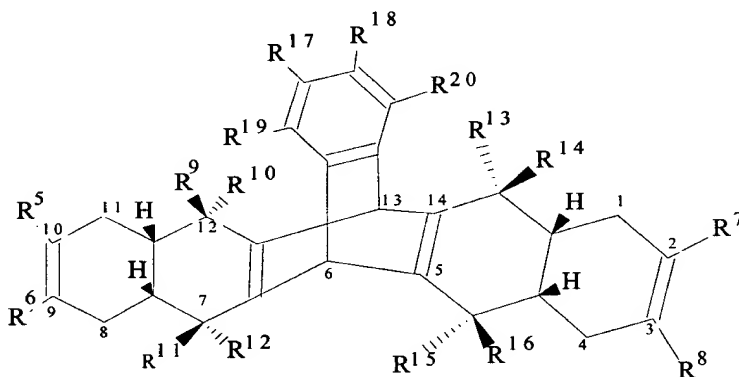


and wherein in reduced forms, each R^5 is independently H, C1-C8 alkyl or -OR;
 and pharmaceutically acceptable salts of the foregoing, as well as optical isomers thereof.

The numbering scheme used herein is shown in the example structure below:



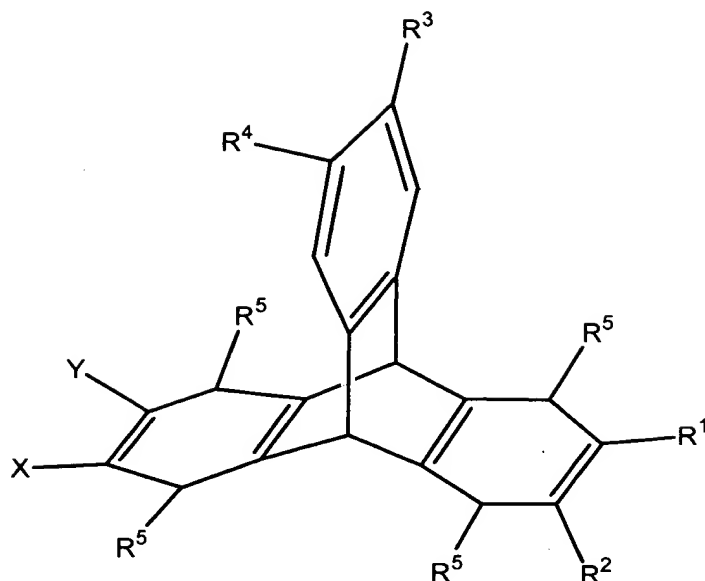
Other compounds of the invention include those with formula:



wherein

R^5 is selected from the group consisting of: R, halogen, NR_2 , SR and H; R^6 is selected from the group consisting of: H, R, SR and NR_2 ; R^7 and R^8 are independently selected from the group consisting of: H, halogen, OR and hydrocarbyl; R^{17} and R^{18} are independently selected from the group consisting of: H, halogen, (preferably bromine), R, SR and NR_2 ; R^{19} and R^{20} are, independently of one another, H, R, or OR; (R^9 and R^{10}) and (R^{11} and R^{12}) and (R^{13} and R^{14}) and (R^{15} and R^{16}) are together =O or are independently H or -OR; R is selected from the group consisting of H, halogen, OR and hydrocarbyl; reduced forms thereof and pharmaceutically acceptable salts of the foregoing, as well as optical isomers thereof.

Other compounds of the invention include those with formula:



wherein X is $-NW(CW_2)_nZ$, where the Ws are independently selected from the group consisting of: H, C1-C8 alkyl, and C1-C8 alkenyl and Z is selected from the group consisting of: R, COR, COOR, CONR₂, OOCR and NRCOR;

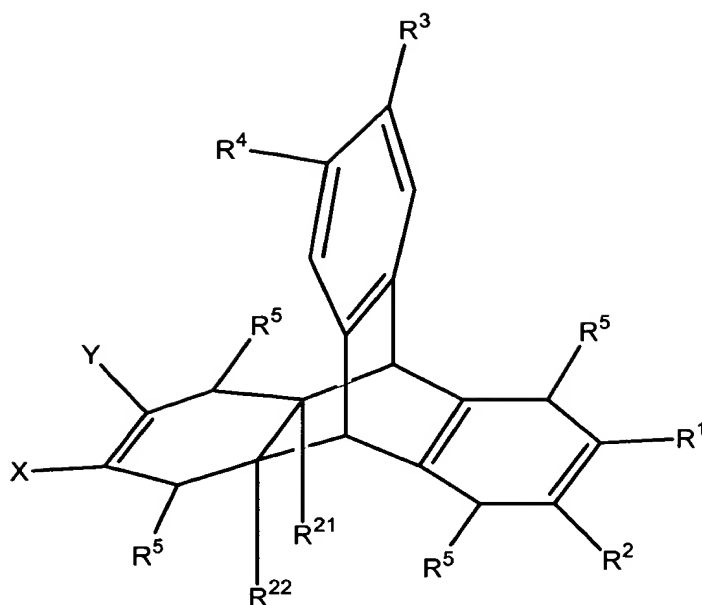
Y is selected from the group consisting of: halogen, C1-C8 alkyl, C1-C8 alkenyl, OR, NR₂, SR, H, COR, OCOR and NRCOR;

R and R¹⁻², are independently selected from the group consisting of: H, OR, and hydrocarbonyl;

R³⁻⁴, independently of one another, are selected from the group consisting of: H, OR, SR, and NR₂;

R⁵, independently of other R⁵s, is selected from the group consisting of: =O, -H and -OT, where T is H or C1-C8 alkyl or alkenyl; and pharmaceutically acceptable salts of the foregoing, as well as optical isomers thereof.

Other compounds of the invention include those with formula:



wherein X is selected from the group consisting of: H, R, SR and NR_2 ;

Y is selected from the group consisting of: halogen, NR_2 , SR, H, and R;

R and R^{1-2} , are independently selected from the group consisting of: H, halogen, OR, and hydrocarbyl;

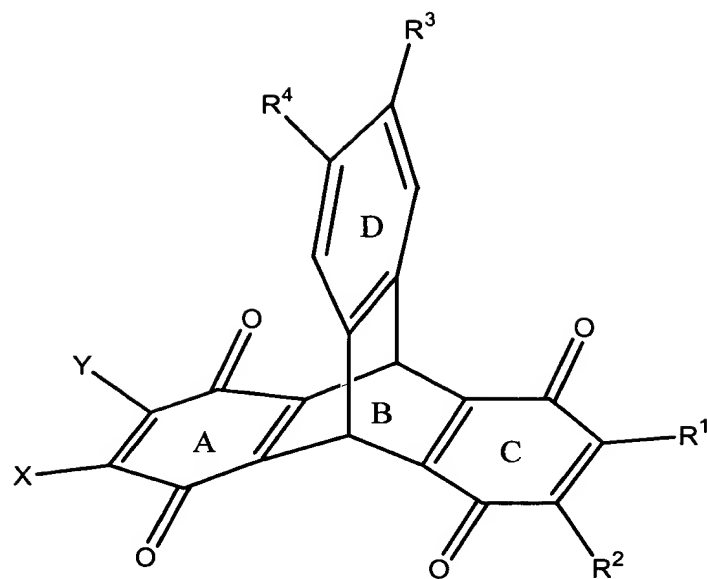
R^{3-4} , independently of one another, are selected from the group consisting of: H, halogen (preferably bromine), R, SR, and NR_2 ;

R^5 , independently of other R^5 s, is selected from the group consisting of: $=\text{O}$, $=\text{NOH}$, $=\text{CHR}$ and reduced forms thereof;

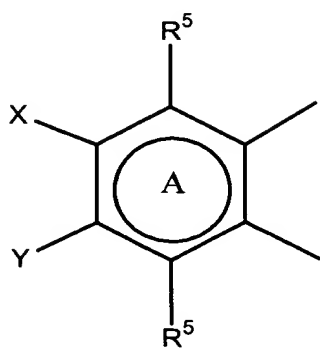
R^{21} and R^{22} are independently selected from the group consisting of: H, R, and OR;

and reduced forms thereof and pharmaceutically acceptable salts of the foregoing, as well as optical isomers thereof.

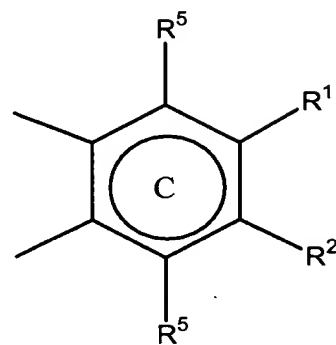
Also provided are compounds of the formula:



and the reduced forms thereof, wherein in said reduced forms, either ring A or ring C or both is reduced to



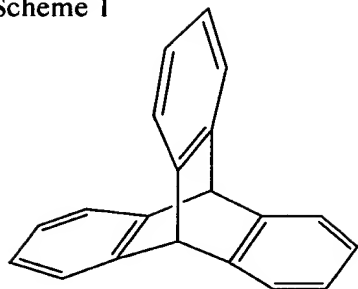
or



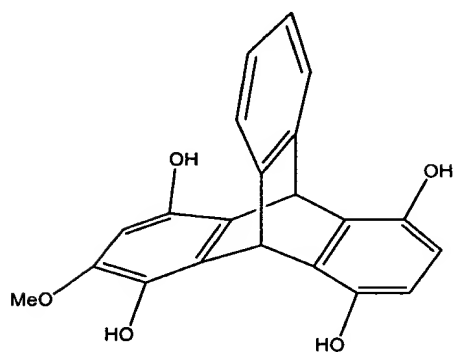
wherein all but one of X, Y, R1 and R2 is independently H, C1-C6 alkyl, C1-C6 alkenyl, OR, SR or NR2 wherein each R is independently H or C1-C6 alkyl and the other R1 or R2 is a solubilizing group; and each R5 is independently H, C1-C6 alkyl or OR. The solubilizing group may be of the formula: $\text{NR}(\text{CR}_2)_n\text{X}$ wherein X is a sugar, R, COR, COOR, CONR_2 , OOCR and NRCOR; R is independently selected from the group consisting of: H, C1-C8 alkyl and C1-C8 alkenyl; n is an integer from 1 to 8.

Scheme 1 shows some of the compounds of the invention and abbreviations used herein.

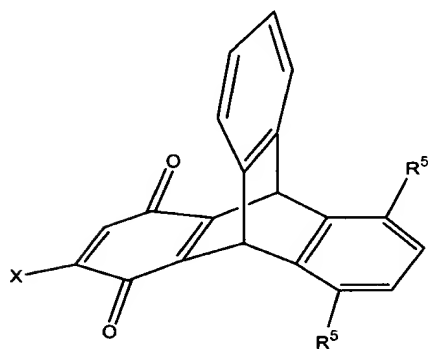
Scheme 1



TT0: triptycene



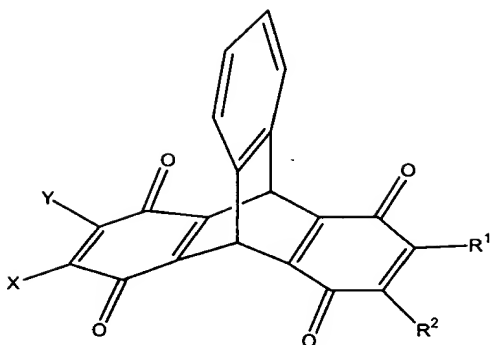
TT3



TT5: $X = R^5 = \text{OMe}$

TT7: $X = R^5 = \text{H}$

TT9: $X = \text{OMe}, R^5 = \text{H}$



TT1: $X = Y = \text{H}$

TT2: $X = \text{OMe}, Y = \text{H}$

TT13: $X = \text{OMe}, Y = \text{Br}$

TT14: $X = \text{NHMe}, Y = \text{Br}$

TT15: $X = \text{MHNe}, Y = \text{OH}$

TT16: $X = \text{OMe}, Y = \text{Br}, R^2 = \text{NMe}_2$

TT17: $X = \text{NHCH}_2\text{CH}_2\text{CO}_2\text{H}, Y = \text{Br}$

TT18: $X = \text{NHCH}_2\text{CH}_2\text{CO}_2\text{Et}, Y = \text{OH}$

TT19: $X = \text{NH}(\text{CH}_2)_3\text{CH}_2(\text{NH}_2)\text{CHCO}_2\text{H}, Y = \text{Br}$

TT20: $X = \text{NH}(\text{CH}_2)_3\text{CH}_2(\text{NH}_2)\text{CHCO}_2\text{H}, Y = \text{OH}$

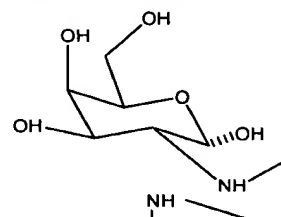
TT21: $X = \text{NHCH}_2\text{CH}_2\text{CO}_2\text{H}, Y = \text{OH}$

TT24A: $X = \text{OMe}, Y = \text{Br}, R^2 = \text{NHMe}$

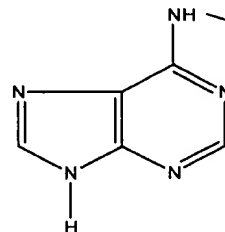
TT24B: $X = \text{OMe}, Y = \text{Br}, R^1 = \text{NHMe}$

N analog 1: $X = \text{NH}(\text{CH}_2)_3\text{CH}_2(\text{NH}_2)\text{CHCO}_2\text{H}, Y = \text{Br}$

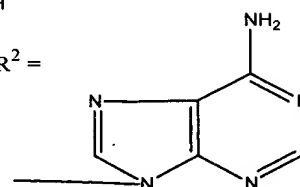
N analog 2: $Y = \text{Br}, X =$



N analog 3: $Y = \text{Br}, X =$

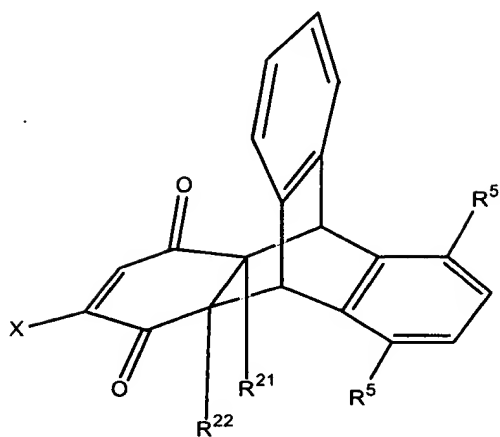


N analog 4: $X = \text{OMe}, Y = \text{Br}, R^2 =$



S analog 9: $Y = \text{Br}, X = \text{SCH}_2\text{CH}(\text{CO}_2\text{H})\text{NH}_2$

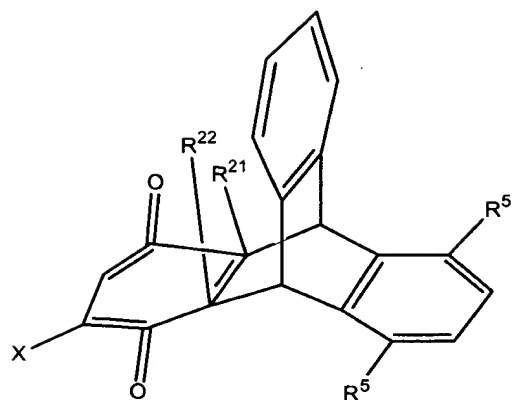
Scheme 1 (continued)



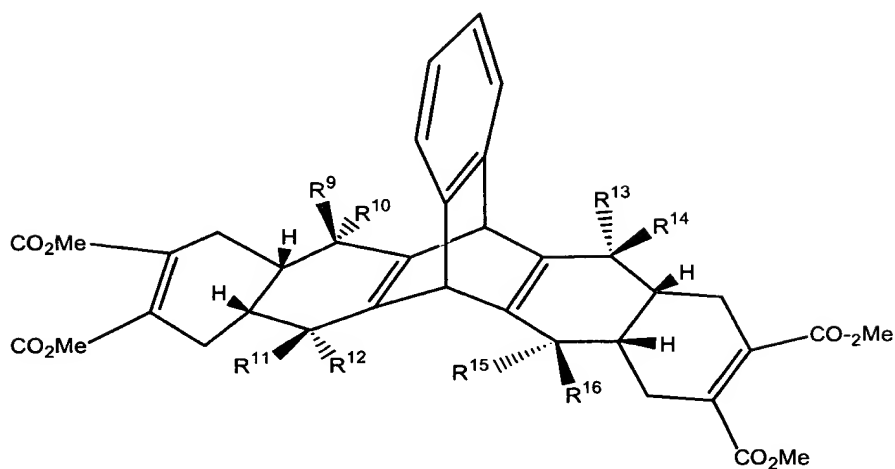
TT6: $X = R^5 = R^{21} = R^{22} = H$

TT8: $X = OMe, R^5 = R^{21} = R^{22} = H$

TT10: $X = CO_2Me, R^5 = OMe, R^{21} = R^{22} = H$



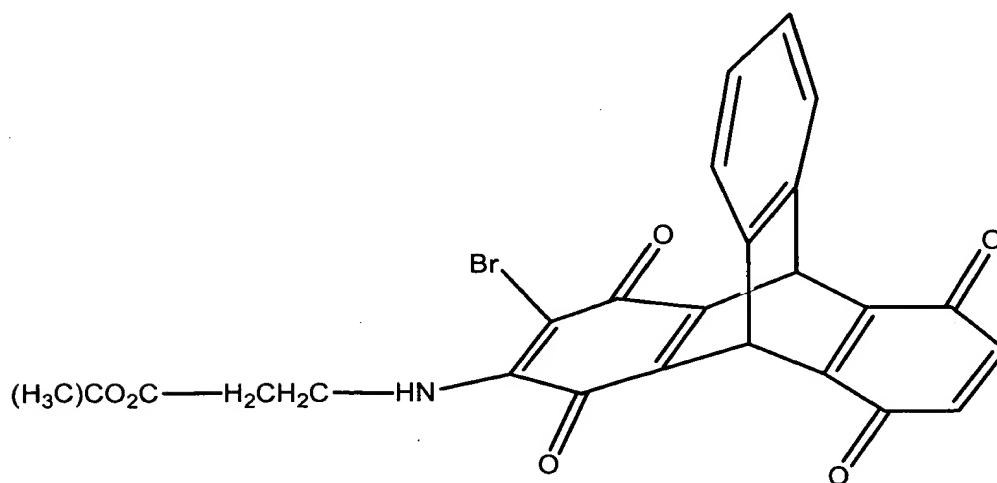
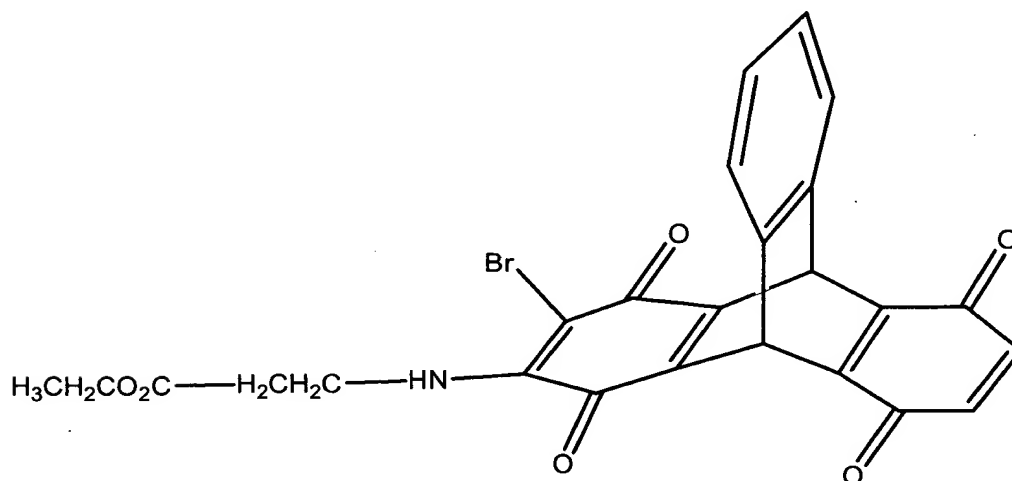
TT11: $X = H, R^5 = OMe, R^{22} = H, R^{21} = CO_2Me$



TT4: $R^9 = R^{10} = R^{11} = R^{12} = R^{13} = R^{14} = R^{15} = R^{16} = O$

TT12: $R^9 = R^{11} = R^{13} = R^{15} = H, R^{10} = R^{12} = R^{14} = R^{16} = OH$

Currently, the most preferred compounds include those compounds of the formulas listed above wherein at least one of X, Y, R¹ and R² is selected from the group consisting of: a nitrogen containing group, a water soluble group, and a sulfur containing group, and the following:



The compounds listed above are useful in treating malaria, cancer, as well as other diseases. Other preferred compounds include those specifically depicted and described in this disclosure.

5 A class of compounds of this invention includes those compounds listed above as presently preferred. Another class of compounds of this invention include TT2 and TT13. Another class of compounds of this invention include TT1, TT7 and TT9. Another class of compounds of this invention include one or members of the class TT3, TT5, TT6, TT8 and TT10. Another class of compounds of this invention includes homologs of the foregoing
10 compounds. One class of compounds is the compounds TT1-13. Another class of compounds is TT14-20. One class of compounds are those where X is selected from the group consisting of : H, OMe and CO₂Me; where Y is selected from the group consisting of: H, Br, and OMe; where R¹ = R² = H; and where positions 1, 4, 5 and 8 are selected from the group OH, OMe, =O, H. Another class of compounds include those where one or more substituents contains one or
15 more N atoms. Another class of compounds includes those where X and Y are not both members of the group containing: H, OMe, Br, CO₂ Me, while positions 1, 4, 5, and 8 are substituted with -OH, -OMe or =O, or mixtures of those substituents. Another class of compounds include those which include at least one amine, amino acid or amine sugar substituent.

20 A preferred class of compounds is those which are water soluble, where one or more substituents, particularly where X, Y, R¹ and/or R² substituents of formula I are replaced with water soluble group or groups that enhance the solubility of the compound and salts thereof.

25 A preferred class are those compounds of formula I where X is a water soluble group or a group that enhances the water solubility of the compound; and salts thereof. These compounds include those where X is RO₂C (CH₂)_nNH, where n is an integer from 1 to 8 and R is as defined for I.

30 Another class of compounds includes those that contain a sulfur containing substituent.

This invention also provides methods for inhibiting cellular transport of nucleosides; inducing DNA fragmentation; inhibiting nucleic acid and/or protein synthesis and decreasing the proliferation and viability of cancer cells (including wild type and multi-drug resistant) or other cells in which the proliferation or viability is desired to be reduced, comprising contacting the cells with an effective amount of a compound of the invention as disclosed herein. This invention provides such compounds in suitable pharmaceutical carriers in dosages effective to provide measurable nucleoside transport blocking, nucleic acid and/or protein synthesis inhibition, DNA cleavage, and/or reduction in tumor cell (including wild type and multi-drug resistant) growth and/or viability. Preferably, the compounds used in the methods of this invention are almost or at least as effective as Daunomycin, a currently used anticancer drug.

Also provided is a method of treating cancer in a host, comprising: administering to said host an effective amount of an active compound of the invention for an effective time. Administration routes include intravenously, parenterally, and other methods known in the art. As used herein, an "effective amount" is an amount which causes a measurable effect on a desired parameter. As used herein, an "effective time" is the time required to cause a measurable or desired effect on a desired parameter.

Also provided is a method for preparation of triptycene analogs, comprising in situ oxidation and [4+2] cycloaddition of substituted benzenes or quinones and optionally substituted anthracenes; and optional oxidation of the resulting compounds. The resulting compounds will be a mixture of methoxy-substituted and carbonyl-substituted triptycene analogs. This mixture can be separated into individual compounds with methods known in the art. The one pot synthesis may be separated into oxidation and cycloaddition steps, if desired. Synthesis of particular groups of compounds of the invention are described in more detail herein.

Also provided is a method to synthesize 1,4-dimethoxyanthracene comprising reduction of quinizarin to give 1,4-anthraquinone; reduction of 1,4-anthraquinone to give 1,4-dihydroxyanthracene; and methylation of 1,4-dihydroxyanthracene to give 1,4-dimethoxyanthracene. These reactions are described in more detail herein. The reduction step (first step; with sodium borohydride) has been reported in: Bedworth, P. V.; Perry, J. W.; Marder, S. R. J. Chem. Soc. Chem. Commun. 1997, 1353-1354 for use in certain synthetic

methods and the following two steps (reduction with sodium hydrosulfite and methylation) have been used in certain syntheses, but preparation of 1,4-dimethoxyanthracene (1) has not been reported in one sequence of reaction.

5 A new synthesis of the compound TT2 is also provided. Either TT3, TT5 or a mixture of both is oxidized to give TT2. A new method to brominate triptycene analogs is also provided. Treatment of a triptycene analog which has a methoxy group on position 2 and a hydrogen on position 3 with N-bromosuccinimide gives an analog with a bromine on position 3. This bromination reaction can be extended to other triptycene analogs. For example, if a methoxy group is on position 6 or 7, bromination will result in a bromine on position 7 or 6, respectively. 10 If the starting compound has methoxy groups on positions 5 and 6, oxidation will give the corresponding analog with carbonyl groups on positions 5 and 6.

15 Triptycene analogs I bearing functionalities at C12 and C13 (R^3 and R^4), can be made in an analogous reaction as described below starting with 6,7-disubstituted 1,4-dimethoxyanthracenes (analogues of compound 1). These 6,7-disubstituted 1,4-dimethoxyanthracenes are prepared from the corresponding analogs of 6,7-disubstituted 1,4-dihydroxy-9,10-anthraquinones (by following the method described below).

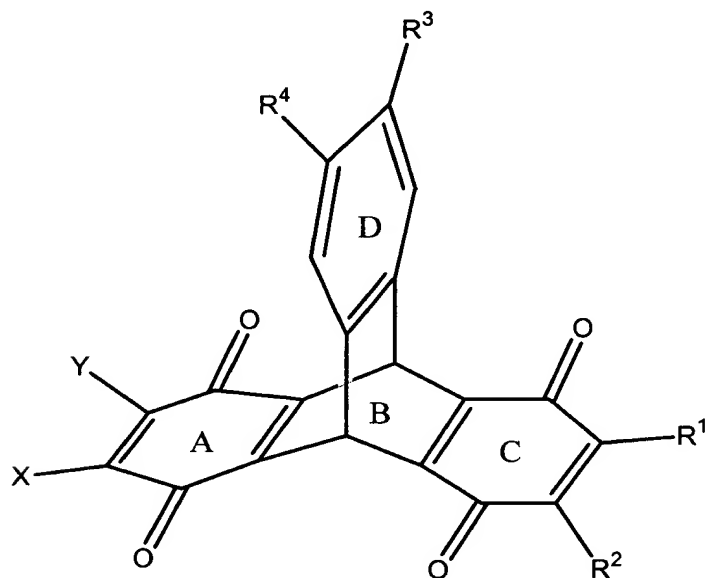
20 Also provided are triptycene analogs prepared by the methods described herein.

25 Compounds containing any combination of substituents or members of the Markush groups specified above are within the scope of the invention. All substituents of the compounds of the invention may be the same, all substituents may be different, or any combination of substituents may be the same or different. Compounds having substituents with a specified function, for example those that impart water solubility to the compound form a special class of compounds of this invention.

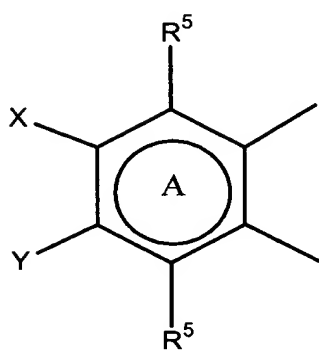
30 The substituents included in the compounds of the invention and used in the methods of the invention may be any substituent not having structures or reactivity which would substantially interfere with the desired activity of the compound, as may readily be determined without undue experimentation by those skilled in the art, for example, by using the assay methods disclosed herein and those methods known to one of ordinary skill in the art.

The reduced forms of all compounds described herein are included in the disclosure. It is understood that when referring to reduced forms of structures herein, any quinone-like ring of the structure may be replaced with a hydroquinone-like ring, as known in the art.

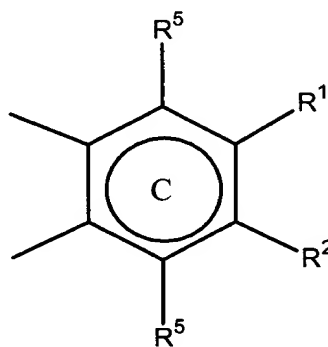
For example, in the following structure,



in reduced forms, either ring A or ring C or both is replaced with



or



wherein in reduced forms, each R^5 is independently a reduced form of $=O$, for example, OR . In the reduced forms, each R^5 may also be H or $C1-C8$ alkyl, for example, as known in the art.

BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1. Comparison of the abilities of novel TTs to inhibit the growth (open) and viability (striped) of L1210 cells after 4 days *in vitro*. For tumor cell growth and viability, L1210 cells were respectively seeded in triplicate at initial densities of 1×10^4 and 1.11×10^4 cells/0.5 ml/well in RPMI 1640 medium containing 7.5% FCS and penicillin (100 IU/ml)-streptomycin (100 μ g/ml), and grown for 4 days in the presence or absence (control) of 256 nM concentrations of the indicated compounds in a humidified incubator at 37°C with 5% CO₂ in air. Cell density was monitored using a Coulter counter. Cell growth results are expressed as % of the number of vehicle-treated control cells after 4 days in culture ($1,465,110 \pm 92,595$; $100 \pm 6\%$; open control). The ability of viable cells/0.5 ml to bioreduce 0.1 ml of MTS:PMS (20:1) reagent over a 3-h incubation period at 37°C was assessed by measuring the absorbance of the water-soluble formazan products at $A_{490 \text{ nm}}$. Cell viability results are expressed as % of the net absorbance of MTS/formazan after bio-reduction by vehicle-treated control cells ($A_{490 \text{ nm}} = 1.164 \pm 0.089$; $100 \pm 8\%$; striped control) after 4 days in culture. The blank value ($A_{490 \text{ nm}} = 0.202$) for cell-free medium supplemented with MTS:PMS reagent has been subtracted from the results. Bars: means \pm SD (n = 3). ^aNot different from TT7 and TT9; ^bP<0.025, smaller than TT1, TT7 and TT9 but not different from TT13; ^cP<0.05, greater than TT1, TT7 and TT9 but not different from TT5, TT6 and TT8; ^dnot different from control; ^eP<0.05, smaller than control.

Figure 2. The same experiment as in Figure 1 with additional compounds.

Figure 3. Comparison of the concentration-dependent inhibitions of L1210 cell growth by the TT analog TT2 (○,●) and DAU (□,■) at days 2 (open symbols) and 4 (closed symbols) *in vitro*. The protocol of the experiment was identical to that of Figure 1. The results are expressed as % of the numbers of vehicle-treated control cells ($100 \pm 4\%$; striped area) after 2 ($213,798 \pm 8,316$ cells/ml) and 4 days ($1,311,274 \pm 55,991$ cells/ml) in culture. Drug concentrations are plotted on a logarithmic scale. Bars: means \pm SD (n = 3). ^aNot different from control; ^bP<0.025 and ^cP<0.01, smaller than control.

Figure 4. Comparison of the concentration-dependent inhibitions of L1210 cell viability by the TT analog TT2 (○,●) and DAU (□,■) at days 2 (open symbols) and 4 (closed symbols) *in vitro*. Cells were seeded in triplicate at initial densities of 100,000 or 11,100 cells/0.5 ml/well and respectively incubated for 2 or 4 days in the presence or absence (control) of the indicated

concentrations of drugs, which are plotted on a logarithmic scale. The ability of viable cells/0.5 ml to bioreduce 0.1 ml of MTS:PMS (20:1) reagent over a 3-h incubation period was assessed as described in Figure 1. Cell viability results are expressed as % of the net absorbance of MTS/formazan after bio-reduction by vehicle-treated control cells ($100 \pm 8\%$; striped area) at days 2 ($A_{490\text{ nm}} = 1.206 \pm 0.093$) and 4 ($A_{490\text{ nm}} = 1.096 \pm 0.086$). Blank values ($A_{490\text{ nm}} = 0.185$ and 0.189 at days 2 and 4) for culture medium supplemented with MTS:PMS reagent have been subtracted from the results. Bars: means \pm SD ($n = 3$). ^aNot different from control; ^b $P < 0.005$, ^c $P < 0.025$ and ^d $P < 0.05$, smaller than control.

Figure 5. Comparison of the concentration-dependent inhibitions of HL-60-S (○), HL-60-RV (□) and HL-60-R8 (△) cell proliferation by DAU at days 2 (A) and 4 (B) *in vitro*. Cell growth results are expressed as % of the numbers of vehicle-treated control tumor cells ($100 \pm 5\%$, striped areas) after 2 ($110,903 \pm 5,878$ cells/ml) and 4 days ($253,462 \pm 12,166$ cells/ml) in culture. Bars: means \pm SD ($n=3$). ^aNot different from control; ^b $P < 0.05$, ^c $P < 0.01$ and ^d $P < 0.025$, smaller than control.

Figure 6. Comparison of the concentration-dependent inhibitions of HL-60-S (●), HL-60-RV (■) and HL-60-R8 (▲) cell proliferation by TT2 at days 2 (A) and 4 (B) *in vitro*. The conditions of the experiments and the determination of the results were identical to those of figure 5. Bars: means \pm SD ($n=3$). ^aNot different from control; ^b $P < 0.05$, ^c $P < 0.025$ and ^d $P < 0.005$, smaller than control.

Figure 7. Comparison of the concentration-dependent inhibitions of HL-60-S (○), HL-60-RV (□) and HL-60-R8 (△) cell viability by DAU at days 2 (A) and 4 (B) *in vitro*. Cell viability results are expressed as % of the net absorbance of MTS/formazan after bio-reduction by vehicle-treated control cells ($100 \pm 6\%$, striped areas) at days 2 ($A_{490\text{ nm}} = 1.635 \pm 0.105$) and 4 ($A_{490\text{ nm}} = 1.428 \pm 0.086$). Blank values ($A_{490\text{ nm}} = 0.412$ and 0.436 at days 2 and 4) for cell-free culture medium supplemented with MTS: PMS reagent have been subtracted from the results. Bars: means \pm SD ($n=3$). ^aNot different from control; ^b $P < 0.05$, ^c $P < 0.025$ and ^d $P < 0.01$, smaller than control.

Figure 8. Comparison of the concentration-dependent inhibitions of HL-60-S (●), HL-60-RV (■) and HL-60-R8 (▲) cell viability by TT2 at days 2 (A) and 4 (B) *in vitro*. The conditions of the experiments and the determination of the results were identical to those of figure 5. Bars: means \pm SD ($n=3$). ^aNot different from control; ^b $P < 0.025$, smaller than control.

Figure 9. Comparison of the concentration-dependent inhibitions of DNA synthesis by the TT analog TT2 (●) and DAU (○) in L1210 cells *in vitro*. Cells ($1.14 \times 10^6/0.5$ ml of RPMI 1640 medium) were incubated at 37°C for 90 min in the presence or absence (control) of the indicated concentrations of drugs, which are plotted on a logarithmic scale, and then pulse-labeled for an additional 30 min to determine the rate of ^3H -thymidine incorporation into DNA. DNA synthesis in vehicle-treated control cells was $27,860 \pm 1,931$ cpm ($100 \pm 7\%$; striped area). The blank value (916 ± 52 cpm) for cells incubated and pulse-labeled at 2°C with 1 μCi of ^3H -thymidine has been subtracted from the results. Bars: means \pm SD ($n = 3$). ^aNot different from control; ^b $P < 0.05$ and ^c $P < 0.01$, smaller than control.

Figure 10. Concentration-dependent inhibition of nucleoside transport by the TT analog TT2 in L1210 cells *in vitro*. Cells ($1.48 \times 10^6/0.5$ ml of RPMI 1640 medium) were preincubated for 15 min at 37°C in the presence or absence (control) of the indicated concentrations of TT2 (●) and DAU (○), which are plotted on a logarithmic scale, before being exposed to 1 μCi of ^3H -thymidine for 30 sec at 37°C. After washing thrice with PBS, intact cell pellets were harvested by centrifugation, incubated for 30 min in 1 ml of HLB containing 0.2% Triton X-100, and these lysates were mixed with scintillation cocktail and counted to estimate the cellular uptake of ^3H -thymidine. Results are expressed as % of ^3H -thymidine transported into vehicle-treated control cells over 30 sec ($15,031 \pm 872$ cpm; $100 \pm 6\%$; striped area). Bars: means \pm SD ($n = 3$). ^aNot different from control; ^b $P < 0.005$, smaller than control.

Figure 11. Comparison of the inhibitory effects of novel TT analogs on DNA synthesis (open) and nucleoside transport (striped) in L1210 cells *in vitro*. The cellular uptake of ^3H -thymidine (striped) and the rate of ^3H -thymidine incorporation into DNA (open) were determined in cells ($1.38 \times 10^6/0.5$ ml of RPMI 1640 medium) respectively preincubated at 37°C for 15 or 90 min in the presence or absence (control) of 7 μM concentrations of the indicated compounds. For the cellular transport of nucleosides, preincubated cells were then exposed to 1 μCi of ^3H -thymidine for 30 sec at 37°C. Results are expressed as % of ^3H -thymidine transported into vehicle-treated control cells over 30 sec ($15,539 \pm 1,767$ cpm; $100 \pm 11\%$; striped control). For DNA synthesis, preincubated cells were then pulse-labeled with 1 μCi of ^3H -thymidine for an additional 30 min at 37°C. Results are expressed as % of ^3H -thymidine incorporation into DNA in vehicle-treated control cells over 30 min ($28,505 \pm 2,594$ cpm; $100 \pm 9\%$; open control). The blank value ($1,075 \pm 171$ cpm) for cells incubated and pulse-labeled at 2°C with 1 μCi of ^3H -

thymidine has been subtracted from the results. Bars: means \pm SD (n = 3). ^aNot different from TT3, TT7, TT9 and TT11; ^bP<0.01, smaller than TT1 but not different from TT13; ^cnot different from control; ^dP<0.025, greater than TT1; ^eP<0.05, smaller than control.

Figure 12. Irreversibility of the inhibitory effects of the novel TT analog TT2 on DNA synthesis (open) and nucleoside transport (striped) in L1210 cells *in vitro*. The reversible inhibitory effects of the tricyclic pyrone H10 are demonstrated in the same experiment. The cellular uptake of ³H-thymidine (striped) and the rate of ³H-thymidine incorporation into DNA (open) were determined in cells (1.64 x 10⁶/0.5 ml of RPMI 1640 medium) respectively preincubated at 37°C for 15 or 60 min in the presence or absence (control) of 10 and 25 μ M TT2 or 25 μ M H10. The protocol of the experiments and the determination of the results were identical to those of Figures 9 and 10, except that, after preincubation, either the drugs were maintained in the culture medium (+) or the cells were spun, washed, and resuspended in fresh medium in order to remove the drugs (-). After preincubation, vehicle-treated controls were similarly spun and washed. For the cellular transport of nucleosides, preincubated cells were then exposed, in the presence (+) or absence (-) of TT2 or H10, to 1 μ Ci of ³H-thymidine for 30 sec at 37°C. Results are expressed as % of ³H-thymidine transported into vehicle-treated control cells over 30 sec (11,054 \pm 754 cpm; 100 \pm 7%; striped control +; 14,486 \pm 1,260 cpm; 100 \pm 9%; striped control -). For DNA synthesis, preincubated cells were then pulse-labeled with 1 μ Ci of ³H-thymidine for an additional 30 min at 37°C in the presence (+) or absence (-) of TT2 or H10. Results are expressed as % of ³H-thymidine incorporation into DNA in vehicle-treated control cells over 30 min (34,052 \pm 1,566 cpm; 100 \pm 5%; open control +; 28,811 \pm 2,506 cpm; 100 \pm 9%; open control -). The blank value (1,328 \pm 86 cpm) for cells incubated and pulse-labeled at 2°C with 1 μ Ci of ³H-thymidine has been subtracted from the results. Bars: means \pm SD (n = 3). Values with similar superscripts are not significantly different from each others.

Figure 13. The open columns represents the rate of ³H - Thymidine incorporation into DNA over 30 minutes, whereas the striped and closed columns represents the cellular transport of ³H - adenosine and ³H-thymidine occurring over only 30 seconds *in vitro*.

Figure 14. Comparison of the concentration-dependent inhibitions of DNA synthesis by DAU (open symbols in A) and TT2 (closed symbols in B) in HL-60-S (circles), HL-60-RV (squares) and HL-60-R8 (triangles) cells *in vitro*. DNA synthesis in vehicle-treated control cells was 14,269 \pm 1,106 cpm (100 \pm 8%, striped areas). The blank value (404 \pm 38 cpm) for control cells incubated and pulse-labeled at 2°C with 1 μ Ci of ³H-thymidine has been subtracted from

the results. Bars: means \pm SD (n=3). ^aNot different from control; ^bP<0.05 and ^cP<0.025, smaller than control.

Figure 15. Comparison of the concentration-dependent inhibitions of the cellular transport of purine (A) and pyrimidine (B) nucleosides by DAU (open symbols) and TT2 (closed symbols) in HL-60-S (circles), HL-60-RV (squares) and HL-60-R8 (triangles) cells *in vitro*. Results are expressed as % of ³H-adenosine (26,032 \pm 2,239 cpm, 100 \pm 9%, striped area in A) and ³H-thymidine (17,934 \pm 1,398 cpm, 100 \pm 8%, striped area in B) transported into vehicle-treated control cells over 30 s. Bars: means \pm SD (n=3). ^aNot different from control; ^bP<0.025, ^cP<0.01 and ^dP<0.005, smaller than control.

Figure 16. Comparison of the concentration-dependent inhibitions of RNA synthesis by the TT analog TT2 (●) and DAU (○) in L1210 cells *in vitro*. Cells (1.2 x 10⁶/0.5 ml of RPMI 1640 medium) were incubated at 37°C for 3h in the presence or absence (control) of the indicated concentrations of drugs, which are plotted on a logarithmic scale, and then pulse-labeled for an additional 1h to determine the rate of ³H-uridine incorporation into RNA. RNA synthesis in vehicle-treated control cells was 47,117 \pm 3,204 cpm (100 \pm 7%; striped area). The blank value (1,746 \pm 155 cpm) for cells incubated and pulse-labeled at 2°C with 2 μ Ci of ³H-uridine has been subtracted from the results. Bars: means \pm SD (n = 3). ^aNot different from control; ^bP<0.05 and ^cP<0.025, smaller than control.

Figure 17. Comparison of the concentration-dependent inhibitions of protein synthesis by the TT analog TT2 (●) and DAU (○) in L1210 cells *in vitro*. Cells (1.51 x 10⁶/0.5 ml of RPMI 1640 medium) were incubated at 37°C for 3h in the presence or absence (control) of the indicated concentrations of drugs, which are plotted on a logarithmic scale, and then pulse-labeled for an additional 1h to determine the rate of ³H-leucine incorporation into protein. Protein synthesis in vehicle-treated control cells was 13,812 \pm 1,070 cpm (100 \pm 8%; striped area). The blank value (2,436 \pm 298 cpm) for cells incubated and pulse-labeled at 2°C with 2.5 μ Ci of ³H-leucine has been subtracted from the results. Bars: means \pm SD (n = 3). ^aNot different from control; ^bP<0.025 and ^cP<0.01, smaller than control.

Figure 18. Effects of the novel TT analog TT2 and DAU on the mitotic index of L1210 cells *in vitro*. The antimitotic activities of VCR and the tricyclic pyrone H10 are demonstrated in the same experiment. Cells (10⁶/0.5 ml of RPMI 1640 medium) were incubated in triplicate for 24h at 37°C in the presence or absence (control) of 0.256, 0.64, 1.6 and 4 μ M TT2, 0.256,

0.64 and 1.6 μ M DAU, 0.2 μ M VCR or 1 μ M H10. After fixation with MeOH:acetic acid (3:1) and staining with 0.1% crystal violet, about 500 cells/slide were scored for mitotic figures and the mitotic index was expressed as the % of mitotic cells in drug-treated cultures divided by the % of mitotic cells in non-treated controls. The mean % of mitotic cells in control at 24h was 1.86 \pm 0.43%. Bars: means \pm SD (n = 3). ^aNot different from control; ^bP<0.05, ^cP<0.01 and ^dP<0.005, smaller than control; ^eP<0.005, greater than control but smaller than VCR.

Figure 19. Comparison of the concentration-dependent effects of novel TT analogs and known DNA-damaging anticancer drugs on DNA cleavage in L1210 cells *in vitro*. Cells ($10^6/0.5$ ml of RPMI 1640 medium) were prelabeled with 1 μ Ci of ³H-thymidine for 2h, washed and resuspended in fresh FCS-containing medium, and incubated at 37°C for 24h in the presence or absence (control) of the indicated concentrations of TT1 (○), TT2 (●), DAU (■) or CPT (▲), which are plotted on a logarithmic scale. After lysing the cells in HLB containing 0.2% Triton X-100, the detergent-soluble DNA fragments present in the supernatants and the intact chromatin DNA remaining in the pellets were separated by centrifugation and their radioactivity estimated by LSC. Results are expressed as [cpm in supernatant/cpm in supernatant + pellet] \times 100 at 24h. For untreated controls (5.3 \pm 0.6% DNA fragmentation; striped area), the supernatant is 741 \pm 79 cpm and the pellet is 13,293 \pm 1,446 cpm. Bars: means \pm SD (n = 3). ^aNot different from control; ^bP<0.05 and ^cP<0.01, greater than control; ^dP<0.05, smaller than CPT but not different from 10 μ M TT1; ^eP<0.05, greater than TT1 but not different from CPT.

Figure 20. Comparison of the abilities of novel TT analogs to induce DNA cleavage in L1210 cells *in vitro*. Cells containing prelabeled DNA were incubated at 37°C for 24h in the presence or absence (control) of 7 μ M concentrations of the indicated compounds. The protocol of the experiment and the determination of the results were identical to those of Figure 12. Bars: means \pm SD (n = 3). ^aNot different from TT7 and TT9; ^bP<0.05, greater than TT9 but not different from TT13; ^cP<0.05, smaller than TT7; ^dnot different from control; ^eP<0.01, greater than control, P<0.025, greater than TT11 but not different from TT3, TT6, TT8 and TT10; ^fP<0.01, greater than control.

Figure 21. Same experiment as Figure 20 with additional analogs.

Figure 22. Irreversibility of the effects of novel TT analogs and known DNA-damaging anticancer drugs on DNA cleavage in L1210 cells *in vitro*. Cells containing prelabeled DNA were incubated at 37°C for various periods of time in the presence or absence (control) of 10 μ M

TT1 or TT2 and 1.6 μ M DAU or CPT. The protocol of the experiment and the determination of the results at 24h were identical to those of Figure 19, except that the drugs were either maintained in the medium for the whole 24-h period of incubation (open) or removed after the first 3 (striped) and 8h (closed). After spinning and washing the appropriate samples to remove the drugs, cells were resuspended in 0.5 ml of fresh medium to complete the 24-h period of incubation in the absence of drugs. Vehicle-treated controls were similarly spun and washed at 3 and 8h. Bars: means \pm SD (n = 3). Values with similar superscripts are not significantly different from each others. ^bNot different from DAU; ^cP<0.025, greater than TT1 but not different from CPT; ^dP<0.05, smaller than CPT.

Figure 23. Comparison of the concentration-dependent inductions of DNA cleavage by DAU (open symbols in A) and TT2 (closed symbols in B) at 24 h in HL-60-S (circles), HL-60-RV (squares) and HL-60-R8 (triangles) cells containing ³H-prelabeled DNA *in vitro*. Results are expressed as [cpm in supernatant/cpm in supernatant + pellet] x 100 at 24 h. For untreated controls (14.7 \pm 1.7% DNA fragmentation, striped areas), the supernatant (DNA fragments) is 12,815 \pm 1,358 cpm and the pellet (intact DNA) is 74,406 \pm 9,078 cpm. Bars: means \pm SD (n=3). ^aNot different from control; ^bP<0.05, ^cP<0.025 and ^dP<0.01, greater than control; ^eP<0.005, smaller than the effect of 1.6 μ M DAU in HL-60-S cells but not different from the effect of 0.1024 μ M DAU in HL-60-S cells; ^fnot different from the effect of 0.04096 μ M DAU in HL-60-S cells; ^gP<0.05, greater than the effect of 4 μ M TT2 in HL-60-RV cells but not different from the effect of 4 μ M TT2 in HL-60-R8 cells; ^hnot different from the effect of 10 μ M TT2 in HL-60-RV and HL-60-R8 cells; ⁱP<0.005, smaller than control.

DETAILED DESCRIPTION OF THE INVENTION

DEFINITIONS

The term "hydrocarbyl" is used herein to refer generally to organic groups comprised of carbon chains to which hydrogen and optionally other elements are attached. CH₂ or CH groups and C atoms of the carbon chains of the hydrocarbyl may be replaced with one or more heteroatoms (i.e., non-carbon atoms). Suitable heteroatoms include but are not limited to O, S, P and N atoms. The term hydrocarbyl includes, but is not limited to alkyl, alkenyl, alkynyl, ether, polyether, thioether, straight chain or cyclic saccharides, ascorbate, aminoalkyl, hydroxylalkyl, thioalkyl, aryl and heterocyclic aryl groups, optionally substituted tricyclic molecules, amino acid, polyalcohol, glycol, groups which have a mixture of saturated and unsaturated bonds,

carbocyclic rings and combinations of such groups. The term also includes straight-chain, branched-chain and cyclic structures or combinations thereof. Hydrocarbyl groups are optionally substituted. Hydrocarbyl substitution includes substitution at one or more carbons in the group by moieties containing heteroatoms. Suitable substituents for hydrocarbyl groups include but are not limited to halogens, including chlorine, fluorine, bromine and iodine, OH, SH, NH₂, COH, CO₂H, OR_a, SR_a, NR_aR_b, CONR_aR_b, where R_a and R_b independently are alkyl, unsaturated alkyl or aryl groups.

The term "alkyl" takes its usual meaning in the art and is intended to include straight-chain, branched and cycloalkyl groups. The term includes, but is not limited to, methyl, ethyl, n-propyl, isopropyl, n-butyl, sec-butyl, isobutyl, tert-butyl, n-pentyl, neopentyl, 2-methylbutyl, 1-methylbutyl, 1-ethylpropyl, 1,1-dimethylpropyl, n-hexyl, 1-methylpentyl, 2-methylpentyl, 3-methylpentyl, 4-methylpentyl, 3,3-dimethylbutyl, 2,2-dimethylbutyl, 1,1-dimethylbutyl, 2-ethylbutyl, 1-ethylbutyl, 1,3-dimethylbutyl, n-heptyl, 5-methylhexyl, 4-methylhexyl, 3-methylhexyl, 2-methylhexyl, 1-methylhexyl, 3-ethylpentyl, 2-ethylpentyl, 1-ethylpentyl, 4,4-dimethylpentyl, 3,3-dimethylpentyl, 2,2-dimethylpentyl, 1,1-dimethylpentyl, n-octyl, 6-methylheptyl, 5-methylheptyl, 4-methylheptyl, 3-methylheptyl, 2-methylheptyl, 1-methylheptyl, 1-ethylhexyl, 1-propylpentyl, 3-ethylhexyl, 5,5-dimethylhexyl, 4,4-dimethylhexyl, 2,2-diethylbutyl, 3,3-diethylbutyl, and 1-methyl-1-propylbutyl. Alkyl groups are optionally substituted. Lower alkyl groups are C₁-C₆ alkyl and include among others methyl, ethyl, n-propyl, and isopropyl groups.

The term "cycloalkyl" refers to alkyl groups having a hydrocarbon ring, particularly to those having rings of 3 to 7 carbon atoms. Cycloalkyl groups include those with alkyl group substitution on the ring. Cycloalkyl groups can include straight-chain and branched-chain portions. Cycloalkyl groups include but are not limited to cyclopropyl, cyclobutyl, cyclopentyl, cyclohexyl, cycloheptyl, cyclooctyl, and cyclononyl. Cycloalkyl groups can optionally be substituted.

Aryl groups may be substituted with one, two or more simple substituents including, but not limited to, lower alkyl, e.g., methyl, ethyl, butyl; halo, e.g., chloro, bromo; nitro; sulfato; sulfonyloxy; carboxy; carbo-lower-alkoxy, e.g., carbomethoxy, carbethoxy; amino; mono- and di-lower-alkylamino, e.g., methylamino, ethylamino, dimethylamino, methylethylamino; amido; hydroxy; lower-alkoxy, e.g., methoxy, ethoxy; and lower-alkanoyloxy, e.g., acetoxy.

The term "unsaturated alkyl" group is used herein generally to include alkyl groups in which one or more carbon-carbon single bonds have been converted to carbon-carbon double or triple bonds. The term includes alkenyl and alkynyl groups in their most general sense. The term is intended to include groups having more than one double or triple bond, or combinations of double and triple bonds. Unsaturated alkyl groups include, without limitation, unsaturated straight-chain, branched or cycloalkyl groups. Unsaturated alkyl groups include without limitation: vinyl, allyl, propenyl, isopropenyl, butenyl, pentenyl, hexenyl, hexadienyl, heptenyl, cyclopropenyl, cyclobutenyl, cyclopentenyl, cyclopentadienyl, cyclohexenyl, cyclohexadienyl, 1-propenyl, 2-butenyl, 2-methyl-2-butenyl, ethynyl, propargyl, 3-methyl-1-pentynyl, and 2-heptynyl. Unsaturated alkyl groups can optionally be substituted.

Substitution of alkyl, cycloalkyl and unsaturated alkyl groups includes substitution at one or more carbons in the group by moieties containing heteroatoms. Suitable substituents for these groups include but are not limited to OH, SH, NH₂, COH, CO₂H, OR_c, SR_c, P, PO, NR_cR_d, CONR_cR_d, and halogens, particularly chlorines and bromines where R_c and R_d, independently, are alkyl, unsaturated alkyl or aryl groups. Preferred alkyl and unsaturated alkyl groups are the lower alkyl, alkenyl or alkynyl groups having from 1 to about 3 carbon atoms.

The term "aryl" is used herein generally to refer to aromatic groups which have at least one ring having a conjugated pi electron system and includes without limitation carbocyclic aryl, aralkyl, heterocyclic aryl, biaryl groups and heterocyclic biaryl, all of which can be optionally substituted. Preferred aryl groups have one or two aromatic rings.

"Carbocyclic aryl" refers to aryl groups in which the aromatic ring atoms are all carbons and includes without limitation phenyl, biphenyl and naphthalene groups.

"Aralkyl" refers to an alkyl group substituted with an aryl group. Suitable aralkyl groups include among others benzyl, phenethyl and picolyl, and may be optionally substituted. Aralkyl groups include those with heterocyclic and carbocyclic aromatic moieties.

"Heterocyclic aryl groups" refers to groups having at least one heterocyclic aromatic ring with from 1 to 3 heteroatoms in the ring, the remainder being carbon atoms. Suitable heteroatoms include without limitation oxygen, sulfur, and nitrogen. Heterocyclic aryl groups include among others furanyl, thienyl, pyridyl, pyrrolyl, N-alkyl pyrrolo, pyrimidyl, pyrazinyl, imidazolyl, benzofuranyl, quinolinyl, and indolyl, all optionally substituted.

"Heterocyclic biaryl" refers to heterocyclic aryls in which a phenyl group is substituted by a heterocyclic aryl group ortho, meta or para to the point of attachment of the phenyl ring to the decalin or cyclohexane. Heterocyclic biaryl includes among others groups which have a phenyl group substituted with a heterocyclic aromatic ring. The aromatic rings in the heterocyclic biaryl group can be optionally substituted.

"Biaryl" refers to carbocyclic aryl groups in which a phenyl group is substituted by a carbocyclic aryl group ortho, meta or para to the point of attachment of the phenyl ring to the decalin or cyclohexane. Biaryl groups include among others a first phenyl group substituted with a second phenyl ring ortho, meta or para to the point of attachment of the first phenyl ring to the decalin or cyclohexane structure. Para substitution is preferred. The aromatic rings in the biaryl group can be optionally substituted.

Aryl group substitution includes substitutions by non-aryl groups (excluding H) at one or more carbons or where possible at one or more heteroatoms in aromatic rings in the aryl group. Unsubstituted aryl, in contrast, refers to aryl groups in which the aromatic ring carbons are all substituted with H, e.g. unsubstituted phenyl ($-C_6H_5$), or naphthyl ($-C_{10}H_7$). Suitable substituents for aryl groups include among others, alkyl groups, unsaturated alkyl groups, halogens, OH, SH, NH_2 , COH, CO_2H , OR_e , SR_e , NR_eR_f , $CONR_eR_f$, where R_e and R_f independently are alkyl, unsaturated alkyl or aryl groups. Preferred substituents are OH, SH, OR_e , and SR_e where R_e is a lower alkyl, i.e., an alkyl group having from 1 to about 3 carbon atoms. Other preferred substituents are halogens, more preferably chlorine or bromine, and lower alkyl and unsaturated lower alkyl groups having from 1 to about 3 carbon atoms. Substituents include bridging groups between aromatic rings in the aryl group, such as $-CO_2-$, $-CO-$, $-O-$, $-S-$, $-P-$, $-NH-$, $-CH=CH-$ and $-(CH_2)_\ell-$ where ℓ is an integer from 1 to about 5, and particularly $-CH_2-$. Examples of aryl groups having bridging substituents include phenylbenzoate. Substituents also include moieties, such as $-(CH_2)_\ell-$, $-O-(CH_2)_\ell-$ or $-OCO-(CH_2)_\ell-$, where ℓ is an integer from about 2 to 7, as appropriate for the moiety, which bridge two ring atoms in a single aromatic ring as, for example, in a 1, 2, 3, 4-tetrahydronaphthalene group. Alkyl and unsaturated alkyl substituents of aryl groups can in turn optionally be substituted as described supra for substituted alkyl and unsaturated alkyl groups.

The terms "alkoxy group" and "thioalkoxy group" (also known as mercaptide groups, the sulfur analog of alkoxy groups) take their generally accepted meaning. Alkoxy groups include but are not limited to methoxy, ethoxy, n-propoxy, isopropoxy, n-butoxy, sec-butoxy, isobutoxy,

tert-butoxy, n-pentyloxy, neopentyloxy, 2-methylbutoxy, 1-methylbutoxy, 1-ethyl propoxy, 1,1-dimethylpropoxy, n-hexyloxy, 1-methylpentyloxy, 2-methylpentyloxy, 3-methylpentyloxy, 4-methylpentyloxy, 3,3-dimethylbutoxy, 2,2-dimethoxybutoxy, 1-1-dimethylbutoxy, 2-ethylbutoxy, 1-ethylbutoxy, 1,3-dimethylbutoxy, n-pentyloxy, 5-methylhexyloxy, 4-methylhexyloxy, 3-methylhexyloxy, 2-methylhexyloxy, 1-methylhexyloxy, 3-ethylpentyloxy, 2-ethylpentyloxy, 1-ethylpentyloxy, 4,4-dimethylpentyloxy, 3,3-dimethylpentyloxy, 2,2-dimethylpentyloxy, 1,1-dimethylpentyloxy, n-octyloxy, 6-methylheptyloxy, 5-methylheptyloxy, 4-methylheptyloxy, 3-methylheptyloxy, 2-methylheptyloxy, 1-methylheptyloxy, 1-ethylhexyloxy, 1-propylpentyloxy, 3-ethylhexyloxy, 5,5-dimethylhexyloxy, 4,4-dimethylhexyloxy, 2,2-diethylbutoxy, 3,3-diethylbutoxy, 1-methyl-1-propylbutoxy, ethoxymethyl, n-propoxymethyl, isopropoxymethyl, sec-butoxymethyl, isobutoxymethyl, (1-ethyl propoxy)methyl, (2-ethylbutoxy)methyl, (1-ethylbutoxy)methyl, (2-ethylpentyloxy)methyl, (3-ethylpentyloxy)methyl, 2-methoxyethyl, 1-methoxyethyl, 2-ethoxyethyl, 3-methoxypropyl, 2-methoxypropyl, 1-methoxypropyl, 2-ethoxypropyl, 3-(n-propoxy)propyl, 4-methoxybutyl, 2-methoxybutyl, 4-ethoxybutyl, 2-ethoxybutyl, 5-ethoxypentyl, and 6-ethoxyhexyl. Thioalkoxy groups include but are not limited to the sulfur analogs of the alkoxy groups specifically listed supra.

"Optional" or "optionally" means that the subsequently described event or circumstance may or may not occur, and that the description includes instances where said event or circumstance occurs and instances in which it does not. For example, "optionally substituted phenyl" means that the phenyl radical may or may not be substituted and that the description includes both unsubstituted phenyl radicals and phenyl radicals wherein there is substitution.

"Amino acids" as used herein include naturally occurring and commercially available amino acids and optical isomers thereof. Typical natural and commercially available amino acids are glycine, alanine, serine, homoserine, threonine, valine, norvaline, leucine, isoleucine, norleucine, aspartic acid, glutamic acid, lysine, ornithine, histidine, arginine, cysteine, homocysteine, methionine, phenylalanine, homophenylalanine, phenylglycine, o-, m-, and p-tyrosine, tryptophan, glutamine, asparagine, proline and hydroxyproline. "Amino acid" as used herein includes amino acid residues and amino acid side chains. An "amino acid residue" is an amino acid radical $--NHCH(R)C(O)--$, wherein R is an amino acid side chain, except for the amino acid residues of proline and hydroxyproline which are $--N(CH_2-CH_2-CH_2)CHC(O)--$ and $--N(CH-CHOHCH_2)CHC(O)--$, respectively. An amino acid side chain is a radical found on the α -carbon of an α -amino acid, where the radical is either hydrogen (side chain of glycine), methyl

(side chain of alanine), or is a radical bonded to the α -carbon by a methylene ($--CH_2--$), or phenyl group.

"Contacting" reaction components with each other refers to providing a medium and/or reaction chamber in which the reaction components are placed together so that they can react with each other. Preferably, the reaction components are suspended or dissolved in a carrier fluid which is a liquid medium. "Maintaining reaction components in contact" means keeping the components together in such a way that they can react with each other.

"Straight chain or cyclic saccharides" include mono-, di- and poly-, straight chain and cyclic saccharides that are optionally substituted with an amino group which is optionally acetylated. Straight chain saccharides that are useful in this invention include but are not limited to those molecules with a chain of 5 or 6 carbon atoms with one or more -OH groups attached, and either an aldehyde or ketone group. Cyclic saccharides are saccharides that are in a ring form. Disaccharides are compounds wherein two monosaccharide groups are linked. Polysaccharides are compounds wherein more than two monosaccharide groups are linked. Specific examples of saccharides useful in this invention include glucose, ribose and glucosamine, among others.

Substituents which impart water solubility include but are not limited to alcohols; polyalcohols; straight chain or cyclic saccharides; amines and polyamines; sulfate groups; phosphate groups; ascorbate groups; alkyl chains optionally substituted with -OH at any position; glycols, including polyethylene glycols, and polyethers. Substituents which impart water solubility are also referred to as solubilizing groups.

This invention is also directed to pharmaceutically acceptable salts of the various formulas and structures disclosed herein. Acid addition salts are prepared by contacting compounds having appropriate basic groups therein with an acid whose anion is generally considered suitable for human or animal consumption. Pharmacologically acceptable acid addition salts include but are not limited to the hydrochloride, hydrobromide, hydroiodide, sulfate, phosphate, acetate, propionate, lactate, maleate, malate, succinate, and tartrate salts. All of these salts can be prepared by conventional means by reacting, for example, the selected acid with the selected basic compound. Base addition salts are analogously prepared by contacting compounds having appropriate acidic groups therein with a base whose cation is generally

considered to be suitable for human or animal consumption. Pharmacologically acceptable base addition salts, include but are not limited to ammonium, amine and amide salts.

Pharmaceutically acceptable esters of compounds of this invention are prepared by conventional methods, for example by reaction with selected acids. Pharmaceutically acceptable esters include but are not limited to carboxylic acid esters RCOO-D (where D is a cationic form of a compound of this invention and where R is H, alkyl or aryl groups).

Effective dosages of the compounds of this invention may be easily determined by those skilled in the art following the teachings hereof and principles known to the art.

The compounds of these inventions may be administered in the form of pharmaceutical preparations including the compounds of these inventions in suitable pharmaceutical carriers to form solutions, lotions, creams, and other dosage forms known to the art. Combinations of such compounds with pharmaceutical carriers are also provided by this invention. Combinations of triptycene analogs described herein along with other compounds which may include other triptycene analogs described herein are also provided by this invention.

This invention is also directed to prodrugs and analogs which on being metabolized will result in any of the effective triptycene analogs of this invention. For example, alkoxy or acetate groups can be metabolized to hydrogens. Labile substituents may be protected employing conventional and pharmaceutically acceptable protecting groups removable on metabolism. Pharmaceutically active compounds may be derivatized by conventional methods to provide for extended metabolic half-life, to enhance solubility in a given carrier, to provide for or facilitate slow-release or timed-release or enhance or affect other drug delivery properties.

Pharmaceutical compositions according to the present invention comprise one or more triptycene compounds, salts or esters of this invention in association with a pharmaceutically acceptable carrier or excipient adapted for use in human or veterinary medicine. Such compositions may be prepared for use in conventional manner in admixture with one or more physiologically acceptable carriers or excipients. The compositions may optionally further contain one or more other therapeutic agents. The compounds, salts or esters of this invention are present in these pharmaceutical compositions in an amount or in a combined amount sufficient to elicit a measurable positive effect on a desired parameter, or a desired physiological effect. The triptycene compounds, salts and esters of this invention may be formulated for oral,

buccal, parenteral, topical or rectal administration. In particular, they may be presented in unit dose form. The compositions may take such forms as suspensions, solutions, or emulsions in oily or aqueous vehicles, and may contain formulatory agents such as suspending, stabilizing and/or dispersing agents. Alternatively, the active ingredient may be in powder form for constitution with a suitable vehicle, e.g. sterile, pyrogen-free water, before use.

The pharmaceutical compositions according to the invention may also contain other active ingredients, such as antimicrobial agents, or preservatives.

The invention further provides a process for preparing a pharmaceutical composition which comprises bringing a triptycene analog of the invention into association with a pharmaceutically acceptable excipient or carrier. The carrier or excipient being selected as is known in the art for compatibility with the desired means of administration, for compatibility with the selected compounds and to minimize detrimental effects to the patient.

The magnitude of a prophylactic or therapeutic dose of a particular compound will, of course, vary with the nature of the severity of the condition to be treated, the particular triptycene analog and its route of administration. It will also vary according to the age, weight and response of the individual patient, all as will be readily ascertainable to those skilled in the art.

The compounds of the present invention are preferably formulated prior to administration. The present pharmaceutical formulations are prepared by known procedures using well-known and readily available ingredients. In making the compositions of the present invention, the active ingredient will usually be mixed with a carrier, or diluted by a carrier, or enclosed within a carrier which may be in the form of a capsule, sachet, paper or other container. When the carrier serves as a diluent, it may be a solid, semi-solid or liquid material which acts as a vehicle, excipient or medium for the active ingredient. The compositions can be in the form of tablets, pills, powders, lozenges, sachets, cachets, elixirs, suspensions, emulsions, solutions, syrups, aerosols (as a solid or in a liquid medium), ointments containing for example up to 10% by weight of the active compound, soft and hard gelatin capsules, suppositories, sterile injectable solutions and sterile packaged powders.

Some examples of suitable carriers, excipients, and diluents include lactose, dextrose, sucrose, sorbitol, mannitol, starches, gum acacia, calcium phosphate, alginates, tragacanth, gelatin, calcium silicate, microcrystalline cellulose, polyvinylpyrrolidone, cellulose, water, syrup,

methylellulose, methyl and propylhydroxybenzoates, talc, magnesium stearate and mineral oil. The formulations can additionally include lubricating agents, wetting agents, emulsifying and suspending agents, preserving agents, sweetening agents or flavoring agents. The compositions of the invention may be formulated so as to provide quick, sustained or delayed release of the active ingredient after administration to the patient by employing procedures well known in the art.

The term "unit dosage form" refers to physically discrete units suitable as unitary dosages for human subjects and other mammals, each unit containing a predetermined quantity of active material calculated to produce the desired therapeutic effect, in association with a suitable pharmaceutical carrier.

This invention is further directed to therapeutic methods employing the triptycene compounds of this invention and pharmaceutical compositions containing them in the treatment of disorders or physiological conditions involving abnormal cell growth or activity. These methods comprise a step of administering to a patient having the disorder or symptoms thereof a pharmaceutical composition comprising one or a mixture of the compounds, salts or esters of this invention where the compounds, or mixtures of compounds of this invention are present in the composition at a level or a combined level sufficient to effect a positive biological response. The present invention provides triptycene analogs that can be used in place of or in combination with currently known pharmaceuticals active against disorders such as cancer and malaria. Compounds of this invention exhibit improved properties (enhanced activity and/or decreased undesired side-effects) for treatment of such disorders as compared to previously known compounds useful for such treatments.

Table 1 lists the chemical names of the compounds depicted in Scheme 1.

Table 1: Nomenclature of various substituted 9,10-Dihydro-9,10-[1',2']benzenoanthracene-1,4,5,8-tetraones.

TT1: 9,10-Dihydro-9,10-[1',2']benzenoanthracene-1,4,5,8-tetraone.

TT2: 2-Methoxy-9,10-dihydro-9,10-[1',2']benzenoanthracene-1,4,5,8-tetraone.

TT13: 2-Bromo-3-methoxy-9,10-dihydro-9,10-[1',2']benzenoanthracene-1,4,5,8-tetraone.

TT14: 2-Bromo-3-(methylamino)-9,10-dihydro-9,10-[1',2']benzenoanthracene-1,4,5,8-tetraone.

TT3: 1,4-Dihydroxy-2,5,8-trimethoxy-9,10-dihydro-9,10-[1',2']benzenoanthracene.

TT5: 2,5,8-Trimethoxy-9,10-dihydro-9,10-[1',2']benzenoanthracene-1,4-dione.

TT7: 9,10-Dihydro-9,10-[1',2']benzenoanthracene-1,4-dione.

TT9: 2-Methoxy-9,10-dihydro-9,10-[1',2']benzenoanthracene-1,4-dione.

TT6: (4a*S**,9a*R**)-4a,9,9a,10-Tetrahydro-9,10-[1',2']benzenoanthracene-1,4-dione.

TT8: (4a*S**,9a*R**)-2-Methoxy-4a,9,9a,10-tetrahydro-9,10-[1',2']benzenoanthracene-1,4-dione.

TT10: (4a*S**,9a*R**)-5,8-Dimethoxy-2-methoxycarbonyl-4a,9,9a,10-tetrahydro-9,10-[1',2']benzenoanthracene-1,4-dione.

TT11: (4a*R**,9a*R**)-5,8-Dimethoxy-4a-methoxycarbonyl-9,9a,10-trihydro-9,10-[1',2']benzenoanthracene-1,4-dione.

TT4: (4a*S**,7a*R**,11a*S**,14a*R**)-Tetramethyl 1,4,4a,5,6,7,7a,8,11,11a,12,13,14,14a-tetradecahydro-5,7,12,14-tetraoxo-6,13-[1',2']benzenopentacene-2,3,9,10-tetracarboxylate.

TT12: (4a*S**,5*S**,7*R**,12*S**,14*R**,7a*R**,11a*S**,14a*R**)-Tetramethyl 1,4,4a,5,6,7,7a,8,11,11a,12,13,14,14a-tetradecahydro-5,7,12,14-tetrahydroxy-6,13-[1',2']benzenopentacene-2,3,9,10-tetracarboxylate.

TT15: 2-Hydroxy-3-(methylamino)-9,10-dihydro-9,10-[1',2']benzenoanthracene-1,4,5,8-tetraone.

TT16: 2-Bromo-3-methoxy-6-(dimethylamino)-9,10-dihydro-9,10-[1',2']benzenoanthracene-1,4,5,8-tetraone.

TT17: 2-Bromo-3-[2-(ethoxycarbonyl)ethylamino]-9,10-dihydro-9,10-[1',2']benzenoanthracene-1,4,5,8-tetraone

TT18: 2-hydroxy-3-[2-(ethoxycarbonyl)ethylamino]-9,10-dihydro-9,10-[1',2']benzenoanthracene-1,4,5,8-tetraone

TT20: 2-Hydroxy-3-[(S)-5-(hydroxycarbonyl)-5-aminopentylamino]-9,10-dihydro-9,10-[1',2']benzenoanthracene-1,4,5,8-tetraone

EXAMPLES

The following non-limiting examples are intended to aid in understanding the invention.

In contrast to their inactive parent compound triptycene (code name TT0), several triptycene (TT) analogs were shown to prevent L1210 leukemic cells from synthesizing macromolecules and growing *in vitro*. TT2 blocks the cellular transport of both purine and pyrimidine nucleosides (IC_{50} : $6\mu M$) over a 30-sec period in vitro, in contrast to the quinone antitumor drug daunomycin. The concentration-dependent induction of DNA cleavage at 24 h and internucleosomal DNA fragmentation at 48-72 h by TT2 peaks at $4-10\mu M$ but disappears at $25\mu M$. TT2 induction of DNA cleavage is inhibited by actinomycin D, cycloheximide and the caspase inhibitor 2-VAD-fmk, suggesting that TT2 activates endonucleases and triggers apoptosis.

The antitumor activity of TT2 has been compared to that of daunomycin (DAU), a clinically valuable anthracycline antibiotic which is structurally different from TT2 but also contains a quinone moiety. TT2 inhibits the proliferation (IC_{50} : 300 nM at day 2 and 150 nM at day 4) and viability (IC_{50} : 250 nM at day 2 and 100 nM at day 4) of L1210 cells to the same maximal degree than DAU, suggesting that the cytostatic and cytotoxic activities of TT2 are a combination of drug concentration and duration of drug exposure. Since TT2 does not increase the mitotic index of L1210 cells at 24h like vincristine, it is unlikely to be an antimitotic drug that disrupts microtubule dynamics. Like DAU, a 1.5- to 3-h pretreatment with TT2 is sufficient to inhibit the rates of DNA, RNA and protein syntheses determined over 30- to 60-min periods of pulse-labeling in L1210 cells *in vitro* (IC_{50} : $6\mu M$). In contrast to DAU, which is inactive, a 15-min pretreatment with TT2 has the advantage of also inhibiting the cellular transport of nucleosides occurring over a 30-sec period *in vitro* (IC_{50} : $6\mu M$), suggesting that TT2 prevents the incorporation of 3H -thymidine into DNA because it rapidly blocks the uptake of 3H -thymidine by the tumor cells. After 24h, TT2 induces as much DNA cleavage as camptothecin and DAU, two anticancer drugs producing DNA-strand breaks and known to respectively inhibit DNA

topoisomerase I and II activities. Interestingly, the abilities of TT2 to block nucleoside transport, inhibit DNA synthesis and induce DNA fragmentation are irreversible upon drug removal, suggesting that this compound may rapidly interact with various molecular targets in cell membranes and nuclei to disrupt the functions of nucleoside transporters and nucleic acids and trigger long-lasting antitumor effects which persist after cessation of drug treatment. Because inhibition of nucleoside transport is highly unusual among DNA-damaging drugs, the use of bifunctional TTs with antileukemic activity in the nM range *in vitro* provides a considerable advantage in polychemotherapy to potentiate the action of antimetabolites and sensitize multidrug-resistant tumor cells.

Materials and methods

Cell culture and drug treatments

TT0 was purchased from Aldrich (Milwaukee, WI). All solutions of synthetic TTs and tricyclic pyrone H10, vincristine (VCR; a gift from Lilly Research Laboratories, Indianapolis, IN) and camptothecin (CPT; from Sigma Chemical Co., St Louis, MO) were dissolved and diluted in dimethyl sulfoxide (DMSO), whereas daunomycin (DAU) (from Sigma) solutions were prepared in 0.1 M potassium phosphate buffer, pH 7.4, containing 0.9% NaCl. Suspension cultures of murine L1210 lymphocytic leukemia cells (American Type Culture Collection, Rockville, MD) were maintained in continuous exponential growth by twice-a-week passage in RPMI 1640 medium supplemented with 7.5% fortified bovine calf serum (FCS; HyClone Laboratories, Logan, UT) and penicillin (100 IU/ml)-streptomycin (100 µg/ml), and incubated in the presence or absence of drugs at 37°C in a humidified atmosphere containing 5% CO₂. Since drugs were supplemented to the culture medium in 1 µl aliquots, the concentration of DMSO in the final incubation volume (0.5 ml) never exceeded 0.2% and did not affect the rates of macromolecule syntheses and growth in L1210 cells. Control cells incubated in the absence of drugs were similarly treated with vehicle only and, in every experiment, all incubates received the same volume of solvent. For drug removal, incubates were spun at 200 x g for 10 min, drug-containing supernatants were discarded, and intact cells were washed thrice with 1 ml of the above RPMI 1640 culture medium and resuspended in 0.5 ml of fresh medium for further incubation.

Suspension cultures of WT, drug-sensitive, human HL-60-S promyelocytic leukemia cells, were obtained from American Type Culture Collection (Manassas, VA), maintained in continuous exponential growth by twice-a-week passage in RPMI 1640 medium supplemented with 8.25% fortified bovine calf serum (FCS; Hyclone Laboratories, Logan, UT) and penicillin

(100 IU/ml)-streptomycin (100 μ g/ml), and incubated in the presence or absence of drugs at 37°C in a humidified atmosphere containing 5% CO₂. The MDR HL-60-RV and HL-60-R8 cells were developed in Melvin S. Center's laboratory (Kansas State University) and similarly maintained in RPMI 1640 medium in the absence of drugs (Marsh W, et al, Cancer Res 46: 4053-4057, 1986; McGrath T et al, and Center MS: Biochem Biophys Res Commun 145: 1171-1176, 1987; Marsh W and Center MS: Cancer Res 47: 5080-5086, 1987; McGrath T, et al, Biochem Pharmacol 38: 3611-3619, 1989; Marquardt D, et al, Cancer Res 50: 1426-1430, 1990; Marquardt D and Center MS: J Natl Cancer Inst 83: 1089-1109, 1991; Krishnamachary N and Center MS; Cancer Res 53: 3658-3661, 1993). Every 4 weeks, these HL-60-RV and HL-60-R8 sublines were exposed to 41 nM DAU for 48 h to stabilize their MDR phenotype. This concentration of DAU, which is not cytotoxic to MDR HL-60 sublines, was removed from the culture medium at least 48 h before experimentation. Since drugs were supplemented to the culture medium in 1 μ l aliquots, the concentration of vehicle in the final incubation volume (0.5ml) did not affect basal activity levels in control tumor cells incubated in the absence of drugs. RFs were determined by dividing the IC₅₀ of the MDR cells by that of the sensitive WT parent cell line.

Cell proliferation assay

For tumor cell growth, L1210 cells were resuspended in fresh FCS-containing RPMI 1640 medium, plated at an initial density of 1×10^4 cells/0.5 ml, and incubated in 48-well Costar cell culture plates (Costar, Cambridge, MA). Except when otherwise specified, cells were grown for 4 days in the presence or absence of drugs and their density was monitored every 24h using a Coulter counter (Coulter Electronics, Luton Beds, UK).

HL-60-S, HL-60-RV and HL-60-R8 cells were resuspended in fresh FCS-containing RPMI 1640 medium, seeded in triplicate at an initial density of 1.5×10^4 cells/0.5 ml and incubated at 37°C in 48-well Costar cell culture plates (Costar, Cambridge, MA). Tumor cells were grown for 4 days in the presence or absence (control) of drugs and their density was monitored every 24 h using a Z1 dual threshold Coulter counter (Beckman Coulter, Miami, FL).

Cell viability assay

L1210, HL-60-S, HL-60-RV and HL-60-R8 cells suspended in FCS-containing RPMI 1640 medium were grown in 48-well Costar cell culture plates for up to 4 days in the presence or absence of drugs to evaluate drug cytotoxicity. Decreasing concentrations of cells, such as 1×10^5 and 1.11×10^4 cells/0.5 ml/well, were initially plated at time 0 in order to collect control

samples with approximately equal cell densities after 2 and 4 days in culture, respectively. The viability of TT-treated cells was assessed from their ability to bioreduce the 3-(4,5-dimethylthiazol-2-yl)-5-(3-carboxymethoxyphenyl)-2-(4-sulfophenyl)-2H-tetrazolium (MTS) reagent (Promega, Madison, WI) in the presence of phenazine methosulfate (PMS; Sigma) into a water-soluble formazan product which absorbs at 490 nm. At the appropriate time after drug treatment, cell samples (about $10^6/0.5$ ml/well for controls) were further incubated at 37°C for 3h in the dark in the presence of 0.1 ml of MTS:PMS (2:0.1) reagent and their relative cell viability was estimated by recording the Abs. at 490 nm, using a Cambridge model 750 automatic microplate reader (Packard, Downers Grove, IL). Blank values for culture medium supplemented with MTS:PMS reagent in the absence of cells were subtracted from the results.

Macromolecule synthesis

For nucleic acid and protein syntheses, L1210 cells were resuspended in fresh FCS-containing RPMI 1640 medium at a density of about $1.1-1.6 \times 10^6$ cells/0.5 ml. To estimate the rate of DNA synthesis, HL-60-S, HL-60-RV and HL-60-R8 cells were resuspended in fresh FCS-containing RPMI 1640 medium at a density of 5×10^5 cells/0.5 ml. Except when otherwise specified, the cells were incubated at 37°C for 90 min in the presence or absence of drugs and then pulse-labeled for an additional 30 min with 1 μ Ci of [methyl- 3 H]thymidine (50 Ci/mmol; Amersham Corp., Arlington Heights, IL) to estimate the rate of DNA synthesis. For RNA and protein syntheses, cells were incubated at 37°C for 3h in the presence or absence of drugs and then pulse-labeled for an additional 1h with 2 μ Ci of [5,6- 3 H]uridine (46 Ci/mmol; ICN Biomedicals, Irvine, CA) or 2.5 μ Ci of [4,5- 3 H]L-leucine (100 Ci/mmol; Moravsek Biochemicals, Brea, CA), respectively. The incubations were terminated by the addition of 0.5 ml of 10% trichloroacetic acid (TCA). After holding on ice for 15 min, the acid-insoluble material was recovered over Whatman GF/A glass microfibre filters and washed thrice with 2 ml of 5% TCA and twice with 2 ml of 100% EtOH. After drying the filters, the radioactivity bound to the acid-precipitable material was determined by liquid scintillation counting (LSC) in 10 ml of Bio-Safe NA (Research Products International Corp., Mount Prospect, IL).

Nucleoside transport

L1210 cells (about 1.5×10^6 cells/0.5 ml) were preincubated for 15 min at 37°C in the presence or absence of TTs and then exposed to 1 μ Ci of 3 H-thymidine for only 30 sec to assess the cellular uptake of nucleoside over such very short period of time. HL-60-S, HL-60-RV and HL-60-R8 cells (1.2×10^6 cells/0.5 ml) were preincubated for 15 min at 37°C in the presence or absence (control) of drugs and then exposed to 1 μ Ci of [2,8- 3 H]adenosine (30 Ci/mmol;

American Radiolabeled Chemicals, St. Louis, MO) or [methyl-³H]thymidine (50 Ci/mmol; Amersham, Arlington Heights, IL) for only 30 s to, respectively, assess the cellular uptake of purine and pyrimidine nucleosides over such very short period of time. Reactions were diluted with 2 ml of ice-cold Ca²⁺/Mg²⁺-free Dulbecco's phosphate buffered saline (PBS) and the unincorporated radiolabel was removed by centrifugation at 200 x g for 10 min. After washing thrice with 2 ml of ice-cold PBS, intact cell pellets were harvested by centrifugation and incubated for 30 min in 1 ml of hypotonic lysis buffer (HLB) containing 10 mM Tris-HCl, pH 8.0, 1 mM EDTA and 0.2% Triton X-100. Cell lysates were mixed with 9 ml of Bio-Safe II (Research Products International) and counted to estimate the cellular uptake of ³H-thymidine. Drug inhibition was expressed as % of ³H-thymidine or ³H-adenosine transported into vehicle-treated control cells over similar 30-sec period.

Mitotic index

L1210 cells (about 1 x 10⁶/0.5 ml of FCS-containing RPMI 1640 medium) were incubated in triplicate for 24h at 37°C in the presence or absence of TTs or known antimitotic drugs, and collected by centrifugation at 200 x g for 10 min to determine their mitotic index. For hypotonic treatment, cells were resuspended in 1 ml of 75 mM KCl for 20 min at 4°C. After fixation in 1 ml of MeOH:acetic acid (3:1), the final cell pellets were collected by centrifugation, resuspended in 75 µl of MeOH:acetic acid (3:1), dispensed onto glass slides, air dried, and stained by spreading 40 µl of 0.1% crystal violet under a coverslip. The % of cells in mitosis was determined microscopically by counting 500 cells/slides. The mitotic index was calculated as the % of mitotic cells in drug-treated cultures divided by the % of mitotic cells in non-treated controls.

DNA cleavage

Drug-induced DNA cleavage was determined by intact chromatin precipitation, using L1210, HL-60-S, HL-60-RV and HL-60-R8 cells which were prelabeled with 1 µCi of ³H-thymidine for 2h at 37°C, washed with 3 x 1 ml of ice-cold PBS, collected by centrifugation, and resuspended in fresh FCS-containing RPMI 1640 medium at a density of about 1 x 10⁶ cells/0.5 ml for L1210 and 0.5 x 10⁵ cells/0.5ml for HL cells. Except when otherwise specified, such cells containing prelabeled DNA were then exposed for 24h to TTs and drugs known to induce DNA fragmentation. After centrifugation at 200 x g for 10 min to discard the drugs and wash the cells, the intact cell pellets were lysed for 30 min in 0.5 ml of HLB, centrifuged at 12,000 x g for 30 min to collect the supernatants, and resuspended in 0.5 ml of HLB. After another similar centrifugation, the radioactivities in the pooled supernatants (detergent-soluble low molecular

weight DNA fragments) and the pellet (intact chromatin DNA) were determined by LSC: % DNA fragmentation = [cpm in supernatant/cpm in supernatant + pellet] x 100. Before being counted in 6 ml of Bio-Safe NA, the intact pelleted chromatin was incubated for 2 h at 60°C in the presence of 0.6 ml of NCS tissue solubilizer (Amersham).

Results

Drugs

The TT quinones under study were synthesized in the laboratory by an *in-situ* oxidation of substituted dihydroxybenzenes followed by [4+2] cycloaddition with 1,4-dimethoxyanthracene and then oxidation. This method of synthesis is reported below. The chemical structures of the TTs tested for their antileukemic activity *in vitro* are shown in Scheme 1 (serial numbers indicate the order of synthesis).

Inhibition of tumor cell growth and viability by TTs

L1210

When tested at 256 nM over a 4-day period, several TTs inhibit the rate of tumor cell growth in relation with their ability to decrease tumor cell viability (Fig. 1). Although most of these drugs are much more active at higher concentrations, at this lower concentration of 256 nM, TT4 and TT12 are inactive and TT11 and TT10 have only marginal and weak inhibitory effects. In contrast, 256 nM TT3, TT5, TT6 and TT8 have moderate cytostatic (32-45% inhibition) and cytotoxic (41-54% inhibition) activities. Under similar conditions, TT1, TT7 and TT9 are more effective and can reduce the number and viability of L1210 cells at 4 days by 55-57% and 61-66%, respectively. TT2 and its C2-brominated derivative TT13 at 256 nM, can inhibit leukemic cell proliferation and viability by 67-68% and 73-75%, respectively (Fig. 1). Figure 2 shows the same experiment for additional TT compounds.

DAU is a clinically valuable anthracycline antibiotic, which also contains a quinone moiety. At concentrations ≥ 640 nM, TT2 inhibits almost totally the proliferation and viability of L1210 cells at day 4 but these maximal cytostatic and cytotoxic activities of TT2 can be mimicked by concentrations of DAU as low as 102 nM and the smallest concentrations of TT2 and DAU that can induce significant antiproliferative and cytotoxic effects after 4 days are 41 and 16 nM, respectively (Figs. 3 & 4). These relative potencies of TT2 and DAU can easily be compared using the full concentration-response curves of Figures 3 and 4, where the striped areas at 100% represent the control levels of L1210 cell growth and viability after 2 and 4 days in culture. The magnitudes of the cytostatic and cytotoxic effects of both TT2 and DAU are

clearly related to the combination of their increasing concentration and duration of action. For instance, 16 nM DAU and 41 nM TT2 are ineffective at day 2 but their antiproliferative (Fig. 3) and cytotoxic (Fig. 4) activities become apparent at day 4. Moreover, 41 nM DAU and 256 nM TT2 are moderately cytostatic (42 and 44% inhibition, respectively) and cytotoxic (32 and 49% inhibition, respectively) at day 2 but respectively decrease L1210 cell proliferation by 77 and 68% and L1210 cell viability by 65 and 79% at day 4 (Figs. 3 & 4). As a result, the antiproliferative activities of DAU and TT2 are respectively characterized by IC_{50} values of 50 and 300 nM at day 2 but 25 and 150 nM at day 4, suggesting that DAU is about 6 times more cytostatic than TT2 in the L1210 tumor system *in vitro* (Fig. 3). Similarly, the inhibitions of tumor cell viability by DAU and TT2 are respectively characterized by IC_{50} values of 70 and 250 nM at day 2 but 30 and 100 nM at day 4, suggesting that DAU is about 3.5 times more cytotoxic than TT2 in this leukemic system *in vitro* (Fig. 4).

HL

The concentration-dependent inhibitions of HL-60-S, HL-60-RV and HL-60-R8 cell proliferation by DAU are respectively characterized by IC_{50} values of 14, 500 and 320 nM at day 2 (Fig. 5A) and 9, 275 and 225 nM at day 4 (Fig. 5B). Such RFs of 22.9-35.7 at day 2 and 25.0-30.6 at day 4 demonstrate that DAU is about 30 times less effective as a cytostatic agent in MDR than in WT HL-60 cells (Fig. 5). In contrast, the concentration-dependent inhibitions of HL-60-S, HL-60-RV and HL-60-R8 cell proliferation by TT2 are respectively characterized by similar IC_{50} values of 400, 500 and 350 nM at day 2 (Fig. 6A) and 300, 260 and 290 nM at day 4 (Fig. 6B). These negligible RFs (0.9-1.3 at day 2 and 0.9-1.0 at day 4) suggest that TT2 retains its effectiveness as a cytostatic agent in MDR cells that have become 30 times less sensitive to the antiproliferative action of DAU (Fig. 6).

Since DAU decreases the viability of HL-60-S, HL-60-RV and HL-60-R8 cells with respective IC_{50} values of 80, 1,900 and 2,500 nM at day 2 (RFs: 23.8-31.3) and 15, 450 and 380 at day 4 (RFs: 25.3-30.0), this drug is about 28 times less potent as a cytotoxic agent in MDR than in WT HL-60 cells (Fig. 7). In contrast, TT2 reduces cell viability as effectively in HL-60-S (IC_{50} : 1,300 and 230 nM at days 2 and 4) as in HL-60-RV (IC_{50} : 1,100 and 340 nM at days 2 and 4) and HL-60-R8 (IC_{50} : 1,600 and 350 nM at days 2 and 4) cells (Fig. 8). Such negligible RFs (0.9-1.2 at day 2 and 1.5 at day 4) demonstrate that the cytotoxic potency of TT2 persists in MDR tumor sublines that have become 28 times less sensitive to the cytotoxic activity of DAU (Fig. 8).

Inhibition of DNA synthesis and nucleoside transport by TTs

L1210

A 2-h treatment with TT2 is sufficient to inhibit, in a concentration-dependent manner, the rate of DNA synthesis determined over a 30-min period of pulse-labeling in L1210 cells *in vitro* (Fig. 9). DNA synthesis is totally inhibited by 25 μ M TT2 but, as compared to DAU which becomes effective against DNA synthesis at 0.256 μ M, concentrations greater than 1.6 μ M must be used to demonstrate the inhibitory effect of TT2 on DNA synthesis. Hence, the concentration-dependent inhibitions of DNA synthesis by DAU (IC₅₀: 1 μ M) and TT2 (IC₅₀: 6 μ M) suggest that, under these experimental conditions, DAU prevents L1210 cells from synthesizing DNA about 6 times more effectively than TT2 (Fig.9).

A critical finding is that, in contrast to DAU which serves as a negative control in the assay, a 15-min treatment with TT2 is sufficient to block, in a concentration-dependent manner, the cellular transport of ³H-thymidine occurring over only 30 sec *in vitro* (Fig. 10). Nucleoside transport is totally inhibited in L1210 cells treated with 25 μ M TT2. But 10 μ M DAU is totally unable to alter the cellular transport of ³H-thymidine (Fig. 10), even though such concentration of DAU inhibits maximally the incorporation of ³H-thymidine into DNA (Fig. 9). Interestingly, the concentration-response curves for the inhibitory effects of TT2 on nucleoside transport (Fig. 10) and DNA synthesis (Fig. 9) are nearly identical and share similar IC₅₀ values of 6 μ M, suggesting that the inhibition of ³H-thymidine incorporation into DNA caused by TT2 at 2 h (Fig. 9) may largely be due to the ability of this TT quinone to immediately block the uptake of ³H-thymidine by the cells (Fig. 10).

Moreover, when compared on an equal 7 μ M concentration basis, the different magnitudes at which various TTs inhibit DNA synthesis at 2h correlate with the different abilities of these compounds to block the cellular transport of nucleosides after 15 min (Fig. 11). Indeed, among all TTs tested at this concentration, TT4 and TT12 alter neither DNA synthesis nor nucleoside transport, whereas TT2 and TT13, which block the most the cellular transport of ³H-thymidine (72-75% inhibition), are also the most potent against the incorporation of ³H-thymidine into DNA (69-71% inhibition) (Fig. 11). Under similar conditions, TT1, TT3, TT7, TT9 and TT11 inhibit nucleoside transport by 44-57% and DNA synthesis by 41-58%, whereas TT5, TT6, TT8 and TT10 are less effective, inhibiting both responses in the 20-35% range only (Fig. 11).

Finally, the abilities of 10 and 25 μ M TT2 to respectively inhibit nucleoside transport by 67 and 93% and DNA synthesis by 69 and 97% are both irreversible upon drug removal (Fig. 12), suggesting that, after a 15- to 60-min pretreatment, the presence of TT quinone in the culture medium is no longer required to continually block the cellular transport of 3 H-thymidine and thereby prevent its incorporation into DNA. For the sake of comparison, the inhibitions of nucleoside transport and DNA synthesis caused by the novel tricyclic pyrone analog H10, which are fully reversible upon drug removal, are demonstrated as a positive control under similar experimental conditions in the L1210 tumor assay system (Fig. 12).

As shown in Figure 13, TT16 is as potent as TT2 on both DNA synthesis and the cellular transport of adenosine and thymidine.

HL

The concentration-dependent inhibitions of DNA synthesis by DAU in HL-60-S (IC_{50} : 0.21 μ M), HL-60-RV (IC_{50} : 2.5 μ M) and HL-60-R8 (IC_{50} : 1.7 μ M) cells are characterized by RF values of 8.1-11.9 (Fig. 14A). In contrast, TT2 inhibits the rate of incorporation of 3 H-thymidine into DNA as effectively in HL-60-S (IC_{50} : 1.8 μ M) as in HL-60-RV (IC_{50} : 2.1 μ M) and HL-60-R8 (IC_{50} : 2.1 μ M) cells and there are no significant RFs (1.2), indicating that TT2 maintains its ability to inhibit DNA synthesis in MDR tumor sublines that have become 10 times less sensitive to the action of DAU (Fig. 14B).

DAU, which fails to significantly inhibit nucleoside transport in WT HL-60-S parental cells, is also unable to do so in their two MDR sublines (Fig. 15). In contrast, TT2 is a quinone antitumor drug that has the advantage of rapidly blocking, in a concentration-dependent manner, the cellular transport of both purine and pyrimidine nucleosides in HL-60-S cells (Fig. 15). Moreover, the ability of TT2 to prevent the cellular transport of 3 H-adenosine and 3 H-thymidine in WT HL-60-S cells (IC_{50} : 3.3 and 2.0 μ M, respectively) fully persists (RFs: 1.0-1.2) in both the MDR HL-60-RV (IC_{50} : 3.2 and 2.0 μ M, respectively) and HL-60-R8 (IC_{50} : 3.7 and 2.3 μ M, respectively) cell lines (Fig. 15).

Inhibition of RNA and protein syntheses by TTs

Besides DNA synthesis, a 3-h treatment with TT2 can also inhibit, in a concentration-dependent manner, the rates of RNA (Fig. 16) and protein (Fig. 17) syntheses determined over 60-min periods of pulse-labeling in L1210 cells *in vitro*. The concentration-response curves for the inhibitions of RNA (Fig. 16) and protein (Fig. 17) syntheses by TT2 are nearly identical to

that for the inhibition of DNA synthesis (Fig. 8): concentrations of TT2 greater than 1.6 μ M must be used to demonstrate effectiveness, total inhibition is achieved at 25 μ M, and the IC₅₀ value in all cases is about 6 μ M. Under similar conditions, DAU inhibits the respective syntheses of RNA (IC₅₀: 0.8 μ M) and protein (IC₅₀: 2 μ M) about 7.5 and 3 times more effectively than TT2 (Figs. 16 & 17).

Effects of TTs on the mitotic index

Control populations of L1210 cells cultured for 24h in the absence of drugs contain only 1.86% of mitotic cells (Fig. 18). In relation with their ability to block tubulin polymerization and cell cycle progression in M-phase, 24-h treatments with VCR and the tricyclic pyrone H10 respectively produce 16- and 7-fold increases in the mitotic index (Fig. 18). Such known microtubule de-stabilizing anticancer drugs, therefore, serve as positive controls in this antimitotic assay. In contrast, none of the concentrations of TT2 tested, even those in the μ M range that are highly cytostatic and cytotoxic at 24h, are able to raise the mitotic index of L1210 cells (Fig. 18), suggesting that TT quinones are unlikely to be antimitotic drugs that disrupt microtubule dynamics to trigger their anticancer activity. In relation with its known ability to first accumulate cells in G₂ and then inhibit cell cycle traverse as its concentration increases, DAU actually decreases the % of mitotic cells by 74-91% (Fig. 18). Since the highest concentrations of TT2 tested significantly decrease the % of mitotic cells by 38-48% (Fig. 18), TTs might also prevent tumor cell cycle progression to mitosis.

Induction of DNA cleavage by TTs

L1210

L1210 cells containing ³H-thymidine-prelabeled DNA were used to quantitatively determine whether TTs could induce DNA fragmentation over a 24-h period *in vitro*. CPT and DAU, two anticancer drugs known to induce DNA-strand breaks by respectively inhibiting Topo I and II activities, are used as positive controls in this DNA fragmentation assay (Fig. 19). As reported before with anthracycline quinone antibiotics, the concentration-dependent induction of DNA cleavage caused by 24-h DAU treatments is biphasic, peaking at 45% in response to 1.6 μ M DAU but declining back to control level (5%) at higher concentrations of DAU (Fig. 19). In contrast, the concentration-dependent increase of DNA cleavage produced by 24-h CPT treatments reaches 57% in response to 1.6 μ M CPT but remains at a plateau of maximal stimulation (60-70%) at higher concentrations of CPT (Fig. 19). After 24 h, the maximal levels of DNA cleavage caused by 10 μ M TT1 (45%) and TT2 (61%) respectively match those induced by 1.6 μ M DAU and CPT (Fig. 19). Although concentrations of TTs higher than those of DAU

are required to induce such peak of DNA cleavage, the shape of the concentration-response curves for the ability of TT1 and TT2 to break DNA resembles more that of DAU than that of CPT (Fig. 19), suggesting that the DNA-damaging effects of TT quinones and DAU might share some similarity.

Overall, the different TTs compared at 7 μ M induce various levels of DNA cleavage at 24 h (Fig. 20) in relation with their respective cytostatic and cytotoxic activities when tested at 256 nM for 4 days (Fig. 1). Indeed, TT2 and TT13, which are the most potent against L1210 cell proliferation and viability (Fig. 1), again induce the most DNA fragmentation (54-57%) after 24h (Fig. 20), whereas TT4 and TT12, which have no antileukemic activity in the growth and viability assays (Fig. 1), also fail to significantly raise the level of DNA cleavage (4-6%) over control (5%) (Fig. 20). TT1, TT7 and TT9, which have good antileukemic activities (Fig. 1), also produce substantial levels of DNA fragmentation (31-40%) (Fig. 20). Moreover, the extent of DNA cleavage is only 14-18% in L1210 cells treated with TT3, TT5, TT6, TT8 or TT10 and the ability of TT11 to break DNA is minimal (7%) (Fig. 20), in relation with the weak and marginal antiproliferative and cytotoxic effects of these compounds in the L1210 system *in vitro* (Fig. 1). Figure 21 shows the same experiment as in Figure 20 with additional analogs.

Finally, L1210 cells treated for only 3 and 8 h with 10 μ M TT1 or TT2 have the same level of DNA fragmentation at 24h (36-45% or 58-67%, respectively) than if they are exposed for the whole 24-h incubation period to TT1 or TT2 (40 or 63%, respectively) (Fig. 22). Under similar conditions, 1.6 μ M DAU and CPT also rapidly trigger molecular events, which are irreversible and produce identical levels of DNA fragmentation at 24h (33-41% and 51-58%, respectively), whether or not those drugs are maintained in the culture medium after 3 or 8h (Fig. 20). It should be noted that no significant elevation of DNA cleavage is detectable in this assay after 3 and 8h of drug exposure and that the increases of DNA fragmentation caused by TTs, DAU and CPT only appear after 12h to reach a maximal level at 24h (data not shown), suggesting that the irreversible events triggered by those drugs during the initial 3h still require a substantial period of time, irrespective of the continual presence or absence of drugs, to fully induce DNA fragmentation. These results suggest that, like anticancer drugs known to induce DNA-strand breaks, TT quinones interact rapidly with cellular targets to induce long-lasting DNA-damaging effects, which develop and persist after drug removal.

HL

Tumor cells containing ^3H -thymidine prelabeled DNA were used to quantitatively determine whether TT2 would induce DNA fragmentation as effectively in WT as in MDR HL-60 cell lines that have become resistant to the DNA-damaging effects of DAU (Fig. 23). As reported before, the concentration-dependent induction of DNA cleavage caused by DAU is biphasic, peaking 67% above control level (15% DNA fragmentation) in HL-60-S cells treated for 24 h with 1.6 μM DAU but declining back to (18% above control at 4 μM), or even below (11 and 14% below control at 10 and 25 μM , respectively), control level at higher concentrations of DAU (Fig. 23A). Although TT2 is somewhat less potent than DAU, the concentration-dependent induction of DNA cleavage caused by this compound in HL-60-S cells is similarly biphasic, with a peak 37% above the control % of DNA fragmentation at 4 μM , followed by a decline toward (14% above control at 10 μM), or even below (11% below control at 25 μM), the control % of DNA fragmentation at higher concentrations (Fig. 23B). Interestingly, the ability of DAU to trigger maximal or submaximal DNA cleavage in WT cells is totally lost, drastically reduced and/or shifted toward higher concentrations in the MDR cell lines (Fig. 23A). For instance, the 0.04-0.64 μM concentrations of DAU producing submaximal stimulations of DNA cleavage (11-58% above the control level of 15%) in HL-60-S cells are almost totally unable to do so in the HL-60-RV and HL-60-R8 cell lines. Moreover, the magnitude of the peak of maximal DNA cleavage caused by DAU in WT cells is not only reduced by 60 and 79% in HL-60-R8 and HL-60-RV cells, respectively, but it is also shifted from 1.6 to 10 μM in the latter MDR subline (Fig. 23A). As a result, the ability of 10-25 μM DAU to decrease DNA cleavage below control level in WT cells is totally or partially lost in the MDR cell lines. In contrast, the ability of TT2 to induce a peak of DNA cleavage at 4 μM and abolish the basal level of DNA cleavage at 25 μM in WT cells is pretty much unaltered in the two MDR sublines, although the magnitude of maximal stimulation observed in HL-60-S cells is reduced by 35 and 39% in the HL-60-R8 and HL-60-RV cells, respectively (Fig. 23B).

The comparison studies show that the different magnitudes at which equimolar concentrations of various TT analogs inhibit tumor cell proliferation match very well the different cytotoxic activities of these compounds (Fig. 1). Moreover, the different levels of DNA fragmentation induced by the various TT analogs (Fig. 20) match exactly the ranking of these compounds for their cytostatic/cytotoxic effects (Fig. 1), suggesting that the ability of TT analogs to break DNA plays a significant role in their molecular mechanism of antitumor activity. In addition, the various TT analogs inhibit DNA synthesis in relation with their effectiveness against nucleoside transport (Fig. 11), suggesting that they prevent DNA assembly because they block the cellular uptake of DNA precursors. Because of their ability to interact with both

membrane and nuclear targets to block nucleoside transport, inhibit nucleic acid and protein syntheses, cleave DNA, and reduce tumor cell growth and viability in the nM range *in vitro*, these TT analogs represent a novel synthetic class of bifunctional anticancer drugs valuable to develop new means of polychemotherapy.

The cytostatic and cytotoxic effects of each concentrations of TT2 increase with the time in culture (Figs. 3 and 4), suggesting that the effectiveness of TT analogs as inhibitors of tumor cell proliferation and viability *in vitro* is a combination of drug concentration and duration of drug exposure. As a result, TT2 inhibits L1210 tumor cell growth at 2 and 4 days with IC₅₀ values of 300 and 150 nM, respectively, in relation with its ability to reduce L1210 tumor cell viability with IC₅₀ values of 250 nM at day 2 and 100 nM at day 4. The IC₅₀ required to reduce tumor cell viability at day 4 (100 nM) may be lower and perhaps more accurate than that observed for tumor cell growth inhibition at day 4 (150 nM) because the Coulter counter data include all viable and nonviable tumor cells that have previously accumulated and remain in the medium at day 4, irrespective of their present metabolic status and reproductive ability. Moreover, decreased tumor cell viability after several days of drug treatment may be a better predictor of anticancer activity than antiproliferation since growth delay may allow survivors to resume dividing and expand clonally once the drug is catabolized or eliminated and its effect is waning.

Overall, the new TT bisquinone TT2 is a potent antitumor agent, which is active in the nM range and is only 3.5-6 times less cytostatic/cytotoxic after 4 days than equimolar concentrations of the clinically proven anthracycline quinone antibiotic DAU, a very potent anticancer drug used as an arbitrary reference in our L1210 tumor cell studies *in vitro* (Figs. 3 and 4). When IC₅₀ values are compared, TT2 also inhibits the syntheses of DNA, RNA and protein in L1210 cells after 2-3h about 3-6 times less effectively than DAU (Figs. 9, 16 and 17) and concentrations of TT2 about 6 times higher than those of DAU are also required to induce the same peak of DNA cleavage in L1210 cells after 24h (Fig. 19). But in addition to mimicking all the antitumor effects of DAU studied, the data demonstrate that TT2 remarkably blocks the cellular transport of nucleosides, which DAU cannot do (Fig. 10), suggesting that these novel antitumor TT bisquinones may have a more versatile mechanism of action and be advantageous in polychemotherapy to potentiate the anticancer effects of antimetabolites and circumvent multidrug resistance (MDR). For both DAU and TT2 concentrations in the 25-150 nM range are sufficient to inhibit tumor cell growth/viability (Figs. 3 and 4), whereas higher concentrations in the 0.8-10 μ M range must be used to inhibit macromolecule syntheses (Figs. 9, 16 and 17) and

maximally induce DNA fragmentation (Fig. 19). But apparent discrepancies in potencies may be due in part to different experimental conditions and cellular responses to various periods of drug exposure: the rates of nucleic acid and protein syntheses over 30-60 min are inhibited in cells treated for only 2-3h with TT2 or DAU, whereas the level of DNA cleavage and the reduction of tumor cell growth/viability are the results of 1- and 4-day long drug treatments, respectively. It should be noted that no significant DNA fragmentation can be detected within the first 8h of TT2 or DAU treatments and that concentrations of DAU and TT2 respectively greater than 0.25 and 1.6 μ M must be used to induce substantial antiproliferative and cytotoxic effects after only 24h of drug exposure (data not shown).

The fact that, within 24h, TT1 and TT2 can produce as much internucleosomal DNA fragmentation in L1210 cells than the known Topo I and II inhibitors CPT and DAU (Fig. 19) suggests that the ability of TT quinones to produce DNA-strand breaks may play a major role in their mechanism of antitumor activity. Since the shapes of the concentration-response curves for the effects of TT1 and TT2 on DNA cleavage resemble the biphasic response to DAU, which has already been reported and discussed before, it is tempting to speculate that the mechanisms by which TT quinones and DAU induce DNA fragmentation share some similarity and that DNA cleavage is not always necessary for high concentrations of TT quinones and DAU to be cytotoxic. The Topo II-associated DNA lesions occurring in cells exposed to anthracycline quinone antibiotics may facilitate subsequent internucleosomal DNA fragmentation by endogenous nucleases and trigger apoptosis. Since apoptosis is an active and cell cycle phase specific process, which requires the expression of specific genes, the syntheses of new RNA and proteins and the activation of endonuclease enzymes, inhibitors of such mechanisms can prevent DNA fragmentation in anthracycline-treated cells. In spite of their increasing cytotoxicity, the highest concentrations of TT1, TT2 and DAU tested in our study might inhibit RNA and protein syntheses, reduce the level of Topo II targets, inactivate endonucleases and/or arrest cell cycle traverse to such degrees that they actually block the molecular mechanisms required for internucleosomal DNA fragmentation and apoptosis and produce the paradoxical biphasic curve of DNA cleavage shown in Figure 19. Taken together, the irreversibilities of the inhibitions of nucleoside transport/DNA synthesis and of the induction of DNA fragmentation upon TT2 removal suggest that, because TT analogs may rapidly and tightly interact with various membrane and intracellular targets, their presence is soon no longer required in the medium to disrupt the structures/functions of nucleoside transporters, nucleic acids and proteins and to trigger long-lasting antitumor events, which persist after cessation of drug treatment.

Combining drugs, which target different molecules and achieve complementary or synergistic antitumor effects, is an important strategy in cancer chemotherapy. A TT quinone inhibiting nucleoside transport and inducing DNA cleavage might disrupt a wider spectrum of molecular targets in populations of unsynchronized tumor cells than another drug affecting a single of these events. Nucleoside transporters in mammalian plasma membranes function by equilibrative (facilitated diffusion) or Na⁺-dependent (concentrative) mechanisms. All nucleosides are substrates for the Na⁺-independent transporters but Na⁺-dependent transporters generally accept purine rather than pyrimidine nucleosides as substrates. Dipyridamole (DPR), 6-(4-nitrobenzylmercapto)purine ribonucleoside (NBMPR) and dilazep are standard potent inhibitors of equilibrative nucleoside transport, whereas phloridzin specifically inhibits Na⁺-dependent nucleoside transport. Like TT2, NBMPR and DPR also reduce ³H-thymidine incorporation into DNA. Nucleoside transport inhibitors block equally well the influx and efflux of nucleosides. L1210 cells possess 3 distinct nucleoside transporters: 2 equilibrative transporters sensitive (es) or insensitive (ei) to NBMPR and one Na⁺-dependent transporter (cif) of low sensitivity to NBMPR and DPR.

For nucleotide synthesis, cells use purine and pyrimidine nucleosides generated either through *de novo* synthesis or through the utilization of salvage pathways. MDR is sometimes associated with increases in the number of nucleoside transporters and their rate of transport, resulting in the increased uptake of adenosine. By blocking the rescue effect of exogenous nucleosides, NBMPR, DPR and dilazep may potentiate or prolong the antitumor activity of antimetabolites which inhibit the *de novo* pathway for nucleoside synthesis. The clinical effectiveness of ADR and DAU is limited by their cumulative cardiotoxicity and ability to induce MDR. The multifactorial mechanisms of MDR to anthracycline quinone antibiotics may include altered expressions of P-glycoprotein (P-gp), Topo II and multidrug resistance-associated protein, increased DNA repair and glutathione-dependent detoxifying enzyme activities, and alterations in cell cycle progression and apoptotic pathways. DPR circumvents ADR resistance and its analog, BIBW 22, is a bifunctional modulator which reverses the MDR phenotype by interfering with both P-gp and nucleoside transport in MDR cells. As bifunctional inhibitors of nucleoside transport and inducers of DNA cleavage, TT quinones might be valuable in polychemotherapy to potentiate the antitumor activity of methotrexate and 5-fluorouracil and sensitize MDR tumor cells that have become unresponsive to the cytotoxicity of other conventional DNA-damaging anticancer agents.

TT2 is also cytostatic (IC_{50} : 300 nM) and cytotoxic (IC_{50} : 230 nM) to WT human HL-60-S leukemic cells in the present study. Another attractive feature of TT2 identified in the present study is the finding that this new quinone antitumor drug retains its effectiveness in two MDR HL-60-RV and HL-60-R8 sublines that have developed different mechanisms of resistance to DAU and, therefore, might not be recognized by either the P-gp or the MRP.

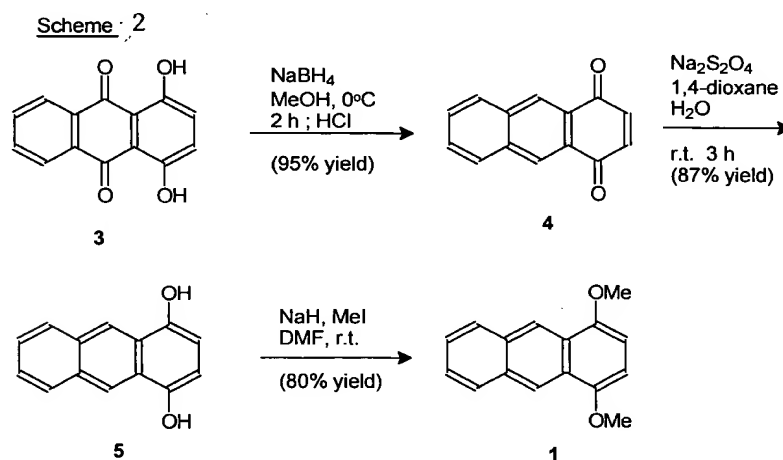
The magnitude at which TT2 inhibits HL-60-S, HL-60-RV and HL-60-R8 cell proliferation (IC_{50} values: 260-300 nM) matches its ability to decrease cell viability (IC_{50} values: 230-350 nM) in the same WT and MDR tumor cell lines at day 4. Moreover, the cytostatic and cytotoxic effects of each concentrations of TT2 obviously increase with the time in culture at days 2 and 4, suggesting that the effectiveness of TT2 as an inhibitor of tumor cell proliferation and viability is a combination of drug concentration and duration of drug exposure. Decreased WT and MDR tumor cell viability after TT2 treatment *in vitro* may be a reliable predictor of anticancer activity *in vivo*. However, it should be noted that DAU is about 3.5-6 times more cytostatic/cytotoxic than TT2 in L1210 cells, even though TT2 appears more potent in the L1210 than in the HL-60-S cells. Hence, TT2 inhibits tumor cell growth, viability and DNA synthesis respectively 33, 15 and 9 times less effectively than DAU in the HL-60-S leukemic system (Figs. 5-8, 10). Moreover, concentrations of TT2 at least 2.5 times greater than those of DAU are required to induce more than 50% of DNA cleavage in HL-60-S cells at 24 h (Fig. 23). Nevertheless, the magnitudes of the inhibitory and stimulatory effects triggered by DAU and somewhat higher concentrations of TT2 are nearly identical in HL-60-S cells. But the critical finding is that the ability of TT2 to inhibit tumor cell growth, viability and DNA synthesis in HL-60-S cells persists unaltered in the HL-60-RV and HL-60-R8 cell lines (RFs: 0.9-1.5) while DAU becomes dramatically less effective against DNA synthesis (RFs: 8.1-11.9), cell proliferation (RFs: 22.9-35.7) and cell viability (RFs: 23.8-31.3) in these MDR HL-60 sublines than in their parental WT counterparts. Finally, TT2 which, in contrast to DAU, has the unique ability to rapidly block the cellular transport of both purine and pyrimidine nucleosides in L1210 (13) and HL-60-S cells, retains such additional advantage in the MDR HL-60 sublines (Fig. 15). These observations substantiate the hypothesis that, because of its bifunctional mechanism of action, TT2 is a quinone antitumor drug which may be more versatile than DAU and able to circumvent MDR.

SYNTHESIS EXAMPLES

Synthesis of **TT2**, **TT13** and Their Analogs:

Although the preparation of 1,4-dimethoxyanthracene (**1**) has been reported (Criswell, et al., J. Org. Chem. 1974, 39, 770-774) by using methylation of quinizarin (**3**) followed by sequential reduction of the diketo function with sodium borohydride in diglyme, the difficulty in the reduction steps prompted us to investigate an easier method in the preparation of **1**.

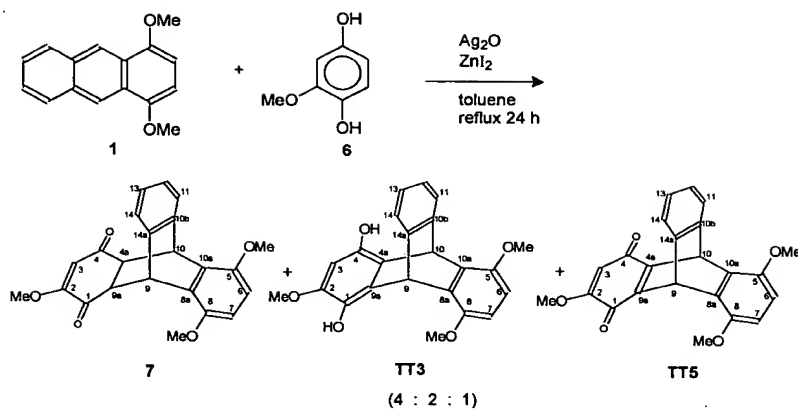
Reduction of **3** with sodium borohydride in MeOH followed by quenching with HCl gave a 95% yield of 1,4-anthraquinone (**4**) (Scheme 2). Reduction of the quinone moiety of **4** with aqueous sodium hydrosulfite in 1,4-dioxane provided 1,4-dihydroxyanthracene (**5**). Methylation of **5** with sodium hydride in DMF at room temperature gave excellent yield of **1**.



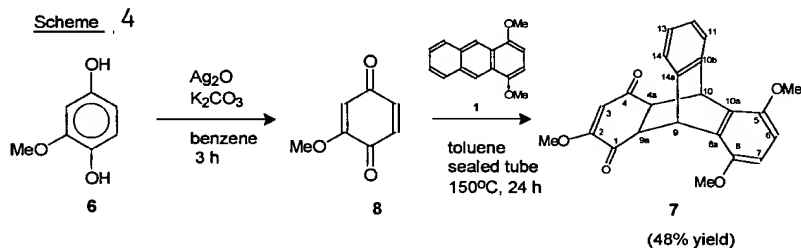
In Situ Oxidation followed by [4+2]-Cycloaddition:

A short syntheses of **TT2**, **TT3**, and **TT5** were discovered. Without the use of quinones as the dienophiles in the Diels-Alder reaction, such as 2-methoxyquinone (**8**), a one-pot oxidation followed by [4+2] cycloaddition of 2-methoxyhydroquinone (**6**) with anthracene **1** was found (Scheme 3). Hence, treatment of 2 equiv of **6**, 1 equiv of anthracene **1**, 2 equiv of silver oxide, and 0.2 equiv of zinc iodide in refluxing toluene for 24 h gave diketone **7** (51% yield; based on reacted **1**), hydroquinone **TT3** (26% yield; based on reacted **1**), quinone **TT5** (13% yield; based on reacted **1**) and 55% recovery of **1**.

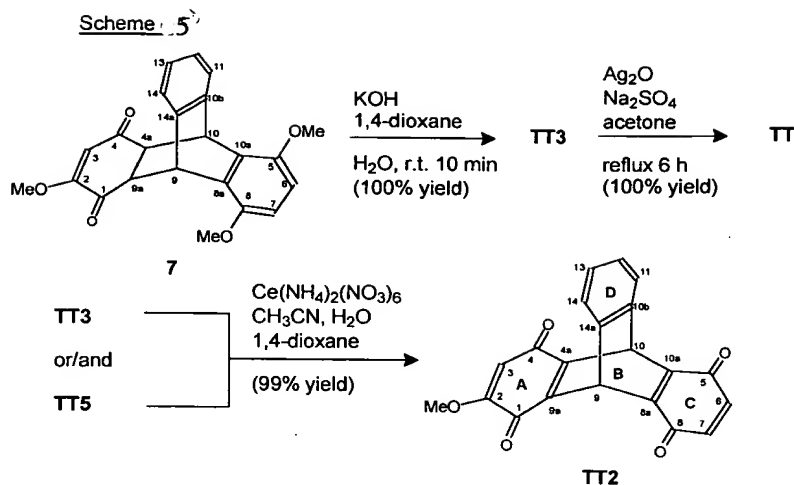
Scheme 3



Alternatively, **7** can be obtained from the Diels-Alder reaction of quinone **8** with anthracene **1** (Scheme 4). Hence, oxidation of **6** with 1.5 equiv of silver oxide and 1.2 equiv of potassium carbonate in benzene at 25 °C for 3 h afforded quantitative yield of quinone **8** which underwent cycloaddition with **1** gave a 48% yield of **7** as a mixture of endo- and exo-isomers (a ratio of 1:1) at C4a and C9a.

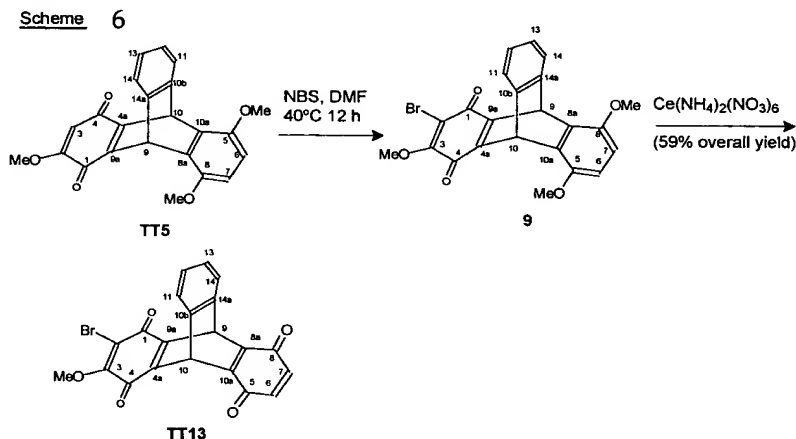


Diketone **7** was converted into hydroquinone **TT3** with 10 equiv of KOH in 1,4-dioxane and water at 25 °C for 1 h in quantitative yield (Scheme 5). Oxidation of **TT3** with 2 equiv of silver oxide and sodium sulfate (anhydrous) in dried acetone under reflux for 6 h gave **TT5** in quantitative yield. Oxidation of **TT5** or **TT3** or a mixture of **TT3** and **TT5** with ceric ammonium nitrate in acetonitrile-1,4-dioxane-water at 25 °C for 12 h gave excellent yields of **TT2**.



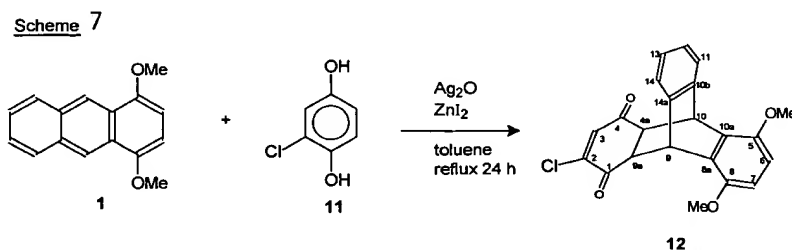
Selective Bromination of 2-Methoxy Triptycene Quinone:

A new bromination reaction was found (Scheme 6). Hence, when **TT5** was treated with *N*-bromosuccinimide (NBS) in DMF at 40 °C for 12 h, a quantitative yield of the C-2 brominated product **9** was obtained. Without purification, compound **9** was directly subjected to the ceric ammonium nitrate oxidation and a 59% yield of **TT13** was achieved.

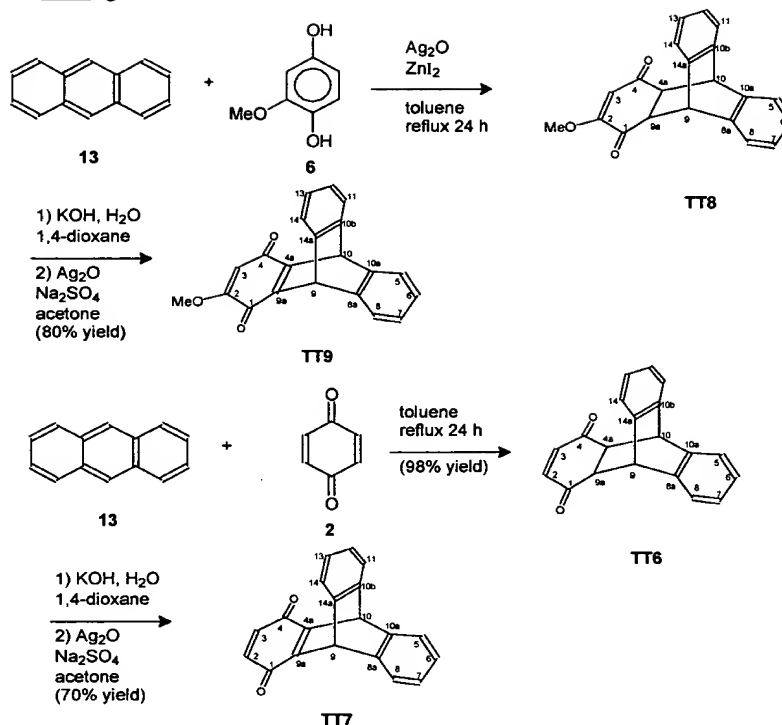


Syntheses of 2-Chloro-4a,9,9a,10-tetrahydro-9,10-[1',2']benzenoanthracene-1,4-dione (**10**), **TT8**, **TT9** and **TT7**:

The *in-situ* oxidation and cyclization reaction is applicable to other hydroquinone substrates. For instance, 2-chlorohydroquinone (**11**), under similar reaction conditions, reacted with 1,4-dimethoxyanthracene (**1**) in the presence of silver oxide and zinc iodide under refluxing toluene to give a 73% yield of the chloro derivative **12** (Scheme 7).



Scheme 8

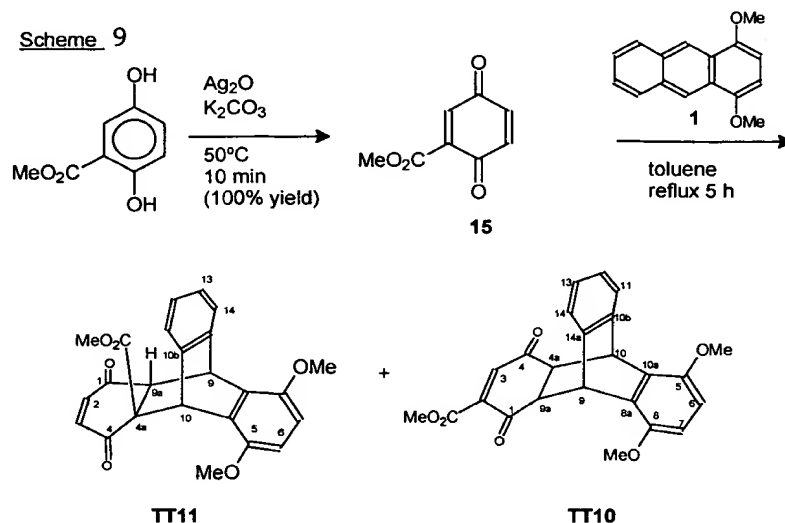


Similarly, treatment of 2-methoxyhydroquinone (**6**) with anthracene (**13**) and silver oxide and zinc iodide gave a 56% yield of the 2-methoxy derivative **TT8** (Scheme 8). Basic isomerization of ketone **14** with 3 equiv of KOH in 1,4-dioxane and water at 25 °C for 2 h followed by oxidation with silver oxide and sodium sulfate (anhydrous) in dried acetone at 25 °C for 3 h to give an 80% yield of quinone **TT9**. Quinone **TT7** was prepared by the Diels-Alder reaction of benzoquinone (**2**) with anthracene **13** to give adduct **TT6** (98% yield) which under similar isomerization and oxidation as those for the conversion of **TT8** into **TT9** gave **TT7** (70% yield for the last two steps).

Syntheses of **TT10** and **TT11**:

Contrary to the above results, when methyl gentisate (**14**) was used in the *in-situ* oxidation followed by [4+2] cycloaddition reaction, only small amount of a mixture of products was isolated. Compound **14** was then oxidized first with 2 equiv of silver oxide and 1.2 equiv

of potassium carbonate in benzene at 50 °C for 10 min under dark gave a quantitative yield of the corresponding quinone **16** (Scheme 9). Diels-Alder reaction of **15** and anthracene **1** at 70 °C for 14 h and then under reflux for 5 h gave 58% yield of **TT11** (as a mixture of endo- and exo-isomers) and 37% yield of **TT10**.



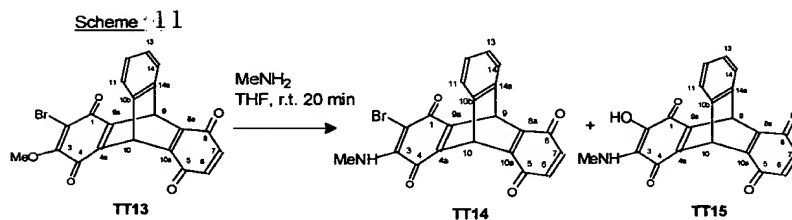
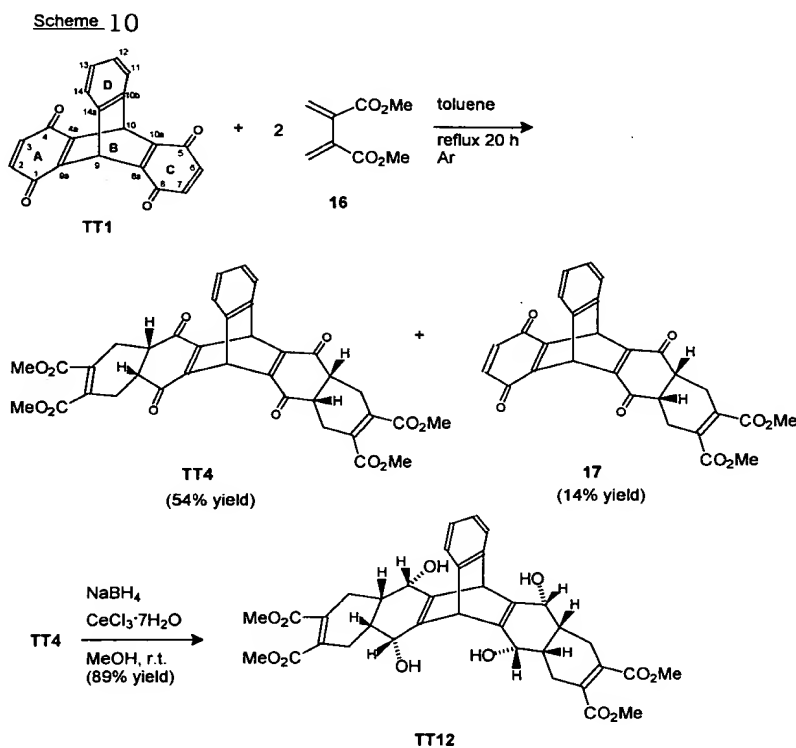
Syntheses of **TT4** and **TT12**:

The triptycene bis-quinones such as **TT1** could be treated with dienes to produce various substituted triptycene analogs such as **TT4** and **TT12** (Scheme 10). Hence, treatment of **TT1** with 2.2 equiv of dimethyl butadiene-2,3-dicarboxylate (**16**) (Hamon, et al., J. Chem. Soc. Chem. Commun. 1981, 873-4) in toluene under reflux for 20 h gave a 54% yield of **TT4** and 14% yield of monoadduct **17**. The stereochemistry of **TT4** was firmly established by a single-crystal X-ray analysis. Reduction of **TT4** with 10 equiv each of sodium borohydride and cerium trichloride heptahydrate in MeOH at room temperature for 12 h afforded an 89% yield of **TT12**. The ^1H and ^{13}C NMR spectrum of **TT12** indicated a single stereoisomer. It is anticipated that the hydride (sodium borohydride) should attack the carbonyl group from the exo face (b-face) and the stereochemistry is therefore assumed.

Syntheses of **TT14**, **TT15**, and **TT16**:

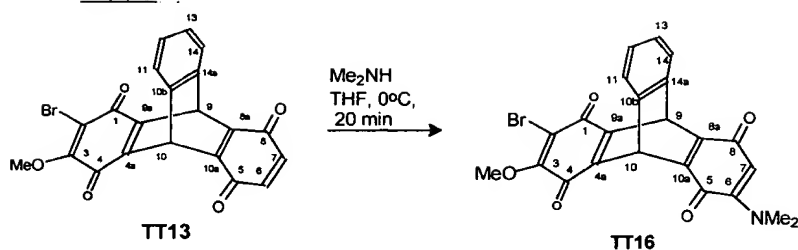
Bromomethoxyquinone **TT13** can be converted into methylaminoquinone **TT14** in 25% yield along with a 29% yield of **TT15** by the treatment with methylamine in THF at room temperature for 20 min (Scheme 11). This reaction is unusual in that the nucleophile, methylamine, displaces the methoxy group of **TT13** instead of the bromine. Again,

unexpectedly, when **TT13** was treated with dimethylamine in THF at 0°C, **TT16** was isolated as the only identifiable product (Scheme 12). The regiochemistry of the dimethylamino group of **TT16** is tentatively assigned.

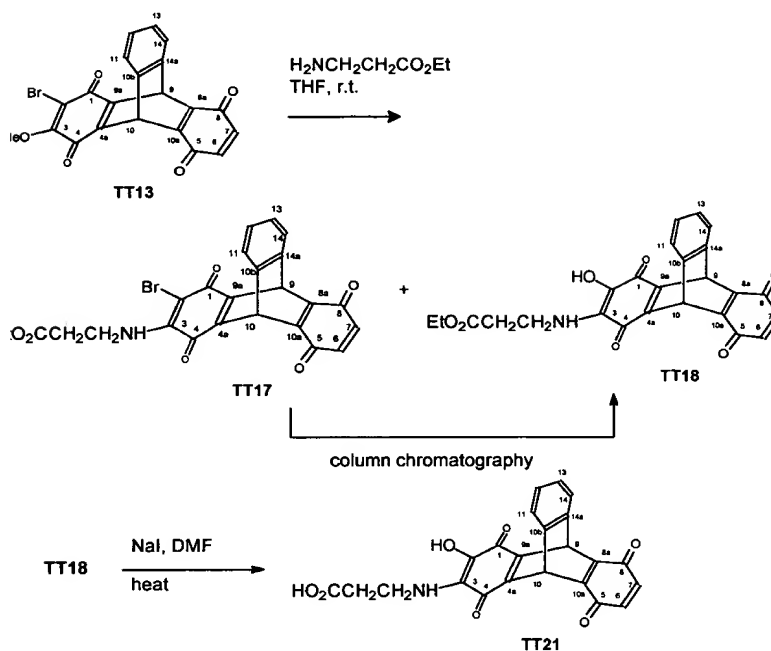


Similar to the addition reaction of methylamine with **TT13**, other primary amines, such as ethyl b-alanine and L-lysine also add to **TT13** to provide the corresponding amino acid adducts **TT17** ~ **TT20** (Scheme 13). Hence, treatment of **TT13** with ethyl b-alanine (derived from ethyl b-alanine hydrochloride with 1 equiv of sodium hydride in THF) in THF and DMF at 25°C to give a mixture of **TT17** and **TT18** (based on the proton NMR spectrum of the crude product). Column chromatographic separation on silica gel gave **TT18** in 69% yield. The ethyl ester function of **TT18** can be removed by treatment with sodium iodide in DMF with heat to give amino acid **TT21** which is water soluble.

Scheme 12



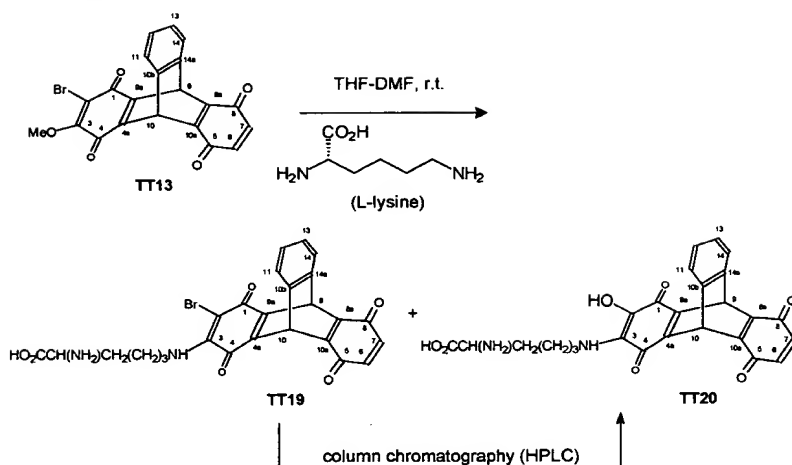
Scheme 13



Under similar reaction conditions, **TT13** was treated with L-lysine (derived from L-lysine monohydrochloride and 1 equiv of NaH in THF) in THF and DMF to give adducts **TT19** and **TT20** which were purified on HPLC to give **TT20** (**TT19** most likely hydrolyzes on the column to give **TT20**).

Hence, a number of water-soluble drugs can be obtained by adding amino acids or amino sugars onto **TT13**, by reactions that are known in the art.

Scheme 13 continued



Preparation of 1,4-anthracenedione (4):

To a dried 100 mL-round-bottom flask equipped with stirring bar and serum cap, was added 1 g (MW 240.21; 0.0042 mol) of quinizarin (3). The compound was dried under vacuum, and then maintained under argon. Methanol (distilled over Mg), 20 mL, was added via syringe. The solution was stirred to dissolve the quinizarin and then cooled over an ice-water bath equipped with a thermometer. To it, 0.638 g (MW 38; 0.0168 mol) of sodium borohydride was added. The resulting mixture was stirred at 0 °C for 1 hour. The progress of the reaction was monitored by TLC (using hexanes: ether 1:1 as the eluant). The reaction was quenched by adding carefully 11 mL of 6 N HCl dropwise at 0 °C over a period of 10 minutes. Orange solids precipitate. The solids were collected by filtration over a fritted filter funnel, and washed several times with distilled water to remove the acid. The solid was then dried under vacuum and recrystallize from acetone-ether to give 0.83 g (95% yield) of yellow crystals, mp 204 - 206 °C.

¹H NMR (CDCl₃) δ ppm: 7.1 (s, 2 H, C 2,3 Hs), 7.7 (dd, *J* = 6.4, 3.2 Hz, 2 H, C 6,7 Hs), 8.1 (dd, *J* = 6.4, 3.2 Hz, 2 H, C 5,8 Hs), 8.6 (s, 2 H, C 9,10 Hs); ¹³C NMR (CDCl₃) δ ppm: 184.68 (s, C=O), 140.06 (d), 134.83 (s), 130.22 (d), 129.59 (d), 128.87 (d), 128.37 (s).

Preparation of 1,4-Dihydroxyanthracene (5):

To a 250 mL-round-bottom flask, was added 2 g (MW 208; 0.0096 mol) of 1,4-anthracenedione (4). To it a solution of 6.6816 g (MW 174; 0.0384 mol) of sodium hydrosulfite in 50 mL of distilled water and 50 mL of 1,4-dioxane were added. The resulting mixture was

stirred at room temperature for 2-3 hours. The reaction progress is checked with TLC (using CH₂Cl₂: hexanes: ether 1:1:0.2 as the eluant). The mixture was transferred to a 500-mL beaker slowly, cooled over an ice-water bath, and added 100 mL of distilled water. Dark green solids precipitated. The beaker was covered with aluminum foil, and placed in the refrigerator for 1 h. The solid was filtered through a glass fritted funnel and the product was washed with 30 mL of distilled water twice. The solid product was dried under vacuum to give 1.754 g (87% yield), mp 167-169 °C; ¹H NMR (CDCl₃ contains a small amount of p-dioxane) δ 8.7 (m, 2 H, C 9,10 Hs), 8.05 (m, 2 H, C 5,8 Hs), 7.5 (s, 2 H, C 6,7 Hs), 6.6 (s, 2 H, C 2,3 Hs). The diol is insoluble in chloroform but soluble in DMSO and DMF.

Preparation of 1,4-Dimethoxyanthracene (1):

To a dried flask, 0.549 g (0.011 mol; 50% oil) of sodium hydride was added. The material was dried under vacuum and flame and maintained under argon. To it 3 mL of distilled ether was added, stirred, and the ether layer containing the oil was removed and discarded. To the sodium hydride flask, 1 g (0.0048 mol) of diol **5** was added and the mixture was dried under vacuum and flame and maintained under argon. DMF (10 mL) was added via syringe and the solution was stirred at room temperature for 1.5 h. TLC of a sample from the reaction solution indicated no starting material. The solution was diluted with 20 mL of water and 6 N HCl until pH = 2. The mixture was extracted with ethyl acetate three time. The combined extract was washed with water twice and brine, dried (MgSO₄), concentrated to give 1.07 g (94% yield) of the crude product (proton NMR spectrum indicated >90% pure). The product was recrystallized from ether:hexane to give 0.89 g (78% yield) of black solids, mp 255 - 258 °C. ¹H NMR (CDCl₃) δ 8.70 (s, 2 H, C9,10H), 7.97 (dd, *J* = 6.1, 3.6 Hz, 2 H, C5,8H), 7.40 (dd, *J* = 6.6, 3.2 Hz, 2 H, C6,7H), 6.55 (s, 2 H, C2,3H), 3.97 (s, 6 H, OCH₃); ¹³C NMR (CDCl₃) δ 149.47 (s, C8a,10a), 131.45 (s, C4a,9a), 128.51 (d, C9,10), 125.47 (d, C5,8), 120.73 (d, C6,7), 100.88 (d, C2,3), 55.61 (s, OCH₃); M.S. (FAB): *m/z* = 238 (M), 239 (MH⁺).

2,5,8-Trimethoxy-4a,9,9a,10-tetrahydro-9,10-[1',2']benzenoanthracene-1,4-dione (**7**), 1,4-Dihydroxy-2,5,8-trimethoxy-9,10-dihydro-9,10-[1',2']benzenoanthracene (**TT3**), and 2,5,8-Trimethoxy-9,10-dihydro-9,10-[1',2']benzenoanthracene-1,4-dione (**TT5**).

A mixture of 0.2 g (0.84 mmol) of 1,4-dimethoxyanthracene (**1**), 0.235 g (1.68 mmol) of methoxyhydroquinone (**6**), 0.39 g (1.68 mmol) of silver oxide and 54 mg (0.17 mmol) of zinc iodide in 20 mL of toluene (distilled over CaH₂) was heated to reflux under argon for 24 h. The

reaction mixture was cooled to room temperature, concentrated and directly subjected to a silica gel column. After elution with a gradient mixture of hexane and ether and then with ethyl acetate and methanol, 0.11 g (55% recovery) of compound **1**, 0.11 g of methoxy-1,4-quinone, 73 mg (51% yield) of **7**, 37 mg (26% yield) of **TT3**, and 18 mg (13% yield) of **TT5** were isolated.

Compound **7**: $^1\text{H NMR}$ (CDCl_3) δ 7.24 - 7.18 (m, 2 H, C12,13 Hs), 7.08 - 7.04 (m, 2 H, C11,14 Hs), 6.65 (s, 2 H, C6,7 Hs), 5.61 (s, 1 H), 5.33 (s, 1 H), 3.82 (s, 6 H, OMe), 3.48 (s, 3 H, OMe), 3.10 (q, $J = 9$ Hz, 2 H, C4a,9a Hs). Compound **TT3**: $^1\text{H NMR}$ (CDCl_3) δ 7.43 (dd, $J = 5.5$, 3 Hz, 1 H), 7.38 (dd, $J = 5.5$, 3 Hz, 1 H), 6.96 (dd, $J = 5.5$, 3 Hz, 2 H, C11,14 Hs), 6.51 (s, 2 H, C6,7 Hs), 6.29 (s, 1 H, C3 H), 6.09 (s, 1 H), 5.98 (s, 1 H), 5.3 (s, 1 H, OH), 4.96 (bs, 1 H, OH), 3.81 (s, 3 H, OMe), 3.805 (s, 3 H, OMe), 3.66 (s, 3 H, OMe). Compound **TT5**: $^1\text{H NMR}$ (CDCl_3) δ 7.43 (dd, $J = 5.5$, 3 Hz, 2 H, C12,13 Hs), 7.0 (dd, $J = 5.5$, 3 Hz, 2 H, C11,14 Hs), 6.52 (s, 2 H, C6,7 Hs), 6.25 (s, 1 H), 6.23 (s, 1 H), 5.71 (s, 1 H, C3 H), 3.79 (s, 6 H, OMe), 3.72 (s, 3 H, OMe).

Isomerization of **7** to **TT3**:

To a solution of 0.77 g (2 mmol) of **7** in 30 mL of p-dioxane and 30 mL of water was added 1.12 g (20 mmol) of KOH. The solution was stirred at room temperature for 1 h, acidified with 1 M HCl, and extracted with methylene chloride three times. The combined extract was washed with brine, dried (MgSO_4), concentrated to give 0.77 g (100% yield) of **TT3**.

Oxidation of **TT3** to **TT5**:

To a mixture of 0.77 g (2 mmol) of **TT3** and 0.6 g of sodium sulfate (anhydrous) in 15 mL of dried acetone under argon at room temperature was added 0.557 g (4 mmol) of silver oxide. The mixture was heated under reflux for 6 h, cooled to room temperature, diluted with methylene chloride, and filtered through Celite. The filtrate was concentrated to give 0.77 g (100% yield) of **TT5**. $^1\text{H NMR}$ spectrum indicates it to be the desired product and was used in the next step without purification.

2-Methoxy-9,10-dihydro-9,10-[1',2']benzenoanthracene-1,4,5,8-tetraone (**TT2**):

To a solution of 0.77 g (0.002 mol) of **TT5** in 16 mL of acetonitrile, 26 mL of p-dioxane and 10 mL of water was added 2 g (3.6 mmol) of ceric ammonium nitrate at room temperature. The solution was stirred for 12 h, diluted with methylene chloride, and washed with water. The organic layer was dried (MgSO_4), concentrated to give the crude product. Recrystallization from ether gave 0.68 g (99% yield) of **TT2**. $^1\text{H NMR}$ (CDCl_3) δ 7.48 (dd, $J = 5.5$, 3 Hz, 2 H, C12,13

Hs), 7.07 (dd, $J = 5.5$, 3 Hz, 2 H, C11,14 Hs), 6.65 (s, 2 H, C6,7 Hs), 6.2 (s, 1 H), 6.18 (s, 1 H), 5.78 (s, 1 H, C3 H), 3.78 (s, 3 H, OMe). MS, CI, m/z 345 ($M+1$).

2-Bromo-3,5,8-trimethoxy-9,10-dihydro-9,10-[1',2']benzenoanthracene-1,4-dione (9):

To a solution of 90 mg (0.24 mmol) of **TT5** in 5 mL of DMF under argon at room temperature was added 52 mg (0.3 mmol) of *N*-bromosuccinimide (NBS). The solution was stirred at 40 °C for 12 h. A sample was withdrawn and checked by TLC to show that no starting material presented. The solution was diluted with ether and washed with twice with water and once with brine, dried ($MgSO_4$), concentrated to give 109 mg (quantitative yield) of the crude product. This material was used in next step without purification. 1H NMR ($CDCl_3$) δ 7.44 (dd, $J = 5.5$, 3 Hz, 2 H, C12,13 Hs), 7.01 (dd, $J = 5.5$, 3 Hz, 2 H, C11,14 Hs), 6.54 (s, 2 H, C6,7 Hs), 6.29 (s, 1 H), 6.22 (s, 1 H), 4.14 (s, 3 H, C3 OMe), 3.81 (s, 3 H, OMe), 3.79 (s, 3 H, OMe).

2-Bromo-3-methoxy-9,10-dihydro-9,10-[1',2']benzenoanthracene-1,4,5,8-tetraone (**TT13**):

To the above product, compound **9**, in a solution of 5 mL of acetonitrile, 6 mL of *p*-dioxane, and 2 mL of water was added 0.66 g (1.2 mmol) of ceric ammonium nitrate. The solution was stirred at room temperature for 6 h, diluted with ether. The mixture was transferred into a separatory funnel, washed with water and brine, dried ($MgSO_4$), concentrated, and column chromatographed on silica gel using a gradient mixture of hexane and ether as eluant to give 60 mg (59% yield) of pure **TT13**; mp 207 - 210 °C. 1H NMR ($CDCl_3$) δ 7.48 (dd, $J = 5.5$, 3 Hz, 2 H, C12,13 Hs), 7.09 (dd, $J = 5.5$, 3 Hz, 2 H, C11,14 Hs), 6.66 (s, 2 H, C6,7 Hs), 6.23 (s, 1 H), 6.16 (s, 1 H), 4.17 (s, 3 H, C3 OMe). ^{13}C NMR ($CDCl_3$) δ 182.2 (s, C1), 182.0 (s, C4), 176.2 (s, C=O), 175.5 (s, C=O), 156.2, 151.6, 151.4, 150.2, 141.8, 135.5, 135.4, 126.2, 126.1, 126.07, 126.0, 125.6, 125.4, 117.2, 61.7 (q, OMe), 43.0 (d), 42.1 (d). MS EI, m/z 424 & 422 (1:1, M^+), 344 ($M-Br$), 300, 287, 232, 152, 126.

2-Chloro-5,8-dimethoxy-4a,9,9a,10-tetrahydro-9,10-[1',2']benzenoanthracene-1,4- dione (**12**):

A solution of 0.1 g (0.42 mmol) of 1,4-dimethoxyanthracene (**1**), 0.121 g (0.84 mmol) of chlorohydroquinone (**11**), 0.195 g (0.84 mmol) of silver oxide, and 27 mg (0.084 mmol) of zinc iodide in 10 mL of toluene was heated under reflux for 10 h. The mixture was concentrated on a rotary evaporator and the residue was subjected to a silica gel column. After elution of a gradient mixture of hexane and ether, 64 mg (73% yield based on reacted **1**) of **12** and 45 mg (45% recovery) of **1**. Compound **12**: 1H NMR ($CDCl_3$) δ 7.42 (m, 2 H, C12,13 Hs), 7.18 (dd, $J = 5.5$, 3 Hz, 2 H, C11,14 Hs), 6.64 (d, $J = 6$ Hz, 1 H), 6.61 (d, $J = 6$ Hz, 1 H), 6.55 (s, 1 H, C3

H), 5.31 (d, $J = 3$ Hz, 1 H), 5.28 (d, $J = 3$ Hz, 1 H), 3.76 (s, 3 H, OMe), 3.75 (d, 3 H, OMe), 3.17 (qd, $J = 9$, 2.6 Hz, 2 H, C4a,9a Hs).

2-Methoxy-4a,9,9a,10-tetrahydro-9,10-[1',2']benzenoanthracene-1,4-dione (TT8):

A mixture of 0.5 g (2.8 mmol) of anthracene (**13**), 0.787 g (5.6 mmol) of methoxyhydroquinone (**6**), 1.298 g (5.6 mmol) of silver oxide and 0.179 g (0.56 mmol) of zinc iodide in 10 mL of toluene was heated under reflux for 24 h under argon. The mixture was diluted with methylene chloride and water, filtered through Celite, and the filtrate was transferred into a separatory funnel. After washing with brine, the organic layer was dried (MgSO_4), concentrated to give dark green solids; NMR spectrum of this material indicated desired product. The solid was recrystallized from ether to give 0.50 g (56% yield) of **TT8** as light green white solids, mp 182-184 °C; ^1H NMR (CDCl_3) d 7.4 (m, 2 H), 7.2 (m, 4 H), 7.1 (m, 2 H), 5.64 (s, C3 H), 4.89 (s, 2 H, C9,10 Hs), 3.51 (s, 3 H, MeO), 3.17 (qd, $J = 9.5$, 3 Hz, 2 H, C4a,9a Hs); ^{13}C NMR (CDCl_3) d 193.25 (s, C=O), 162.84, 161.76, 141.63, 139.84, 139.29, 126.68, 126.66, 126.57, 126.51, 124.91, 124.61, 123.89, 123.82, 113.57, 56.12 (OMe), 49.56, 49.15, 48.87, 48.84.

2-Methoxy-9,10-dihydro-9,10-[1',2']benzenoanthracene-1,4-dione (TT9):

To a solution of 0.2 g (0.63 mmol) of ketone **TT8** in 5 mL of p-dioxane and 1 mL of water was added 0.106 g (1.9 mmol) of KOH. The solution was stirred at room temperature for 2 h, neutralized with 1 N HCl, and extracted with methylene chloride twice. The combined extract was washed with brine, dried (MgSO_4), concentrated to give the crude product diol. This compound was used in the following oxidation reaction without purification. A solution of the above diol, 0.4 g (2.8 mmol) of sodium sulfate (anhydrous), and 0.4 (1.7 mmol) of silver oxide in 12 mL of acetone was stirred at room temperature for 3 h. The mixture was diluted with methylene chloride, filtered through Celite, washed with brine, dried (MgSO_4), concentrated, and column chromatographed on silica gel using a gradient mixture of hexane and ether as eluant to give 0.16 g (80% yield) of **TT9**; mp 195-197 °C; ^1H NMR (CDCl_3) d 7.43 (m, 2 H, C12,13 Hs), 7.03 (dd, $J = 5$, 3 Hz, 2 H, C11,14 Hs), 5.83 (s, 1 H), 5.80 (s, 1 H), 5.74 (s, 1 H, C3 H), 3.76 (s, 3 H, OMe); ^{13}C NMR (CDCl_3) d 183.45 (s, C=O), 178.2 (s, C=O), 158.48 (s), 152.93 (s), 150.07 (s), 143.71 (s, 2 C), 143.57 (s, 2 C), 125.52 (d, 4 C), 124.43 (d, 2 C), 124.3 (d, 2 C), 105.71 (d), 56.48 (q, OMe), 47.5 (d), 47.17 (d).

4a,9,9a,10-Tetrahydro-9,10-[1',2']benzenoanthracene-1,4-dione (TT6):

A solution of 1 g (5.6 mmol) of anthracene (**13**) and 2.42 g (22 mmol) of 1,4-benzoquinone (**2**) in 20 mL of toluene was heated under reflux under argon for 24 h. The solution was cooled to room temperature, diluted with 100 mL of methylene chloride, washed with water, dried (MgSO₄), and concentrated to give yellow solids. Proton NMR spectrum indicated desired product and starting quinone **2**. The solids were washed with ether several time (the product is insoluble in ether and benzoquinone dissolves), and the remaining solids were dried under vacuum to give 1.56 g (98% yield) of **TT6**; mp 219-221 °C; ¹H NMR (CDCl₃) δ 7.40 (dd, *J* = 5, 3 Hz, 2 H), 7.2 ~ 7.17 (m, 4 H), 7.08 (dd, *J* = 5, 3 Hz, 2 H), 6.32 (s, 2 H, C2,3 Hs), 4.87 (s, 2 H, C9,10 Hs), 3.14 (s, 2 H, C4a,9a Hs); ¹³C NMR (CDCl₃) δ 198.29 (s, C=O), 141.49 (s), 140.53 (d), 139.64 (s), 126.68 (d), 126.59 (d), 124.68 (d), 123.83 (d), 49.0 (d), 48.85 (d).

9,10-Dihydro-9,10-[1',2']benzenoanthracene-1,4-dione (**TT7**):

The procedure is the same as that of the preparation of **TT9** from **TT8**. A solution of 1 g (3.5 mmol) of **TT6** and 0.784 g (14 mmol) of KOH in 50 mL of p-dioxane and 10 mL of water was stirred at room temperature for 2 h. The solution was acidified with 1 N HCl until pH = 1, extracted twice with methylene chloride, and the combined methylene chloride layer was washed with brine, dried (MgSO₄), concentrated to give dark brown solids. The proton NMR spectrum indicated a mixture of the diol (major product) and **TT7** (minor). The mixture was stirred with 2 g (8.6 mmol) of silver oxide and 2 g (14 mol) of anhydrous sodium sulfate in 60 mL of dried acetone at room temperature under argon for 3 h. The mixture was filtered through Celite, rinse with methylene chloride, and the filtrate was concentrated to give dark brown solids. Column chromatographic separation of the crude product with silica gel using a gradient mixture of hexane and ether to give 0.7 g (70% yield) of **TT7**. Mp 295-298 °C; ¹H NMR (CDCl₃) δ 7.42 (dd, *J* = 5, 3 Hz, 4 H), 7.03 (dd, *J* = 5, 3 Hz, 4 H), 6.6 (s, 2 H, C2,3 Hs), 5.8 (s, 2 H, C9,10 Hs); ¹³C NMR (CDCl₃) δ 183.52 (s, C=O), 151.92 (s), 143.57 (s, 4 C), 135.39 (d, 2 C), 125.57 (d, 4 C), 124.42 (d, 4 C), 47.36 (d, C9,10).

5,8-Dimethoxy-2-methoxycarbonyl-4a,9,9a,10-tetrahydro-9,10-[1',2']benzeno-anthracene-1,4-dione (**TT10**) and 5,8-Dimethoxy-4a-methoxycarbonyl-9,9a,10-trihydro-9,10-[1',2']benzenoanthracene-1,4-dione (**TT11**):

A mixture of 0.2 g (0.00119 mol) of methyl gentisate (**14**), 0.2 g (0.0014 mol) of potassium carbonate and 0.6 g (0.0026 mol) of silver oxide in 10 mL of benzene (freshly distilled) was stirred at 50 °C (bath temperature) under argon for 10 min under dark. The reaction mixture was cooled to room temperature, filtered through Celite, rinsed with small

amount of ether, concentrated on a rotary evaporator and then under vacuum to give 0.198 g (100% yield) of methoxycarbonyl-1,4-benzoquinone (**15**); ^1H NMR (CDCl_3) δ 7.13 (s, 1 H, C3 H), 6.84 (s, 2 H, C5,6 Hs), 3.92 (s, 3 H, OMe). The proton NMR spectrum indicates the material is about 100% pure and is used in the next step without purification.

A solution of 73 mg (0.44 mmol) of **15** and 70 mg (0.294 mmol) of 1,4-dimethoxyanthracene (**1**) in 1 mL of toluene was heated at 70 $^\circ\text{C}$ for 14 h and under reflux for 5 h under argon. The solution was cooled to room temperature and subjected to a silica gel column using gradient mixtures of hexane, ether and ethyl acetate to give 69 mg (58% yield) of **TT11** and 44 mg (40% yield) of **TT10**. Compound **TT11**: mp 176-178 $^\circ\text{C}$; ^1H NMR (CDCl_3) δ 7.24 (m, 1 H), 7.18 (m, 1 H), 7.05 (dd, $J = 5, 3$ Hz, 2 H), 6.65 (dd, $J = 10, 9$ Hz, 2 H, C6,7 Hs), 6.38 (d, $J = 10$ Hz, 1 H, C2 H), 6.28 (d, $J = 10$ Hz, 1 H, C3 H), 5.67 (s, 1 H, C10 H), 5.27 (d, $J = 2.4$ Hz, 1 H, C9 H), 3.86 (s, 3 H, OMe), 3.81 (s, 3 H, OMe), 3.61 (s, 3 H, OMe), 3.5 (d, $J = 2.4$ Hz, 1 H, C9a H); ^{13}C NMR (CDCl_3) δ 196.67 (s, C=O), 193.48 (s, C=O), 169.23 (s, C=O of ester), 149.85 (s), 148.08 (s), 140.61 (2), 140.22 (d), 139.31 (d), 131.4 (s), 129.11 (s), 126.7 (d), 126.59 (d), 125.93 (d), 124.68 (d), 111.2 (s), 109.4 (d), 104.36 (d), 63.4 (s), 56.23 (q), 55.91 (q), 53.9 (q), 53.17 (d), 45.02 (d), 42.1 (d). Compound **TT10**: ^1H NMR (CDCl_3) δ 7.4 (m, 2 H), 7.18 (m, 2 H), 6.54 (s, 2 H), 6.42 (s, 1 H of one isomer; exo or endo), 6.25 (s, 1 H of one isomer), 5.28 (m, 1 H), 5.25 (m, 1 H), 3.86 (s, 3 H, OMe), 3.83 (s, 3 H, OMe), 3.82 (s, 3 H, OMe), 3.15 (m, 1 H), 3.12 (m, 1 H).

9,10-Dihydro-9,10-[1',2']benzenoanthracene-1,4,5,8-tetraone (**TT1**):

Preparation of *endo*-(5a*S*, 8a*R*, 9*S*, 10*R*)-1,4-dimethoxy-5a,8a,9,10-tetrahydro-9,10[1',2']benzenoanthracene-5,8-dione (**18a**) and *exo*-(5a*R*, 8a*S*, 9*S*, 10*R*)-1,4-Dimethoxy-5a,8a,9,10-tetrahydro-9,10-[1',2']benzenoanthracene-5,8-dione (**18b**).

To a solution of 1.19 g (0.005 mol) 1,4-dimethoxyanthracene (**1**) in 20 mL of toluene was added a solution of 0.54 g (0.005 mol) 1,4-benzoquinone (**2**) in 15 mL of toluene, the resulting solution was heated at reflux for 3 hr., added 1.08 g (0.01 mol) 1,4-benzoquinone. After 24 hr., another portion of 1.08 (0.01 mol) 1,4-benzoquinone was added. The solution was refluxed for 19 h. Subsequent evaporation of solvent and sublimation of excess 1,4-benzoquinone gave a mixture of 1.85 g of isomers **18a** and **18b** (~100% yield) in a ratio of 1:1. Further purification was performed and the products (1.73g) were used for the next step. The *endo*-, *exo*- isomers were assigned according to ¹H NMR spectra analysis. The deshielding effect between enone ring and benzene ring in the *exo*- configuration resulted in a downfield shift of proton signals on these two rings ,especially the benzene protons (7.41ppm, 7.17ppm), compared with those of *endo*- isomer (7.20ppm, 7.06ppm). Mp. 198~201 °C; **18a** (*endo*-): ¹HNMR (CDCl₃) δ 7.20 (dd, *J* = 5.5, 3.2 Hz, 2 H, C3',6'), 7.06 (dd, *J* = 5.5, 3.2 Hz, 2 H, C4', 5'H), 6.59 (s, 2 H, C2,3H), 6.28 (s, 2 H, C6,7H), 5.31 (s, 2 H, C9,10H), 3.82 (s, 6 H, OMe). **18b** (*exo*-): Mp 218-221 °C ¹HNMR (CDCl₃) δ 7.41 (dd, *J* = 5.5, 3.2 Hz, 2 H, C3',6H'), 7.17 (dd, *J* = 5.5, 3.2 Hz, 2 H, C5', 4'H), 6.59 (s, 2 H, C2,3H), 6.31 (s, 2 H, C6,7H), 5.31 (s, 2 H, C9,10H), 3.75 (s, 6 H, OMe). The structure of this *exo* isomer (the two C4a,9a Hs orient at the same side with the phenyl ring) was proven by a single crystal X-ray diffraction.

Exoisomer: ¹³C NMR (CDCl₃)

δ 198.44, 148.500, 140.46, 139.95, 131.21, 126.52, 124.86, 109.09, 56.04, 48.51, 42.16.

Endoisomer: ¹³C NMR (CDCl₃)

δ 198.12, 149.13, 141.808, 140.29, 128.32, 126.45, 123.99, 109.32, 126.45, 123.99, 109.76, 56.39, 48.80, 42.38.

Preparation of 1,4-dimethoxy-5,8-dihydroxy-9,10-dihydro-9,10-[1',2']benzenoanthracene (**19**):

To a solution of 1.73 g (0.005 mol) of **18a&b** in 100 mL of 1,4-dioxane was added 10.61g of potassium hydroxide in 100 mL of water. The resulting mixture was stirred at 35 °C for 18.5 hr., neutralized by 6N hydrochloric acid solution, and extracted with methylene chloride twice (40 ml each), and once with ethyl acetate (40 ml). The combined extracts were dried over anhydrous magnesium sulfate, and concentrated to give 1.75 g of crude product **19** (~100% crude yield). No purification was performed at this stage. ¹H NMR (CDCl₃)

d: 3.80 (s, 6 H, -OCH₃), 6.20 (s, 2H, C 9,10 Hs), 6.34 (s, 2H, C 6,7 Hs), 6.51 (s, 2H, C 2,3 Hs), 7.00 (dd, J=5.5, 3.2 Hz, 2H, C 12,15 Hs), 7.43 (dd, J=5.5, 3.2 Hz, 2H, C 13,14 Hs).

5,8-Dimethoxy-9,10-dihydro-9,10[1'2']benzenoanthracene-1,4-dione (**20**):

5 A mixture of 1.73 g (0.005 mol) of crude **19**, 6 g (0.042 mol) of sodium sulfate and 5.8 g (0.025 mol) of silver oxide in 150 mL of acetone was heated at reflux for 22 hr.. The reaction mass was filtered through Celite and washed with methylene chloride. The filtrate was dried over anhydrous magnesium sulfate, concentrated and column chromatographed on silica gel using a gradient mixture of ethyl acetate and hexane as eluant to give 1.15 g of pure **20** in a yield of 67.25% (based on crude **19**). The melting point and IR agree with reported data.¹ M.p. 266~271 °C (decom.), lit.¹ 271.5~272.5 °C; IR: 3060 cm⁻¹ (Ar-H), 1660 cm⁻¹ (C=O), 1584 cm⁻¹ (aromatic C=C), 1258 cm⁻¹ (C-O); ¹H NMR (CDCl₃) δ 7.44 (dd, J = 5.3, 3.2 Hz, 2 H, C12,13 Hs), 7.00 (dd, J = 5.3, 3.2 Hz, 2 H, C11,14 Hs), 6.54 (s, 2 H, C6,7 Hs), 6.51 (s, 2 H, C2,3 Hs), 6.23 (s, 2 H, C9,10 Hs), 3.79 (s, 6 H, OMe); ¹³C NMR (CDCl₃) δ 183.52 (s, C=O), 152.75 (s, C-OMe), 149.51 (s), 144.14 (s), 135.23 (d), 133.44 (s), 125.28 (d), 124.51 (d), 109.36 (d, C2,3), 56.30 (s, OMe), 41.30 (d, CH).

Preparation of 9,10-Dihydro-9,10-[1',2']-benzenoanthracene-1,4,5,8-tetrone (**TT1**):

20 To a solution of 0.4 g (1.17 mmol) of compound **20** in 30 mL of acetonitrile was added dropwise a solution of 3.2 g (5.84 mmol) of ceric ammonium nitrate in 70 mL of a mixture of dioxane and water (3:1) at r.t under argon. After stirred for 3 h, the reaction mixture was extracted with methylene chloride four times (40 mL each). The combined methylene chloride extracts were dried over anhydrous magnesium sulfate, concentrated and column chromatographed on silica gel using a gradient mixture of ethyl acetate and petroleum ether as eluant to give 0.37 g of bisquinone **TT1** in a yield of about 100%. Recrystallization in methylene chloride and petroleum ether gave **TT1** as yellow needle-like crystals; m.p. 303~305 °C (dec.), lit.¹ >220 °C (dec.); ¹H NMR (CDCl₃) δ 7.49 (dd, J = 5.3, 3.0 Hz, 2 H, C12,13 Hs), 7.08 (dd, J = 5.3, 3.0 Hz, 2 H, C11,14 Hs), 6.65 (s, 4 H, =CH-CO), 6.18 (s, 2 H, CH); ¹³C NMR (CDCl₃) δ 182.22 (s, C=O), 151.51 (s), 142.20 (s), 135.41 (d), 125.91 (d), 125.47 (d), 42.13 (d).

Preparation of dimethyl butadiene-2,3-dicarboxylate (**16**):

(Z)-Dimethyl 2,3-dimethyl-2-butenedioate (**21**):

To a solution of 2 g (0.0158 mol) of 2,3-dimethylmaleic anhydride in 40 mL of methanol was added 0.2 mL of concentrated sulfuric acid. After refluxing for 7 days, the solution was diluted with 100 mL of methylene chloride, neutralized with aqueous sodium bicarbonate solution. The methylene chloride layer was washed with 30 mL of brine, the aqueous layer was extracted with methylene chloride four times (30 mL twice and 15 mL twice). The combined methylene chloride layer and extracts were dried over anhydrous magnesium sulfate, concentrated to give 2.11 g of crude product. ¹H NMR check indicated about 25% of starting material existing, the crude mixture with 0.2 mL of concentrated sulfuric acid in 40 mL of methanol was heated at reflux again for 5 days. The solution was concentrated until 30% of methanol was left, neutralized with saturated sodium bicarbonate aqueous solution, followed by addition of 30 mL of brine solution, and the mixture was extracted with ethyl acetate four times (100 mL once, 30 mL three times). The combined ethyl acetate extracts were dried over anhydrous magnesium sulfate, concentrated to give 3.26 g of crude product **21** (~100% yield). It was of satisfactory purity for the next step. ¹H NMR (CDCl₃) δ 3.77 (s, 6 H, OMe), 1.95 (s, 6 H, Me)

(Z)- and (E)-Dimethyl-2,3-bis[bromomethyl]-2-butenedioate (**22a** & **22b**):

A mixture of 3.2384 g (0.0188 mol) of butenedioate **21**, 7.37 g (0.04 mol) of *N*-bromosuccinimide (NBS) and 100 mg (cat.) of 2,2'-azobisisobutyronitrile (AIBN) in 50 mL of carbon tetrachloride was heated under reflux and irradiated with a 275 W sunlamp for 3 h. The reaction solution was cooled, diluted with 150 mL of ether, and filtered. The filtrate was concentrated, and flash column chromatographed using a gradient mixture of ether-hexane as eluant to give 1.33 g of oily liquid *trans*-isomer **22b** (18.4% yield) and 4.40 g of oily liquid *cis*-isomer **22a** (71.6% yield) with a total yield of 90%. The assignment of **22a** and **22b** was based on the reported ¹H NMR data.

22a: ¹H NMR (CDCl₃) δ 4.50 (s, 4 H, CH₂ Br), 3.91 (s, 6 H, OMe); ¹³C NMR (CDCl₃) δ 165.54 (s, C=O), 136.95 (s, C=), 52.97 (s, OMe), 26.75 (t, CH₂Br)

22b: ¹H NMR (CDCl₃) δ 4.24 (s, 4 H, CH₂Br), 3.84 (s, 6 H, OMe); ¹³C NMR (CDCl₃) δ 165.96 (s, C=O), 137.06 (s, C=), 52.99 (s, OMe), 23.95 (CH₂Br).

Dimethyl 1,3-butadiene-2,3-dicarboxylate (**16**):

To a solution of 6.53 g (0.02 mol) of bromoesters **22** in 20 mL of acetone was added 9.49 g (0.06 mol) of sodium thiosulphate and 9.96 g (0.06 mol) of potassium iodide. After refluxing for 2 hr., The mixture was cooled, poured onto 50 g of ice, and extracted with methylene chloride three times (150 mL once, 50 mL twice). The combined methylene chloride extracts were washed with 20 mL of saturated aqueous sodium chloride, dried over anhydrous magnesium sulfate, concentrated, and column chromatographed on silica gel using a gradient mixture of hexane- ethyl acetate as eluant to give 2.8206 g of butadiene **16** in a yield of 83%. ¹H NMR (CDCl₃) δ 6.71(dd, *J* = 1.4, 0.6 Hz, 2 H, =CH), 5.83 (dd, *J* = 1.4, 0.6 Hz, 2 H, =CH), 3.76 (s, 6 H, OMe); ¹³C NMR (CDCl₃) δ 166.20 (s, C=O), 138.42 (s, =C), 127.79 (t, =CH₂), 52.09 (q, OMe).

(4a*S**,7a*R**,11a*S**,14a*R**)-Tetramethyl 1,4,4a,5,6,7,7a,8,11,11a,12,13,14,14a-tetradecahydro-5,7,12,14-tetraoxo-6,13-[1',2']benzenopentacene-2,3,9,10-tetracarboxylate (TT4) and Compound **17**:

A solution of 0.14 g (0.45 mmol) of **TT1** and 0.17 g (1 mmol) of compound **16** in 5 mL of toluene was maintained under argon and heated under reflux for 22 h. The solution was directly subjected onto a silica gel column and eluted with a gradient mixture of hexane, methylene chloride, and ethyl acetate to give 0.160 g (54% yield) of **TT4** and 30 mg (14% yield) of compound **17**. Recrystallization of **TT4** from methylene chloride gave light yellow crystals. Single crystal X-ray diffraction analysis was carried on a crystal and the structure was solved. The following Figure shows the ORTEP drawing of the compound. ¹H NMR (CDCl₃) δ 7.45 (m, 2 H), 7.09 (m, 2 H), 6.07 (s, 2 H, C6,13 Hs), 3.75 (s, 12 H, OMe), 3.2 (bs, 4 H, C4a,7a,11a,14a Hs), 2.62 ~ 2.5 (m, 8 H); ¹³C NMR (CDCl₃) δ 192.36 (s, C=O), 167.12 (s, C=O of ester), 154.04 (d), 141.49 (s), 132.51 (d), 126.02 (s), 125.57 (s), 52.33 (q, OMe), 45.5 (d), 43.27 (d), 25.48 (t).

Compound **17**: ¹H NMR (CDCl₃) δ 7.42 (dd, *J* = 5, 3 Hz, 2 H), 7.01 (dd, *J* = 5, 3 Hz, 2 H), 6.38 (s, 2 H), 6.10 (s, 2 H), 3.71 (s, 6 H, OMe), 3.17 (bs, 2 H, CHC=O), 2.60 (dd, *J* = 17, 4 Hz, 2 H), 2.43 (dd, *J* = 17, 4 Hz, 2 H). (19).

(4*aS**, 5*S**, 7*R**, 12*S**, 14*R**, 7*aR**, 11*aS**, 14*aR**) - Tetramethyl 1,4,4*a*,5,6,7,7*a*,8,11,11*a*,12,13,14,14*a*-tetradecahydro-5,7,12,14-tetrahydroxy-6,13-[1',2']benzenopentacene-2,3,9,10-tetracarboxylate (TT12):

To a solution of 0.10 g (0.15 mmol) of **TT4** in 7 mL of methanol (distilled over magnesium turning) under argon were added 0.52 g (1.4 mmol) of CeCl₃•7H₂O and 51 mg (1.4 mmol) of sodium borohydride. The solution was stirred at room temperature for 12 h, diluted with 70 mL of methylene chloride, and washed with aqueous ammonium chloride and brine. The organic layer was dried (MgSO₄), concentrated to give 88 mg (89% yield) of **TT12**. Proton NMR spectrum indicates the material is pure. ¹H NMR (CDCl₃) δ 7.3 (dd, *J* = 5, 3 Hz, 2 H), 7.01 (dd, *J* = 5, 3 Hz, 2 H), 5.15 (s, 2 H, C6,13 Hs), 4.47 (bs, 4 H, CHO), 3.74 (s, 12 H, OMe), 3.2 (bs, 4 H, OH), 2.53 (dd, *J* = 17, 4 Hz, 4 H), 2.27 (dd, *J* = 17, 4 Hz, 4 H), 2.06 (m, 4 H); ¹³C NMR (CDCl₃ and acetone-d₆) δ 168.24 (s, C=O of ester), 145.74 (s), 145.45 (s), 134.46 (s), 123.49 (d), 121.76 (d), 68.32, 59.66, 51.28, 49.36, 34.19.

2-Bromo-3-(methylamino)-9,10-dihydro-9,10-[1',2']benzenoanthracene-1,4,5,8-tetraone (**TT14**) and 2-hydroxy-3-(methylamino)-9,10-dihydro-9,10-[1',2']benzenoanthracene-1,4,5,8-tetraone (**TT15**):

A solution of 24 mg (0.057 mmol) of **TT13** and 1.8 mg (0.057 mmol) of methylamine in 1 mL of THF was stirred under argon at room temperature for 20 min. The solution was diluted with ether, washed with water and brine, dried (MgSO₄), and concentrated. Column chromatographic separation of the crude on silica gel using a gradient mixture of hexane and ether as eluant to give 6 mg (24% yield) of **TT14** and 6 mg (29% yield) of **TT15**. For **TT14**: ¹H NMR (CDCl₃) δ 7.48 (dd, *J* = 5, 3 Hz, 1 H), 7.45 (dd, *J* = 5, 3 Hz, 1 H), 7.07 (dd, *J* = 5, 3 Hz, 2 H), 6.64 (s, 2 H, C6,7 Hs), 6.28 (s, 1 H), 6.11 (s, 1 H), 5.88 (bs, 1 H, NH), 3.33 (d, *J* = 5.6 Hz, 3 H, MeN). For **TT15**: ¹H NMR (CDCl₃) δ 7.45 (m, 2 H, Ar-H), 7.04 (m, 2 H, Ar-H), 6.41 (2d, *J* = 6.3 Hz, 2 H, C6,7-Hs), 6.23 (s, 1 H), 6.1 (s, 1 H), 5.9 (bs, 1 H, NH), 4.8 (s, 1 H, OH), 3.33 (d, *J* = 5.8 Hz, 3 H, MeN).

2-Bromo-3-methoxy-6-(dimethylamino)-9,10-dihydro-9,10-[1',2']benzenoanthracene-1,4,5,8-tetraone (**TT16**):

To a solution of 35 mg (0.083 mmol) of **TT13** in 2 mL of THF, a stream of dimethylamine was added via a syringe for few minutes. The solution turned dark brown color

immediately, the dimethylamine gas was disconnected, and the solution was stirred at 0°C for 10 minutes. The solvent and excess of dimethylamine were removed via rotary evaporator and residue was column chromatographed on silica gel using a gradient mixture of hexane and ethyl acetate as eluant to give 5 mg of recovered starting material **TT13** and 28 mg of the product **TT16**. ¹H NMR (CDCl₃) δ 7.45 (m, 2 H, Ar-H), 7.04 (m, 2 H, Ar-H), 6.22 (s, 1 H), 6.15 (s, 1 H), 5.4 (s, 1 H, C7H), 3.81 (s, 3 H, OMe), 3.12 (s, 6 H, Me₂N).

2-Bromo-3-[2-(ethoxycarbonyl)ethylamino]-9,10-dihydro-9,10-[1',2']benzenoanthracene-1,4,5,8-tetraone (**TT17**) and 2-hydroxy-3-[2-(ethoxycarbonyl)ethylamino]-9,10-dihydro-9,10-[1',2']benzenoanthracene-1,4,5,8-tetraone (**TT18**).

To a mixture of 0.5 g (0.00325 mol) of b-alanine ethyl ester hydrochloride in 5 mL of THF under argon, 0.156 g (0.00325 mol; 50% oil dispersion) was added. The mixture was stirred at room temperature for 1 h, THF was evaporated under vacuum and while solids, b-alanine ethyl ester and sodium chloride, were obtained.

To a solution of 64 mg (0.151 mmol) of **TT13** in 2 mL of THF and 0.5 mL of DMF under argon was added 0.302 mmol each of the above mixture of b-alanine ethyl ester and sodium chloride. The solution was stirred at room temperature for 16 hours, diluted with methylene chloride, the organic layer was washed with water twice, brine, dried (MgSO₄), and concentrated. The proton NMR spectrum of the crude products indicated a mixture of **TT17** and **TT18**. This crude product was subjected to column chromatographic separation on silica gel using a gradient mixture of hexane and ethyl acetate as eluant to give 46 mg (69% yield) of **TT18**. ¹H NMR (CDCl₃) δ 7.45 (m, 2 H, Ar-H), 7.04 (m, 2 H, Ar-H), 6.42 (ABq, *J* = 8.4 Hz, 2 H, C6,7 H's), 6.23 (s, 1 H, C9H), 6.10 (s, 1 H, C10H), 4.15 (q, *J* = 7 Hz, 2 H, CH₂), 4.10 (broad s, 2 H, CH₂N), 2.64 (t, *J* = 6 Hz, 2 H, CH₂CO), 1.25 (t, *J* = 7 Hz, 3 H, CH₃).

2-Hydroxy-3-[(*S*)-5-(hydroxycarbonyl)-5-aminopentylamino]-9,10-dihydro-9,10-[1',2']benzenoanthracene-1,4,5,8-tetraone (**TT20**).

To a mixture of 0.5 g (0.0027 mol) of L-lysine monohydrochloride in 5 mL of THF under

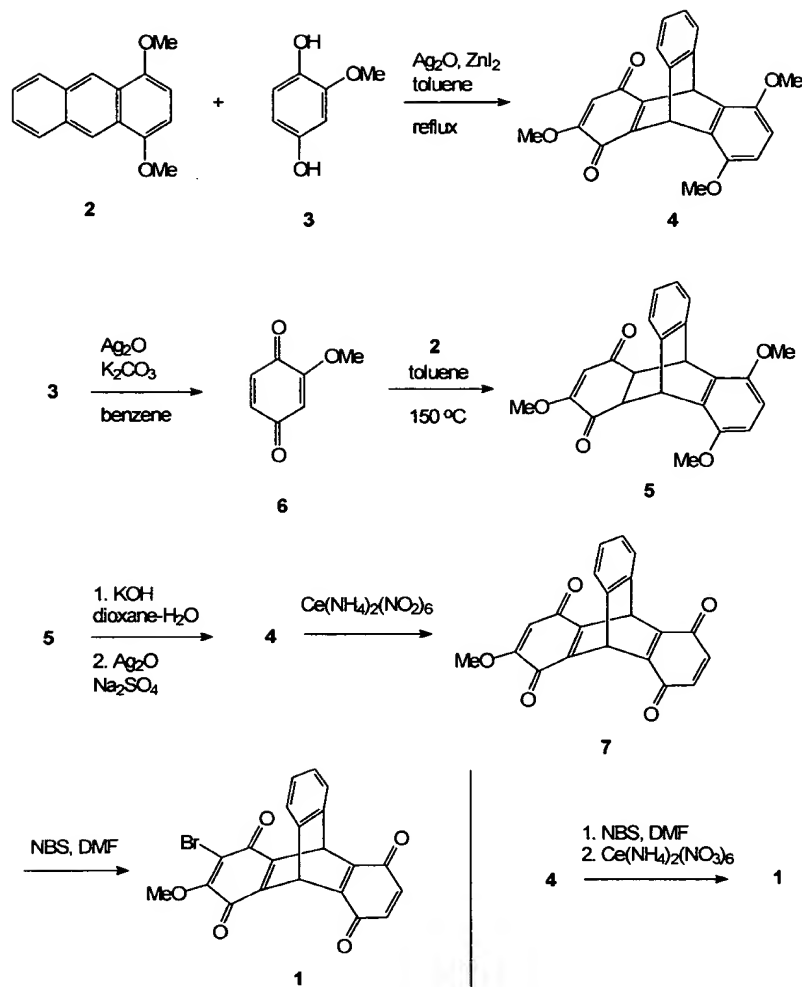
argon was added 0.1315 g (0.0027 mol; 50% oil) of sodium hydride. The mixture was stirred at room temperature for 2 h, concentrated to dryness under vacuum to give a mixture of L-lysine and sodium chloride as white solids. To a solution of 40 mg (0.095 mmol) of **TT13** in 2 mL of THF and 0.5 mL of DMF was added 20 mg of the above mixture of L-lysine and sodium chloride. The mixture was stirred at room temperature for 2 days, concentrated to dryness under vacuum and the crude product was purified by HPLC using a Econosphere C8 (10 m) semi-preparative column (250 mm x 10 mm), 40% acetonitrile in water as solvent, to give pure **TT20** (came out in 2 minutes). ¹H NMR (D₂O) δ 7.54 (m, 2 H, Ar-H), 7.10 (m, 2 H, Ar-H), 6.56 (s, 2 H, C6,7 H's), 6.2 (s, 1 H, C9H), 6.07 (s, 1 H, C10H), 3.76 (t, *J* = 6 Hz, 1 H, CH), 3.03 (dd, *J* = 8 Hz, 7 Hz, 2 H, CH₂N), 1.90 (m, 2 H, CH₂), 1.72 (m, 2 H, CH₂), 1.48 (m, 2 H, CH₂).

ONE POT SYNTHESIS OF 5,8-DISUBSTITUTED TRIPTYCENE MONOQUINONES AND INHIBITION STUDIES.

In the reported synthesis of 5,8-disubstituted triptycene monoquinones, (Bartlett, P. D.; Ryan, M. J.; Cohen, S. G. *J. Am. Chem. Soc.* 1942, 64, 2649; Skvarchenko, V. R.; Shalaev, V. K.; Klabunovskii, E. I. *Russ. Chem. Rev.* 1974, 43, 951; Iwamura, H.; Maino, K. An intramolecular triptycene quinhydrone. *J. Chem. Soc. Chem. Commun.* 1978, 720.; Quast, H.; Fuchsbauer, H. -L. *Chem. Ber.* 1986, 119, 1016-1038; Quast, H.; Fuchsbauer, H. -L. *Chem. Ber.* 1986, 119, 2414; Patney, H. K. *Synthesis* 1991, 694. Charge-transfer complexes; Lipczynska-Kochany, E.; Iwamura, H. *Chem. Lett.* 1982, 1075; Daub, J.; Jakob, L.; Salbeck, J. *Chem. Ber.* 1988, 121, 2187. Radical Anions: Russell, G. A.; Suleman, N. K. *J. Am. Chem. Soc.* 1981, 103, 1560-1561. Liquid Crystals; Norvez, S. *J. Org. Chem.* 1993, 58, 2414) a sequence of three reactions was carried out: Diels-Alder reaction of 1,4-dimethoxyanthracene (Criswell, T. R.; Klanderman, B. H. *J. Org. Chem.* 1974, 39, 770) and p-benzoquinone followed by isomerization of the adducts (endo- and exo-adducts) with potassium hydroxide and then oxidation with silver oxide. These three reactions have been successfully combined into one simple operation. Hence, heating of anthracene 2 (derived from the reduction of 1,4-anthracenedione (Perchellet, E. M.; Magill, M. J.; Huang, X.; Brantis, C. E.; Hua, D. H.; Perchellet, J. P. *Anti-cancer Drugs*, 1999, 10, 749) with sodium hydrosulfite followed by sodium hydride and iodomethane), 1.4 equiv of methoxyhydroquinone (3), 2.7 equiv of silver oxide, and 0.2 equiv of zinc iodide in toluene under reflux for 4 days gave a 70% yield of triptycene monoquinone 4 (Scheme 14).

Presumably, silver oxide oxidizes methoxyhydroquinone to methoxy-p-benzoquinone which undergoes Diels-Alder reaction with anthracene 2 to give adducts 5 (endo- and exo-isomers). Compound 5 then undergoes oxidation with silver oxide to give quinone 4. A catalytic amount of zinc iodide was added to facilitate the Diels-Alder reaction. No other regioisomers (such as isomers with methoxy substituent attached at C4a) were detected. To verify the reaction sequence, methoxyquinone 6, obtained from the oxidation of 3 with silver oxide and potassium carbonate in benzene (98% yield), was treated with anthracene 2 in toluene at 150 °C in a sealed tube to give adducts 5 (48% yield). Isomerization of ketones 5 with KOH in p-dioxane and water followed by oxidation with silver oxide gave monoquinone 4 (95% overall yield). Oxidation of monoquinone 4 with ceric ammonium nitrate afforded a 94% yield of triptycene bisquinone 7. Regioselective bromination of 7 was achieved by the treatment with N-bromosuccinimide (NBS) in DMF at 25 °C for 10 h to give bromoquinone 1 (45% yield). Alternatively, compound 1 can also be obtained from the bromination of monoquinone 4 with NBS in DMF at 40 °C followed by oxidation with ceric ammonium nitrate (59% overall yield). The bromination of quinones with NBS has not been reported previously, and apparently, reactivity of the methoxyquinone moiety towards NBS is greater than that of quinone and 1,4-dimethoxyphenyl moieties. Likely, the methoxy substituent of 7 or 4 (at C2) enhances nucleophilicity of adjacent double bond carbon resulting the bromination.

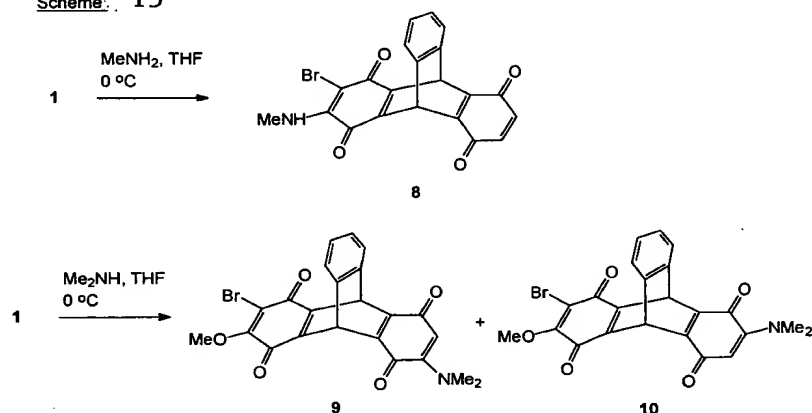
Scheme 14



To synthesize new analogs, 1 was treated with primary and secondary amines (Scheme 15). Surprisingly, compound 1 reacts with primary amines such as methylamine and secondary amines such as dimethylamine to give different regio-isomeric products. Hence, addition of bromide 1 with methylamine in THF at 0°C for 20 min. gave a 66% yield of displacement product 8. On the other hand, when 1 was treated with dimethylamine in THF at 0°C , a 96% yield of regioisomers 9 and 10 (a ratio of 1:1) was isolated. Compounds 9 and 10 were separated by silica gel column chromatography. The regiochemistry of 9 and 10 has not been determined. The presence of bromine atom in the products was indicated by their mass spectra in which the

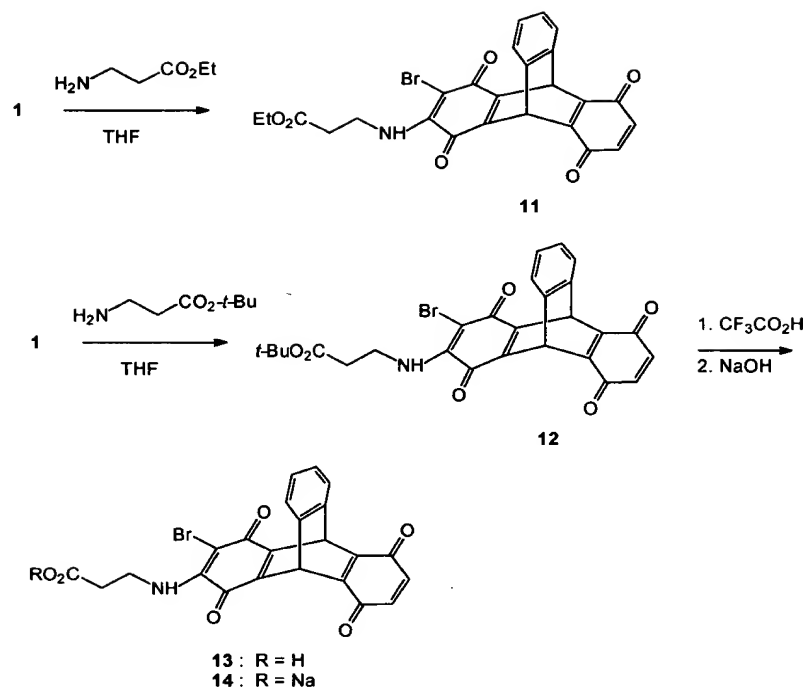
M+2 peaks (^{81}Br isotope) almost equal in intensity to the molecular (EI) or quasimolecular (CI) ions. No other by-products are identifiable in these reactions. These unusual addition reactions are unprecedented. It has been reported that p-benzoquinone undergoes addition reactions with aliphatic primary amines to give a mixture of products including mono- and di-adducts from the 1,4-addition reactions, and tetrachloro-1,4-benzoquinone and 2,3-dichloro-1,4-naphthoquinone react with secondary amines to give displacement products (addition to the double bond followed by elimination of chlorine). Contrary to these literature results, bromide 1 undergoes displacement of the methoxy group instead of bromine with primary amines and simple 1,4-addition on the unsubstituted quinone ring followed by oxidation with secondary amines, exclusively. Presumably, a less basic primary amine, methylamine, undergoes 1,4-addition reaction on the greater electron-deficient bromoquinone ring. Since the C2-bromine of 1 is an electron-withdrawing group, the anion resulted from the addition of an amine on C3 would be more stabilized than that from the addition on C2; resulting a displacement of methoxy group to provide compound 8. On the other hand, a more basic secondary amine, dimethylamine, is less affected by electronic (since it is a more reactive amine) effect of the quinone ring and is more affected by steric effect, in turn it prefers the addition on the unsubstituted (less hindered) quinone moiety to provide compounds 9 and 10.

Scheme 15



Due to the potent anticancer activity of amine 8 (vide infra) and a need of water soluble analogs, aliphatic primary amines containing ester function were used to synthesize various triptycene quinones, and their biological activities were studied (Scheme 16).

Scheme 1.6



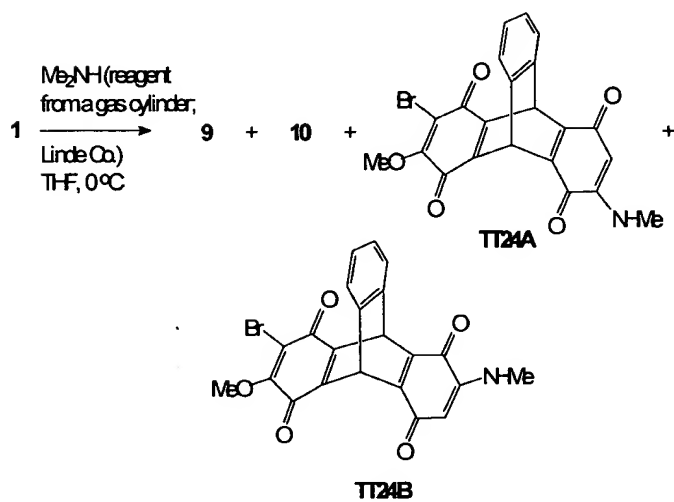
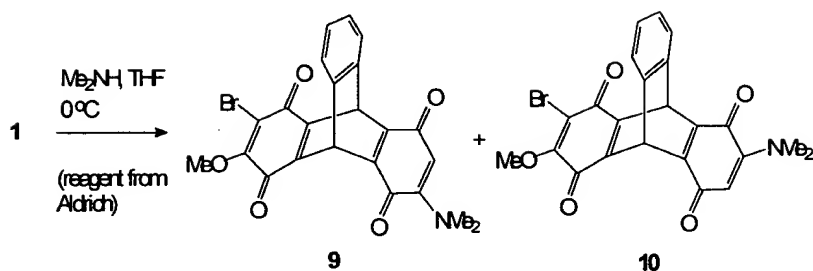
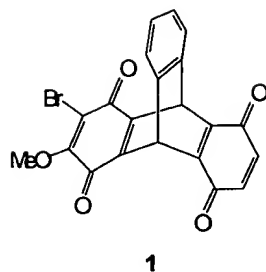
Treatment of bromide **1** with ethyl 2-aminopropanoate in THF at $-40\text{ }^{\circ}\text{C}$ for 6 h afforded a 46% yield of displacement product **11** along with starting material **1** (30% recovery). No other regioisomers were detected. Similarly, reaction of **1** with *t*-butyl 2-aminopropanoate at $-40\text{ }^{\circ}\text{C}$ for 8 h gave a 62% yield of **12**. Recrystallization of **12** in hexane:ethyl acetate (5:1) afforded single crystals which structure was unequivocally shown by X-ray analysis. Deprotection of the *t*-butyl ester of **12** with trifluoroacetic acid in dichloromethane gave a quantitative yield of acid **13**. Sodium salt **14**, a water soluble material, was obtained from the treatment of **13** with 1 equiv of sodium hydroxide.

When compound **1** was treated with dimethylamine in THF (1 M solution in THF; purchased from Aldrich Chemical Company), a mixture of **9** and **10** (1:1) was obtained (Scheme 18). However, when compound **1** was treated with dimethylamine in THF which prepared by adding dimethylamine (a gas purchased from Linde Company) gas into THF, a mixture of **9**, **10**, **TT24A**, and **TT24B** (4:4:1:1) was obtained. Compounds **9**, **10**, and **TT24** were separated by column chromatography. **TT24A** and **TT24B** were inseparable, however, carbon-13 NMR

spectrum indicates two isomers.

5 Likely, a small amount of methylamine is contaminated in the dimethylamine gas, this methylamine then added to the unsubstituted quinone ring of triptycene bisquinone to produce **TT24**. This mode of addition is different from the reaction of **1** with pure methylamine. It is suggested that dimethylamine acts as a base and remove the proton of methylamine to provide methylamine anion which is more reactive than methylamine itself, and in turn undergoes addition reaction on the unactivated C=C of the quinone moiety.

Scheme 18



Anticancer testing of **TT24** show it has an IC_{50} value of 48 nM (or 0.048 μ M). The compound is as active as daunomycin.

Anticancer and antimalarial activities of triptycene bisquinones have been evaluated. Table 2 summarizes IC_{50} values (the concentrations of drugs required to inhibit by 50% the viability of L1210 leukemic cells at day 4) of various triptycene bisquinones in the inhibition of L1210 leukemic cell growth. In general, IC_{50} values are in the 0.24 ~ 0.43 μ M range with the exception of carboxylic acid 13 and its sodium salt 14. Possibly, the passage ability of sodium salt 14 through cell membrane decreases as water solubility increases; in turn, ability in destroying cancer cells decreases. The IC_{50} value of daunomycin, a known anticancer drug, under similar treatment conditions is 0.041 μ M. Triptycene bisquinones induce DNA cleavage and inhibit nucleoside transport. In contrast, daunomycin, inhibits topoisomerases I and II, does not block nucleoside transport. Moreover, triptycene bisquinones 7, 8, 9, and ritonavir inhibit Plasmodium falciparum 3D7 (a malaria strain) with IC_{50} values of 8, 4.7, 5.6, and 9.7 μ M, respectively. Malaria protease plasmepsin II was also inhibited by compounds 8,9, and ritonavir and IC_{50} values of these compounds are 9.7, 23.6, and 0.10 μ M, respectively. Presumably, these reactive triptycene bisquinones undergo addition reactions with lysine, tryptophan, histidine, and cysteine residues of proteins to produce cross-linked proteins.

Table 2. Cytotoxicities of triptycene bisquinone analogs in L1210 leukemic cell system in vitro.

Compound	1	7	8	9	11	12	13	14
IC_{50} (μ M)	0.291	0.270	0.430	0.300	0.240	0.310	1.48	4.00

General Methods. Nuclear magnetic resonance spectra were obtained at 400 MHz for 1H and 100 MHz for ^{13}C in deuteriochloroform, unless otherwise indicated. Infrared spectra are reported in wavenumbers (cm^{-1}). Mass spectra were taken from a Hewlett Packard 5890A Series II, GC-MS. FAB spectra were taken by using Xe beam (8 KV) and m-nitrobenzyl alcohol as matrix. Methoxyhydroquinone (3), quinizarin, silver oxide, zinc iodide, sodium borohydride methylamine in THF, dimethylamine in THF, ethyl 3-aminopropanoate hydrochloric acid salt,

and t-butyl 3-aminopropanoate hydrochloric acid salt were purchased from Aldrich Chem. Co. Davisil silica gel, grade 643 (200~425 mesh), was used for the flash column chromatographic separation.

1,4-Dimethoxyanthracene (2).

To a cold (0 °C) methanol (20 mL) solution of 1.00 g (0.0042 mol) of quinizarin under argon was added 0.638 g (0.0168 mol) of sodium borohydride. The resulting mixture was stirred at 0 °C for 1 h. To it, 11 mL of 6 N HCl was added dropwise at 0 °C over a period of 10 min. The precipitated orange solids were collected, washed several times with distilled water, dried under vacuum, and recrystallized from acetone-ether to give 0.83 g (95% yield) of anthracene-1,4-dione¹⁷ as yellow crystals, mp 204 – 206 °C. ¹H NMR d 8.60 (s, 2 H, C 9,10 Hs), 8.10 (dd, J = 6.4, 3.2 Hz, 2 H, C 5,8 Hs), 7.70 (dd, J = 6.4, 3.2 Hz, 2 H, C 6,7 Hs), 7.10 (s, 2 H, C 2,3 Hs); ¹³C NMR d 184.7 (s, C=O), 140.1 (d), 134.8 (s), 130.2 (d), 129.6 (d), 128.9 (d), 128.4 (s).

To 2.00 g (1.00 mmol) of 1,4-anthracenedione, a solution of 6.68 g (38.0 mmol) of sodium hydrosulfite in 50 mL of water and 50 mL of 1,4-dioxane was added. The resulting mixture was stirred at 25 °C for 3 h, and added 100 mL of water. The mixture was cooled over an ice-water bath, and the precipitated dark green solids were collected by filtration, washed twice with water, and dried under vacuum to give 1.75 g (87% yield) of 1,4-dihydroxyanthracene, mp 167-169 °C; ¹H NMR d 8.70 (s, 2 H, C 9,10 Hs), 8.05 (m, 2 H, C 5,8 Hs), 7.50 (s, 2 H, C 6,7 Hs), 6.60 (s, 2 H, C 2,3 Hs).

To a 0.275 g (11.0 mmol) of pre-washed (with distilled diethyl ether) sodium hydride under argon were added 1.00 g (4.80 mmol) of 1,4-dihydroxyanthracene, 0.75 mL (12.0 mmol) of iodomethane, and 10 mL of DMF (distilled over CaH₂). The solution was stirred at 25 °C for 1.5 h, diluted with 20 mL of water, and acidified with 6 N HCl (pH ~2). The mixture was extracted three times with ethyl acetate, and the combined extract was washed twice with water, and brine, dried (MgSO₄), concentrated to give 1.07 g (94% yield) of compound 2. Recrystallization from ether:hexane (1:1) gave 0.89 g (78% yield) of yellow solids: mp 132-133 °C (Lit.⁶ 134-136 °C); MS, FAB, m/z 239 (M+1), 238 (M+); ¹H NMR d 8.70 (s, 2 H, C9,10H), 7.97 (dd, J = 6.1, 3.6 Hz, 2 H, C5,8H), 7.40 (dd, J = 6.6, 3.2 Hz, 2 H, C6,7H), 6.55 (s, 2 H, C2,3H), 3.97 (s, 6 H, OCH₃); ¹³C NMR d 149.5 (s, C8a,10a), 131.5 (s, C4a,9a), 128.5 (d, C9,10), 125.5 (d, C5,8), 120.7 (d, C6,7), 100.9 (d, C2,3), 55.6 (s, OCH₃).

2,5,8-Trimethoxy-9,10-dihydro-9,10-[1,2]benzenoanthracene-1,4-dione (4).

A mixture of 2.00 g (8.40 mmol) of 1,4-dimethoxyanthracene (2), 1.411 g (10.1 mmol) of methoxyhydroquinone (3), 3.90 g (16.8 mmol) of silver oxide, and 0.536 g (1.68 mmol) of zinc iodide in 30 mL of toluene under argon was refluxed for 3 days. To the mixture, 0.25 g (1.79 mmol) of 3 and 1.30 g (5.60 mmol) of silver oxide were added and the reaction mixture was refluxed for another day. The reaction mixture was cooled, diluted with 200 mL of dichloromethane, filtered through Celite, and the filtrate was washed with aqueous NH_4Cl , and brine, dried (MgSO_4), concentrated, and column chromatographed on silica gel using a gradient mixture of hexane and ethyl acetate as solvent gave 2.21 g (70% yield) of 4: mp. 108~110 °C; ^1H NMR d 7.43 (dd, $J = 5.5$, 3 Hz, 2 H), 7.00 (dd, $J = 5.5$, 3 Hz, 2 H), 6.52 (s, 2 H, C6,7 Hs), 6.25 (s, 1 H), 6.23 (s, 1 H), 5.71 (s, 1 H, C3 H), 3.79 (s, 6 H, OMe), 3.72 (s, 3 H, OMe). ^{13}C NMR d 183.5 (C=O), 178.2 (C=O), 158.4, 153.8, 150.9, 150.5, 149.6, 149.5, 144.3, 144.1, 133.6, 133.5, 125.2 (2C), 124.5, 124.4, 109.5, 105.6, 56.4, 56.38, 56.3, 41.5, 41.2. Anal. Calcd for $\text{C}_{23}\text{H}_{18}\text{O}_5$: C, 73.79; H, 4.85. Found: C, 73.51; H, 5.07.

2,5,8-Trimethoxy-4a,9,9a,10-tetrahydro-9,10-[1,2]benzenoanthracene-1,4-dione (5)

A mixture of 1.00 g (7.10 mmol) of methoxyhydroquinone (3), 2.50 g (10.7 mmol) of silver oxide, and 1.20 g (8.50 mmol) of K_2CO_3 in 50 mL of benzene was stirred under argon at 25 °C for 3 h, the mixture was filtered through Celite, and washed with 5 mL of dichloromethane. The filtrate was concentrated to give 0.970 g (99% yield) of methoxybenzoquinone (6): ^1H NMR d 6.72 (s, 2 H), 5.95 (s, 1 H), 3.84 (s, 3 H). ^{13}C NMR d 187.4, 181.6, 137.1, 134.4, 111.5, 107.6, 56.2. This material was used in the next step without purification.

A solution of 0.70 g (2.90 mmol) of 2 and 1.00 g (7.20 mmol) of 6 in 10 mL of toluene in a sealed tube was heated at 150 °C for 1 day, cooled to 25 °C, concentrated to dryness, and column chromatographed on silica gel using a gradient mixture of hexane and ethyl acetate as solvent gave 0.52 g (48% yield) of compounds 5 as a mixture of exo- and endo- (1:1) isomers along with 0.62 g of 6. The exo- and endo-isomers were partially separated by silica gel column and the stereochemistry have not assigned. Compound 5, less polar isomer: MS, m/z 376 (M^+), 375. ^1H NMR d 7.24 - 7.18 (m, 2 H), 7.08 - 7.04 (m, 2 H), 6.65 (s, 2 H, C6,7 Hs), 5.61 (s, 1 H),

5.33 (bs, 2 H), 3.82 (s, 6 H, OMe), 3.48 (s, 3 H, OMe), 3.12 (ABd, J = 9 Hz, 1 H), 3.06 (ABd, J = 9 Hz, 1 H). More polar isomer: MS, m/z 376 (M⁺), 375. ¹H NMR d 7.40 (m, 2 H), 7.16 (m, 2 H), 6.60 (ABd, J = 8.8 Hz, 1 H), 6.56 (Abd, J = 8.8 Hz, 1 H), 5.65 (s, 1 H, =CH), 5.33 (bs, 2 H, C9,10 Hs), 3.76 (s, 3 H, OMe), 3.73 (s, 3 H, OMe), 3.51 (s, 3 H, OMe), 3.13 (Abd, J = 9 Hz, 1 H), 3.08 (Abd, J = 9 Hz, 1 H). This mixture of compounds was used in the next step.

1,4-Dihydroxy-2,5,8-trimethoxy-9,10-dihydro-9,10-[1,2]benzenoanthracene.

To a solution of 0.77 g (2.0 mmol) of 5 in 30 mL of 1,4-dioxane and 30 mL of water was added 1.12 g (20 mmol) of KOH. The solution was stirred at 25 °C for 1 h, acidified with 1 N HCl, and extracted three times with dichloromethane. The combined extract was washed with brine, dried (MgSO₄), and concentrated to give 0.77 g (100% yield) of 1,4-dihydroxy-2,5,8-trimethoxy-9,10-dihydro-9,10-[1,2]benzenoanthracene: MS, m/z 376 (M⁺). ¹H NMR d 7.43 (dd, J = 5.5, 3 Hz, 1 H), 7.38 (dd, J = 5.5, 3 Hz, 1 H), 6.96 (dd, J = 5.5, 3 Hz, 2 H), 6.51 (s, 2 H, C6,7 Hs), 6.29 (s, 1 H, C3 H), 6.09 (s, 1 H), 5.98 (s, 1 H), 5.3 (s, 1 H, OH), 4.96 (bs, 1 H, OH), 3.81 (s, 3 H, OMe), 3.805 (s, 3 H, OMe), 3.66 (s, 3 H, OMe). This compound was used in the next step.

Oxidation of 1,4-Dihydroxy-2,5,8-trimethoxy-9,10-dihydro-9,10-[1,2]benzenoanthracene to 4.

To a mixture of 0.77 g (2.0 mmol) of 1,4-dihydroxy-2,5,8-trimethoxy-9,10-dihydro-9,10-[1,2]benzenoanthracene and 0.60 g (4.2 mmol) of sodium sulfate (anhydrous) in 15 mL of dried acetone under argon at 25 °C was added 0.56 g (4.0 mmol) of silver oxide. The mixture was heated under reflux for 6 h, cooled to 25 °C, diluted with dichloromethane, and filtered through Celite. The filtrate was concentrated to give 0.77 g of the crude product. Column chromatography on silica gel using a gradient mixture of hexane and ethyl acetate as solvent gave 0.73 g (95% yield) of 4.

2-Methoxy-9,10-dihydro-9,10-[1,2]benzenoanthracene-1,4,5,8-tetraone (7).

To a solution of 0.77 g (2.0 mmol) of 4 in 16 mL of acetonitrile, 26 mL of 1,4-dioxane and 10 mL of water was added 2.0 g (3.6 mmol) of ceric ammonium nitrate at 25 °C. The solution was stirred for 12 h, diluted with dichloromethane, and washed with water. The organic layer was dried (MgSO₄), concentrated to give the crude product. Recrystallization from ether

gave 0.65 g (95% yield) of 7: MS, CI, m/z 345 (M+1), 317 (-CO); ¹H NMR d 7.48 (dd, J = 5.5, 3 Hz, 2 H), 7.07 (dd, J = 5.5, 3 Hz, 2 H), 6.65 (s, 2 H, C6,7 Hs), 6.2 (s, 1 H), 6.18 (s, 1 H), 5.78 (s, 1 H, C3 H), 3.78 (s, 3 H, OMe). ¹³C NMR d 182.2, 182.17, 182.0, 176.9, 158.5, 152.5, 151.6, 151.5, 149.6, 142.2, 142.0, 135.4, 135.3, 125.8 (2C), 125.4, 125.3, 105.7, 56.6 (OMe), 42.2, 41.9. Anal. Calcd for C₂₁H₁₂O₅: C, 73.25; H, 3.51. Found: C, 73.01; H, 3.80.

2-Bromo-3-methoxy-9,10-dihydro-9,10-[1,2]benzenoanthracene-1,4,5,8-tetraone (1).

To a solution of 0.30 g (0.87 mmol) of 7 in 20 mL of DMF under argon at 25 °C was added 0.16 g (0.87 mmol) of N-bromosuccinimide. After the solution was stirred for 10 h, it was diluted with water, and extracted twice with ethyl acetate. The combined extract was washed with brine, dried (MgSO₄), concentrated, and column chromatographed on silica gel using a gradient mixture of hexane and ether as solvent gave 0.29 g (45% yield) of 1: mp 207 – 210 °C; MS EI, m/z 424 & 422 (1:1, M+), 344 (M-Br), 300, 287, 232, 152, 126; ¹H NMR d 7.48 (dd, J = 5.5, 3 Hz, 2 H), 7.09 (dd, J = 5.5, 3 Hz, 2 H), 6.66 (s, 2 H, C6,7 Hs), 6.23 (s, 1 H), 6.16 (s, 1 H), 4.17 (s, 3 H, OMe); ¹³C NMR d 182.2 (s, CO), 182.0 (s, CO), 176.2 (s, CO), 175.5 (s, CO), 156.2, 151.6, 151.4, 150.2, 141.8, 135.5, 135.4, 126.2, 126.1, 126.07, 126.0, 125.6, 125.4, 117.2, 61.7 (OMe), 43.0, 42.1. Anal. Calcd for C₂₁H₁₁BrO₅: C, 59.60; H, 2.62. Found: C, 59.33; H, 2.87.

Synthesis of 1 from bromination of 4 followed by oxidation.

To a solution of 90 mg (0.24 mmol) of 4 in 5 mL of DMF under argon at 25 °C was added 52 mg (0.30 mmol) of N-bromosuccinimide (NBS). The solution was stirred at 40 °C for 12 h, diluted with diethyl ether, washed twice with water, and once with brine, dried (MgSO₄), and concentrated to give 109 mg (quantitative yield) of 2-bromo-3,5,8-trimethoxy-9,10-dihydro-9,10-[1,2]benzenoanthracene-1,4-dione: ¹H NMR d 7.44 (dd, J = 5.5, 3 Hz, 2 H), 7.01 (dd, J = 5.5, 3 Hz, 2 H), 6.54 (s, 2 H, C6,7 Hs), 6.29 (s, 1 H), 6.22 (s, 1 H), 4.14 (s, 3 H, OMe), 3.81 (s, 3 H, OMe), 3.79 (s, 3 H, OMe). This material was used in next step without purification.

To a solution of 0.109 g (0.240 mmol) of 2-bromo-3,5,8-trimethoxy-9,10-dihydro-9,10-[1,2]benzenoanthracene-1,4-dione in 5 mL of acetonitrile, 6 mL of 1,4-dioxane, and 2 mL of water, was added 0.660 g (1.20 mmol) of ceric ammonium nitrate. The solution was stirred at 25 °C for 6 h, diluted with ether, washed with water, and brine, dried (MgSO₄), concentrated, and

column chromatographed on silica gel using a gradient mixture of hexane and ether as eluant to give 60 mg (59% yield) of 1.

2-Bromo-3-(methylamino)-9,10-dihydro-9,10-[1,2]benzenoanthracene-1,4,5,8-tetraone (8).

A solution of 0.200 g (0.473 mmol) of 1 and 0.47 mL (0.946 mmol) of methylamine (2.0 M in THF) in 1 mL of THF was stirred under argon at 0 °C for 1 h. The solution was concentrated to dryness, and column chromatographed on silica gel using a gradient mixture of hexane and ethyl acetate as solvent to give 0.132 g (66% yield) of 8: mp. °C; MS, CI m/z 424, 422 (~1:1; M+1); ¹H NMR d 7.48 (dd, J = 5, 3 Hz, 1 H), 7.45 (dd, J = 5, 3 Hz, 1 H), 7.07 (dd, J = 5, 3 Hz, 2 H), 6.64 (s, 2 H, C6,7 Hs), 6.28 (s, 1 H), 6.10 (s, 1 H), 5.88 (bs, 1 H, NH), 3.33 (d, J = 5.6 Hz, 3 H, MeN); ¹³C NMR (acetone-d₆) d 183.33, 183.3, 178.2, 170.9, 154.4, 152.3, 152.0, 148.0, 147.0, 143.7, 143.5, 136.4, 136.3, 126.5 (2C), 126.0, 125.7, 125.0, 44.3, 42.9, 33.2. Anal. Calcd for C₂₁H₁₂BrNO₄: C, 59.74; H, 2.86. Found: C, 59.74; H, 2.76.

2-Bromo-3-methoxy-6-(dimethylamino)-9,10-dihydro-9,10-[1,2]benzenoanthracene-1,4,5,8-tetraone (9) and 2-Bromo-3-methoxy-7-(dimethylamino)-9,10-dihydro-9,10-[1,2]benzenoanthracene-1,4,5,8-tetraone (10).

To a solution of 0.200 g (0.473 mmol) of 1 in 2 mL of THF under argon at 0 °C, was added 0.24 mL (0.473 mmol) of dimethylamine (2.0 M in THF). After stirring at 0 °C for 2 h, the reaction solution was concentrated to dryness, and column chromatographed on silica gel using a mixture of benzene and ethyl acetate (10:1) as solvent to give 0.099 g (48% yield) of 9 (less polar isomer; the regiochemistry has not been determined) and 0.098 g (48% yield) of 10 (more polar).

Less polar isomer: MS, CI, m/z 468, 466 (~1:1; M+1); ¹H NMR d 7.47 ~ 7.43 (m, 2 H, Ar-H), 7.05 ~ 7.02 (m, 2 H, Ar-H), 6.22 (s, 1 H), 6.14 (s, 1 H), 5.38 (s, 1 H, C7H), 3.80 (s, 3 H, OMe), 3.11 (s, 6 H, Me₂N). ¹³C NMR d 181.4 (2C), 181.2 (2C), 153.7, 150.4, 149.3, 144.1, 143.9, 142.2, 141.8, 139.1, 127.1, 125.5, 125.3, 124.7, 124.2, 102.5, 61.1, 42.8, 42.7, 42.0, 41.4.

More polar isomer: MS, CI, m/z 468, 466 (~1:1; M+1); ¹H NMR d 7.48 ~ 7.42 (m, 2 H, Ar-H), 7.04 ~ 7.01 (m, 2 H, Ar-H), 6.24 (s, 1 H), 6.16 (s, 1 H), 5.39 (s, 1 H, C7H), 3.79 (s, 3 H, OMe), 3.10 (s, 6 H, Me₂N). ¹³C NMR d 181.6 (2C), 181.4 (2C), 154.0, 150.7, 149.6, 144.9, 144.0, 142.6, 141.5, 139.8, 127.2, 125.6 (2C), 125.0, 1

24.3, 102.4, 61.2, 43.0 (2C), 42.0, 41.9. HRMS m/z 466.0285 (466.0290, calcd for C₂₃ H₁₇ Br NO₅, M-H⁺).

2-Bromo-3-[2-(ethoxycarbonyl)ethylamino]-9,10-dihydro-9,10-[1,2]benzenoanthracene-1,4,5,8-tetraone (11).

To a mixture of 5.70 mg (0.236 mmol) of NaH (pre-washed with distilled ether) in 2 mL of DMF under argon at 0 °C was added 36.3 mg (0.236 mmol) of b-alanine ethyl ester hydrochloride (ethyl 3-aminopropanoate hydrochloride). The solution was stirred at 0 °C for 30 min, cooled to -46 °C, and a solution of 100 mg (0.236 mmol) of 1 in 1 mL of DMF was added via cannula. The solution was stirred at -46 °C for 6 h, diluted with a mixture of ethyl acetate and benzene (1:1), washed twice with water, brine, dried (Na₂SO₄), concentrated, and column chromatographed on silica gel using a gradient mixture of hexane and ethyl acetate as solvent to give 55 mg (46% yield) of 11 and 30 mg (30% recovery) of 1. Compound 11: mp. 119~121 °C; MS, m/z 510, 508 (~1:1; M+1), 422, 420 (1:1); ¹H NMR d 7.45 ~ 7.42 (m, 2 H, Ar-H), 7.08 ~ 7.04 (m, 2 H, Ar-H), 6.65 (s, 2 H, C6,7 Hs), 6.3 (bs, 1 H, NH), 6.27 (s, 1 H), 6.10 (s, 1 H), 4.16 (q, J = 7 Hz, 2 H, OCH₂), 4.03 (q, J = 6 Hz, 2 H, CH₂N), 2.64 (t, J = 6 Hz, 2 H, CH₂CO), 1.26 (t, J = 7 Hz, 3 H, CH₃); ¹³C NMR d 184.0, 183.6, 182.6, 182.3, 177.3, 171.7, 154.5, 151.8, 151.7, 147.4, 144.3, 142.4, 142.2, 135.8, 135.5, 126.2, 126.1, 125.9, 125.3, 61.3, 43.7, 42.2, 40.5, 35.4, 14.4. Anal. Calcd. For C₂₅H₁₈ Br NO₆: C, 59.07; H, 3.57. Found: C, 58.69; H, 3.72.

2-Bromo-3-[2-(t-butoxycarbonyl)ethylamino]-9,10-dihydro-9,10-[1,2]benzenoanthracene-1,4,5,8-tetraone (12).

To a mixture of 12.0 mg (0.496 mmol) of NaH (pre-washed with distilled ether) in 2 mL of DMF under argon at 0 °C was added 90.0 mg (0.496 mmol) of b-alanine t-butyl ester hydrochloride. The solution was stirred at 0 °C for 30 min, cooled to -46 °C, and a solution of 201 mg (0.496 mmol) of 1 in 1 mL of DMF was added via cannula. The solution was stirred at

–46 °C for 8 h, diluted with a mixture of ethyl acetate and benzene (1:1), washed twice with water, brine, dried (Na₂SO₄), concentrated, and column chromatographed on silica gel using a mixture of hexane and ethyl acetate (10:1) as solvent to give 165 mg (62% yield) of 12: mp. 171~173 °C; MS, m/z 538, 536 (~1:1; M+1), 482, 480 (1:1); ¹H NMR d 7.45 ~ 7.42 (m, 2 H, Ar-H), 7.08 ~ 7.04 (m, 2 H, Ar-H), 6.64 (s, 2 H, C6,7 Hs), 6.27 (s, 1 H), 6.22 (bs, 1 H, NH), 6.10 (s, 1 H), 4.00 (q, J = 6.7 Hz, 2 H, CH₂N), 2.55 (t, J = 6.7 Hz, 2 H, CH₂CO), 1.44 (s, 9 H, CH₃); ¹³C NMR d 182.3, 182.0, 177.0, 173.5, 170.6, 154.1, 151.4, 151.37, 147.0, 144.2, 142.1, 141.9, 135.4, 135.2, 128.8, 125.8, 125.76, 125.5, 125.0, 81.5, 43.5, 41.9, 40.5, 36.2, 28.0 (3C). Anal. Calcd for C₂₇H₂₂BrNO₆: C, 60.46; H, 4.13. Found: C, 60.75; H, 4.39.

N-(2-Bromo-9,10-dihydro-1,4,5,8-tetraoxo-9,10-[1,2]benzenoanthracene-3-yl)-3-aminopropanoic acid (13).

A solution of 0.100 g (0.187 mmol) of 12 and 0.2 mL (2.60 mmol) of trifluoroacetic acid in 5 mL of dichloromethane was stirred at 0 °C for 1 h and then at 25 °C for 2 h. The solution was concentrated to dryness, and recrystallized from benzene to give 0.090 g (100% yield) of purple solids: mp. 149 ~ 151 °C; MS, m/z 482, 480 (~1:1; M+1), 422, 420 (1:1); ¹H NMR d 7.49 ~ 7.46 (m, 2 H), 7.08 ~ 7.06 (m, 2 H), 6.64 (s, 2 H), 6.27 (s, 1 H), 6.22 (bs, 1 H, NH), 6.10 (s, 1 H), 4.04 (q, J = 6 Hz, 2 H, CH₂N), 3.40 (bs, 1 H, OH), 2.73 (t, J = 6 Hz, 2 H, CH₂); ¹³C NMR (CD₃OD) d 182.8, 182.77, 177.2, 174.2, 174.1, 151.6, 151.4, 147.6, 145.6, 142.9, 142.6, 137.9, 135.4, 129.0, 128.9, 128.2, 128.18, 126.1, 125.2, 43.8, 42.4, 42.3, 34.8. Anal. Calcd for C₂₃H₁₄BrNO₆: C, 57.52; H, 2.94. Found: C, 56.72; H, 2.88.

Preparation of TT24:

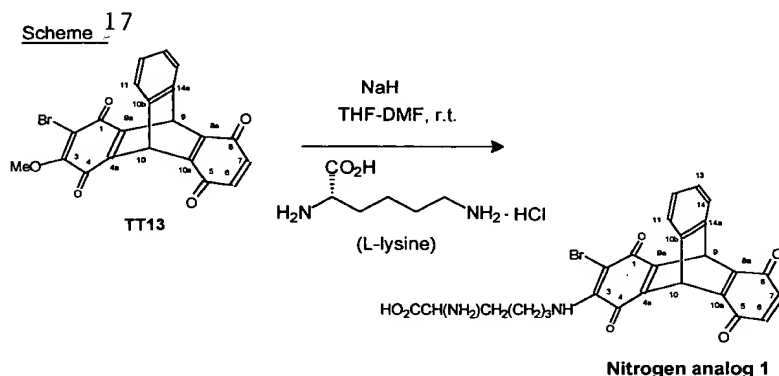
To 20 mL of THF solution, 1 g (0.022 mol) of dimethylamine gas (from a gas cylinder purchased from Linde Company) was added. A 1.1 M of dimethylamine in THF was resulted. To a cold (0°C) solution of 0.081 g (0.19 mmol) of triptycene bisquinone **1** in 2 mL of THF under argon was added 0.17 mL (0.19 mmol) of dimethylamine. The solution turn to purple immediately and was stirred at 0 °C for 10 min. The solvent was removed on a rotary evaporator and then under vacuum and the crude product was column chromatographed on silica gel using a gradient mixture of hexane and ethyl acetate as the eluent to give compounds **1**, **2**, and TT24.

Compound **TT24**: ^1H NMR (CDCl_3) δ 7.44 – 7.40 (m, 2 H), 7.03 (m, 2 H), 6.27 (s, 1 H), 6.15 and 6.14 (2 s, 1 H), 5.69 (q, J = 5.6 Hz, NH), 5.26 (s, 1 H), 3.80 & 3.79 (2 s, 3 H, OMe), 2.79 (d, J = 5.6 Hz, 3 H, NMe); ^{13}C NMR (CDCl_3) δ 181.7, 181.6, 179.5, 156.2, 147.8, 147.6, 143.9, 143.8, 143.6, 142.3, 141.8, 141.3, 139.6, 139.1, 131.2, 126.9, 125.52, 125.49, 125.46, 125.4, 124.8, 124.75, 124.3, 124.2, 102.3, 95.8, 95.7, 61.1, 60.4, 42.2, 41.9, 41.3, 41.0, 29.3 (2 C).

SYNTHESIS OF OTHER N ANALOGS AND S ANALOGS

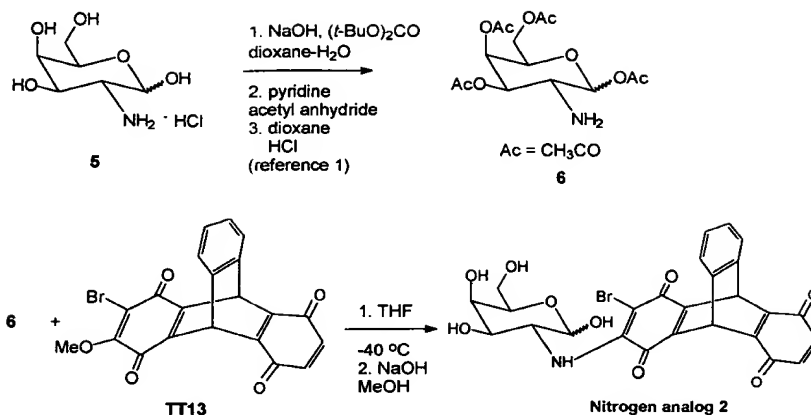
As illustrated in Scheme 17, nitrogen analog **1** has been prepared. Treatment of **TT13** with 1 equivalent each of L-lysine hydrochloric acid and sodium hydride in a 1:1 mixture of THF and DMF under argon at room temperature gave amino acid analog **1** which is a water soluble drug.

Scheme 19 outlines the synthetic route to prepare nitrogen analogs 2-4. Treatment (Kenani, A.; Bailly, C.; Helbecque, N.; Houssin, R.; Bernier, J. -L.; Henichart, J. -P. *Eur. J. Med. Chem.* **1989**, *24*, 371-377.) of D-galactosamine hydrochloride (commercially available) with sodium hydroxide and di-*t*-butylcarbonate in 1,4-dioxane and water, followed by protection of the hydroxyl function with excess of acetyl anhydride in pyridine, and removal of the Boc protecting group with hydrochloric acid in 1,4-dioxane produces amine **6**. Addition of **6** with **TT13** in THF at -40°C followed by removal of the acetoxy protecting group affords nitrogen analog **2**.



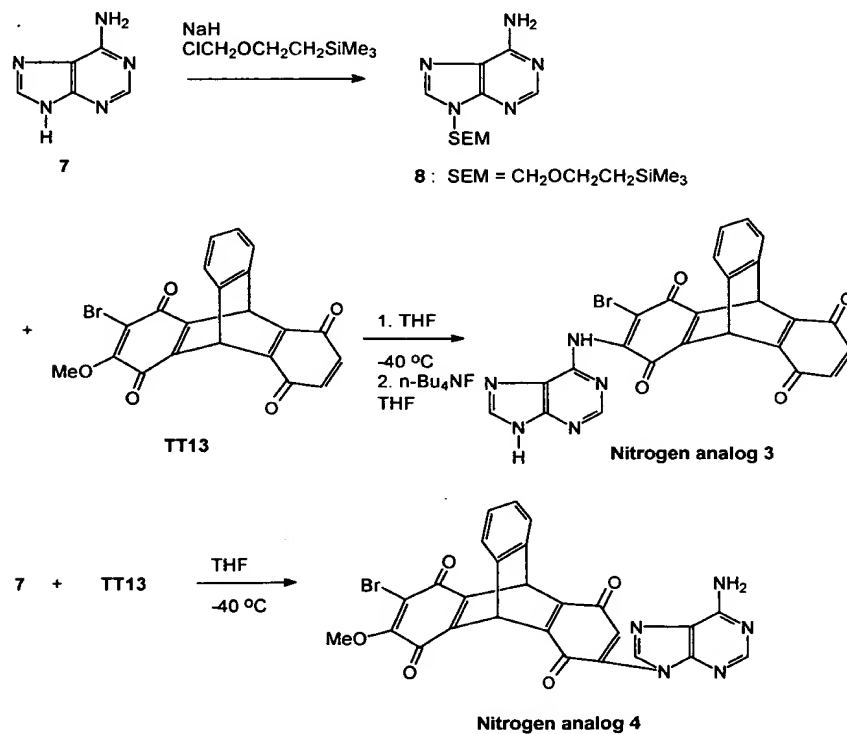
Similarly, glucosamine hydrochloride can be used to prepare glucosamine analog (of **2**; instead of galactosamine analog).

Scheme 19



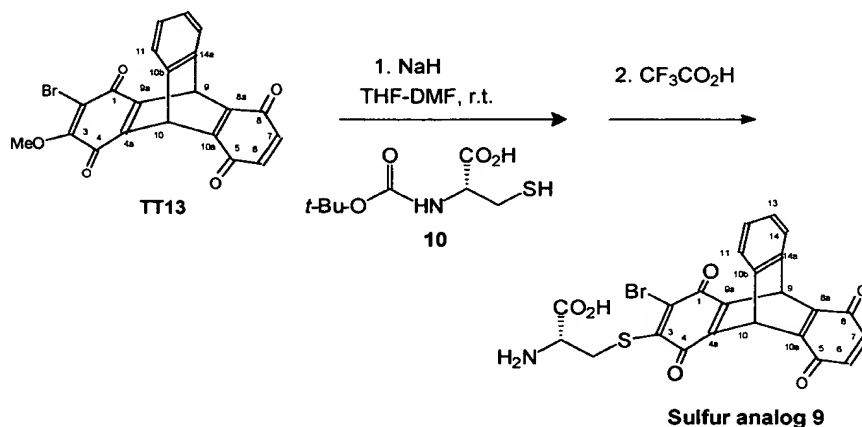
Syntheses of nitrogen analogs **3** and **4** are depicted in Scheme 20. Protection of N-9 of adenine (**7**) with (trimethylsilyl)ethyl chloromethyl ether (SEM-Cl) and sodium hydride in DMF produces compound **8**. Addition of **8** with TT13 in THF at -40 °C followed by removal of the SEM protecting group with tetra-*n*-butylammonium fluoride in THF furnishes nitrogen analog **3**. Direct treatment of adenine and catalytic amount of sodium hydride in THF at -40 °C gives nitrogen analog **4**. The N9 hydrogen of adenine (**7**) is more acidic than that of C6 amine group, hence, N9 nitrogen should react with TT13.

Scheme 20



Sulfur analogs: Uses of thiol containing amino acid such as N-protected L-cysteine provides sulfur analog (Scheme 21). Hence, addition of *N*-Boc cysteine and sodium hydride in DMF-THF followed by trifluoroacetic acid (removal of the Boc protecting group) will afford sulfur analog .

Scheme. 21



The above four nitrogen analogs and the sulfur analog are water soluble materials. Other nitrogen and sulfur analogs are prepared by methods known to one of ordinary skill in the art, using substitutions which are known to one of ordinary skill in the art.

Compounds of this invention other than those particularly shown may be prepared without undue experimentation by those skilled in the art of synthetic chemistry by methods analogous to those specifically disclosed herein or in publications and patent applications incorporated by reference or by methods known in the art.

Although the description above contains many specificities these should not be construed as limiting the scope of the invention, but merely providing illustrations of some of the presently preferred embodiments of the invention. Thus, the scope of the invention should be determined by the appended claims and their legal equivalents, rather than by the examples given. All references cited herein are incorporated by reference to the extent not inconsistent with the disclosure herein.

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